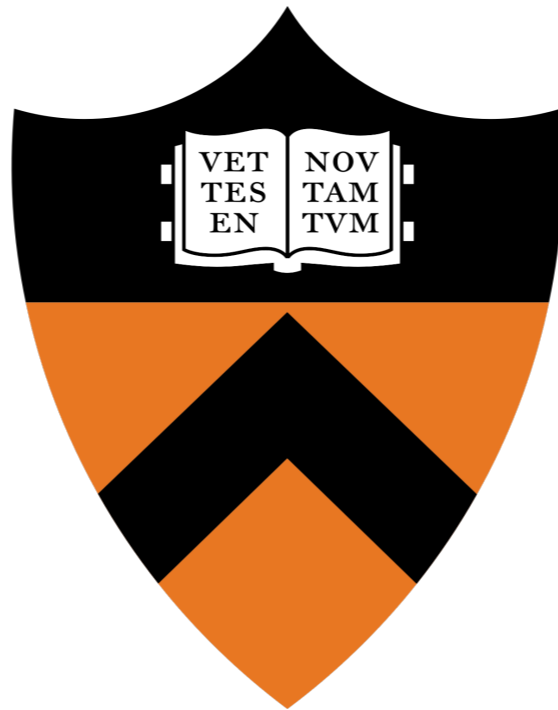


Catalysis by Small Peptides
an Introduction

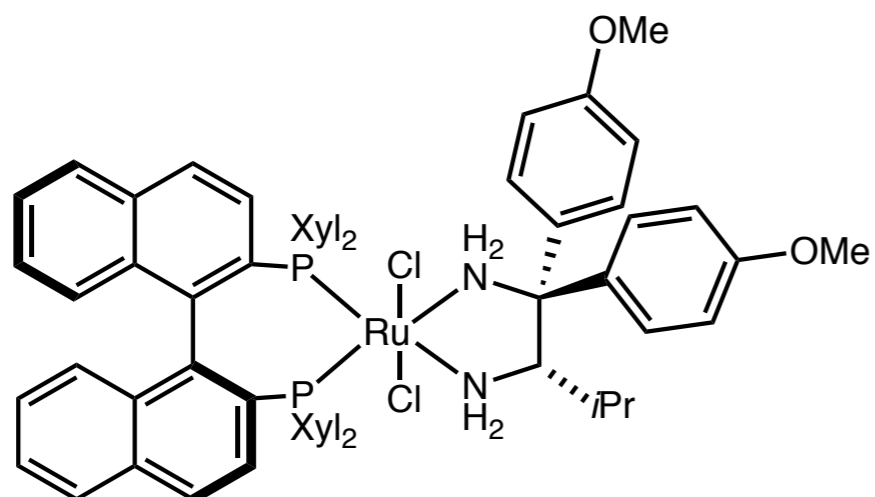


Mario Wiesenfeldt

November 4th, 2020

Catalysis by Small Peptides

Catalysis by small molecules

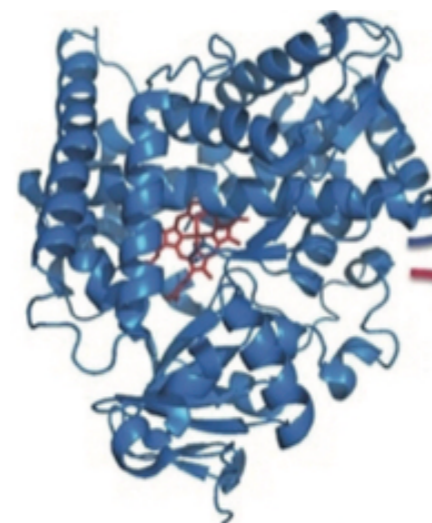


design &
understanding



**diverse
reactivity**

Enzyme catalysis

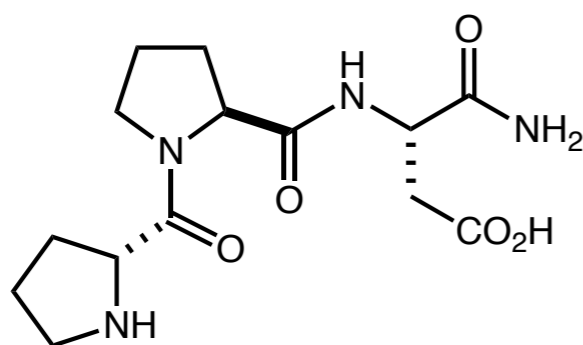


tailor-made non-
covalent interactions



**exceptional
selectivity**

Catalysis by small peptides



2–20 amino acids

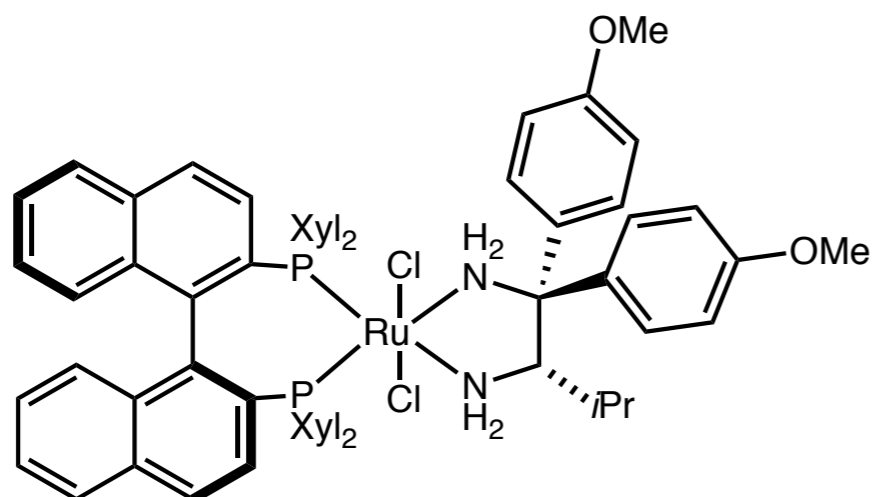
modular and facile synthesis

non-covalent interactions with substrates – **high selectivity**

nonnatural amino acids – **diverse reactivity**

Catalysis by Small Peptides

Catalysis by small molecules

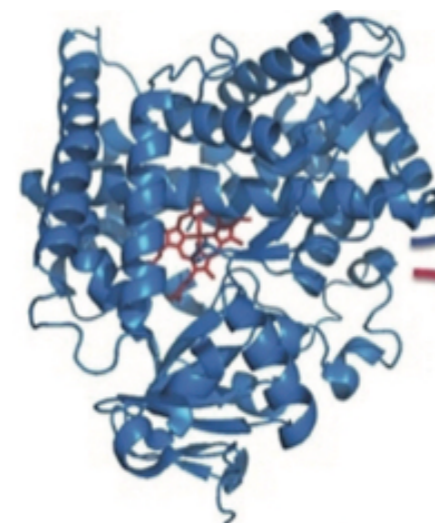


design &
understanding



enzyme-like
selectivity

Enzyme catalysis

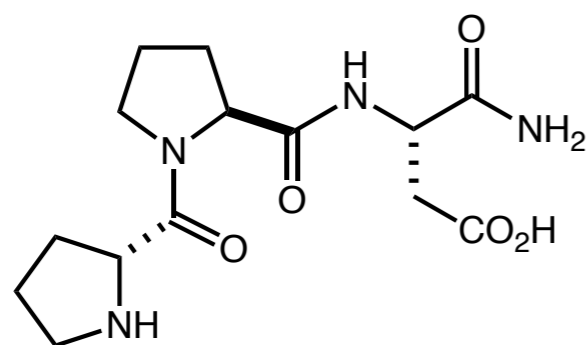


directed evolution or merger
with (photo-) catalysis



nonnatural
reactivity

Catalysis by small peptides



2–20 amino acids

modular and facile synthesis

non-covalent interactions with substrates – **high selectivity**

nonnatural amino acids – **diverse reactivity**

Nonnatural reactivity by photocatalysis: Biegasiewicz, K. F.; Cooper, S. J.; Gao, X.; Oblinsky, D.; Kim, J. H.; Garfinkle, S. E.; Joyce, L. A.; Sandoval, B.; Scholes, G. D.; Hyster, T. *Science* **2019**, *364*, 1166. Comparison between enzymes, transition metals, and small peptides: Lewis, C. J. *ACS Catal.* **2013**, *3*, 2954. (Selected) nobel lectures on asymmetric catalysis and directed evolution: Noyori, R. *Angew. Chem., Int. Ed.* **2019**, *58*, 14420 – Arnold, F. *Angew. Chem., Int. Ed.* **2002**, *41*, 2008.

Small Peptides as Organocatalysts

- Introduction
- Catalytically relevant properties of small peptides
- Small peptides as organocatalyst
- Small peptides in Lewis acid and transition metal catalysis
- Small peptides in photoredox catalysis
- Summary

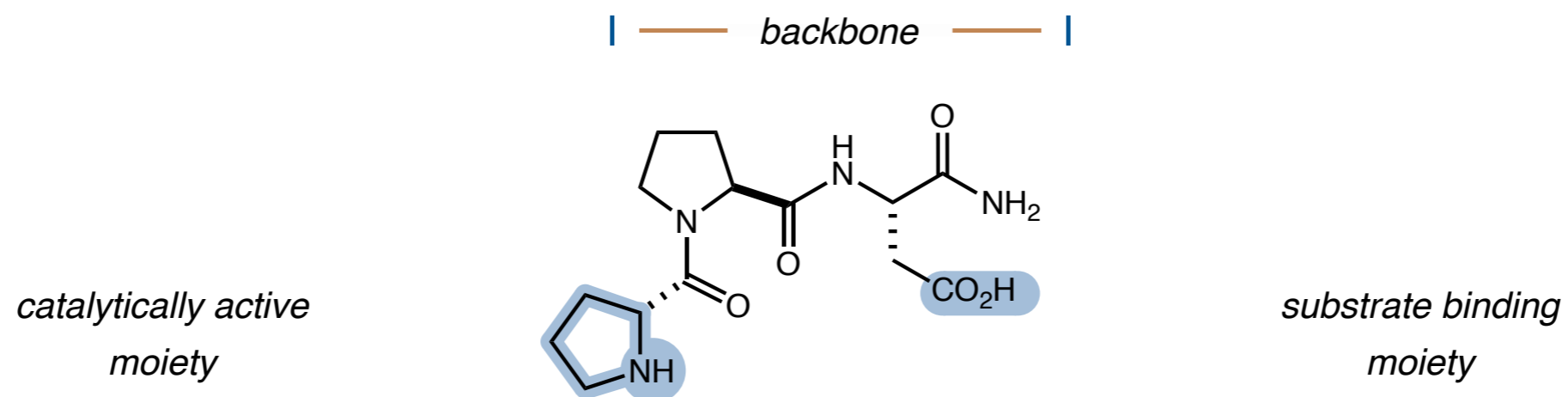
This group meeting does not cover the available literature comprehensively. Instead, selected key studies are discussed in order to explain the underlying concepts.

Small Peptides as Organocatalysts

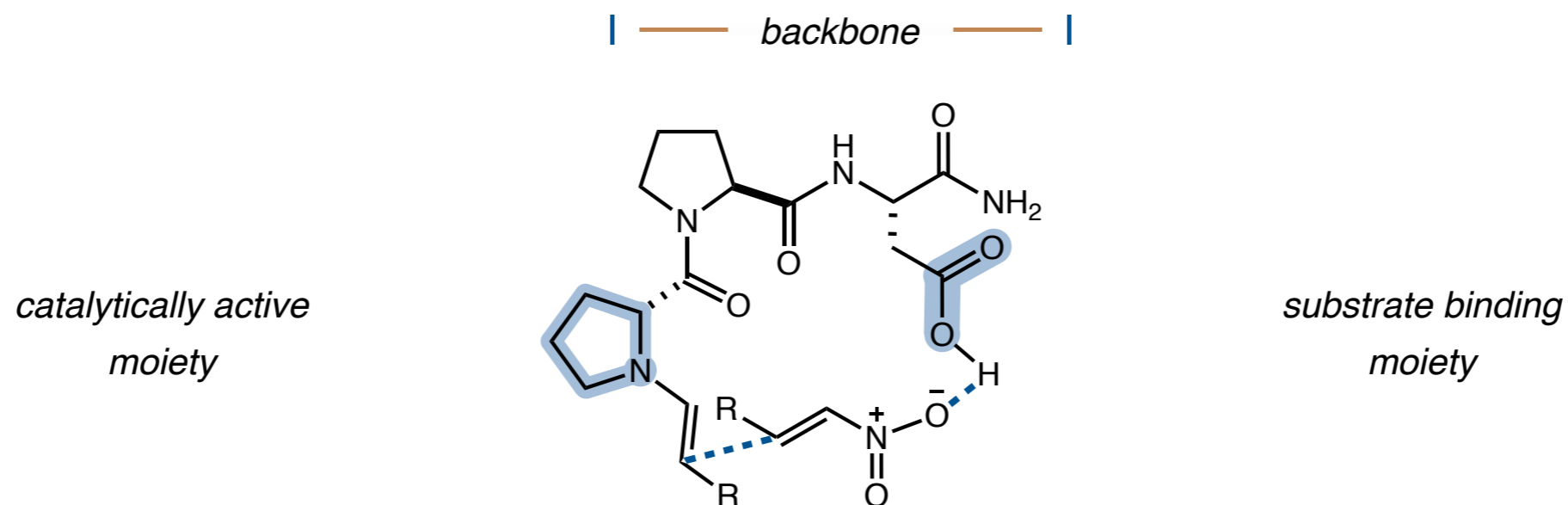
- Introduction
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Structure of a Peptide Catalyst



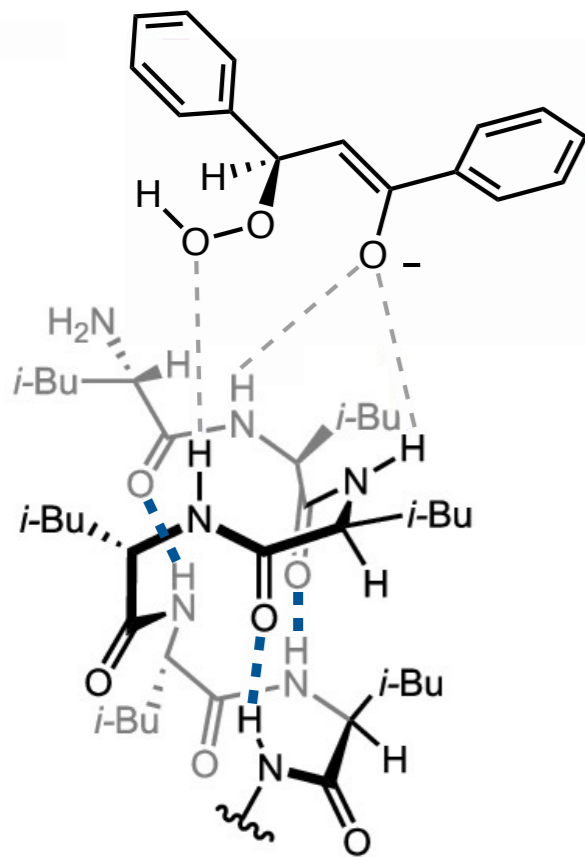
Structure of a Peptide Catalyst



- backbone serves as a tailor-made spacer between the *catalytically active moiety* and the *substrate binding moiety*
- complex three-dimensional geometry determined by primary and secondary protein structure
- conformational flexibility due to high number of rotatable bonds

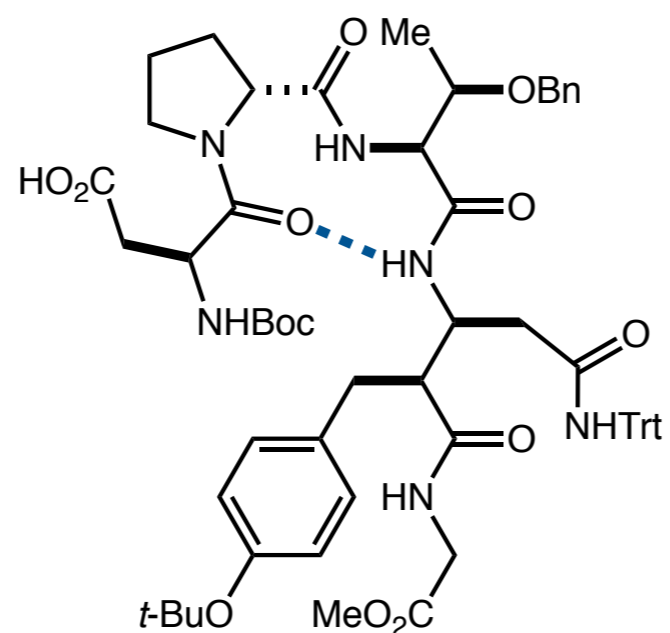
Design of the Secondary Structure

Natural secondary structure motifs



α -helix

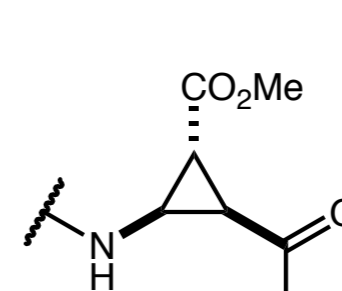
Juliá-Collona epoxidation



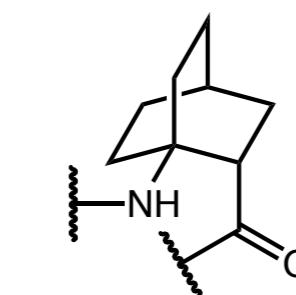
type II β -turn

epoxidation catalyst (Miller)

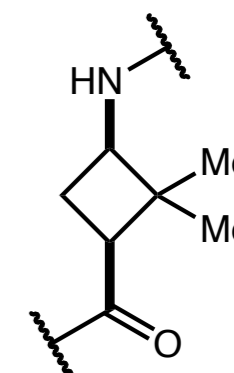
Rigid unnatural amino acids



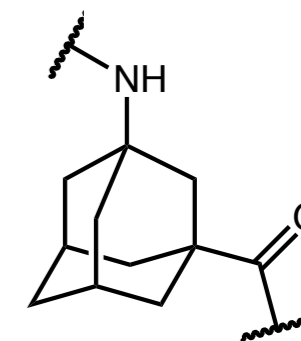
Reiser



Calmes, Amblard



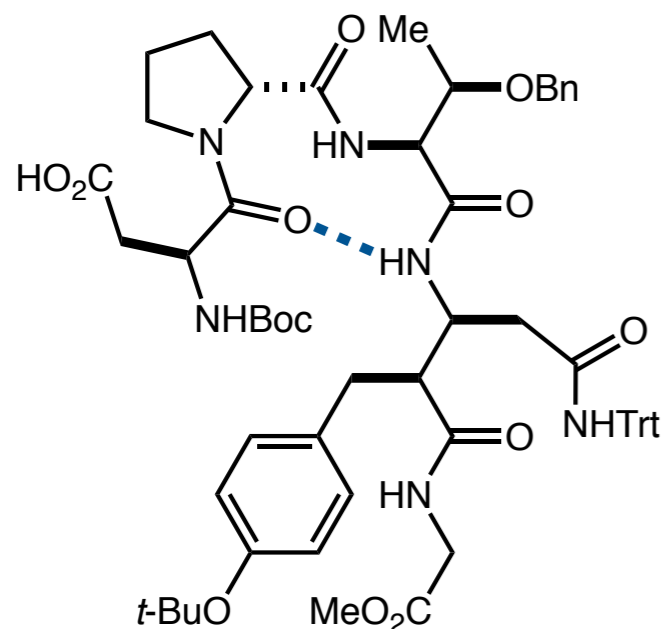
Bur



Schreiner

Peptide design is often centered around such structures with reduced conformational flexibility.

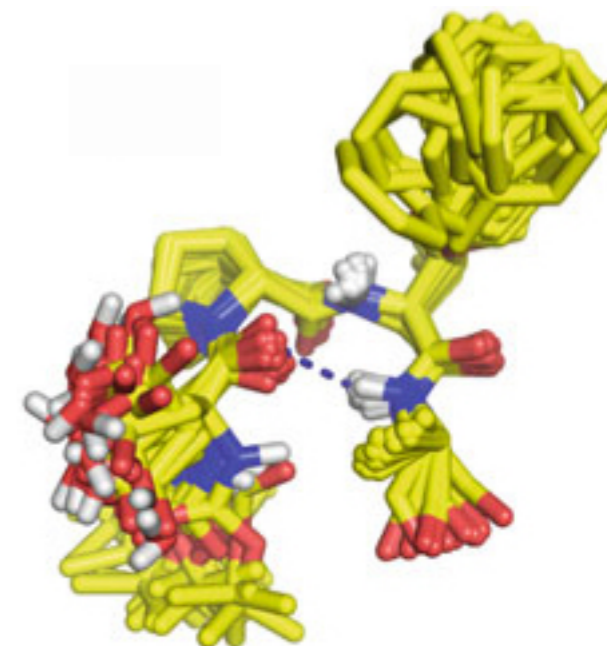
Design of the Secondary Structure



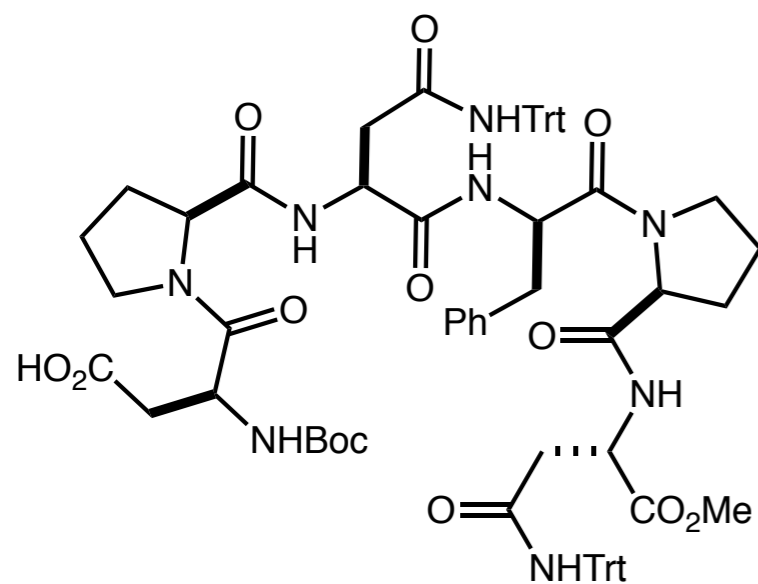
type II β -turn
epoxidation catalyst (Miller)

NMR &
calculations

highly regioselective in the
epoxidation of farnesol



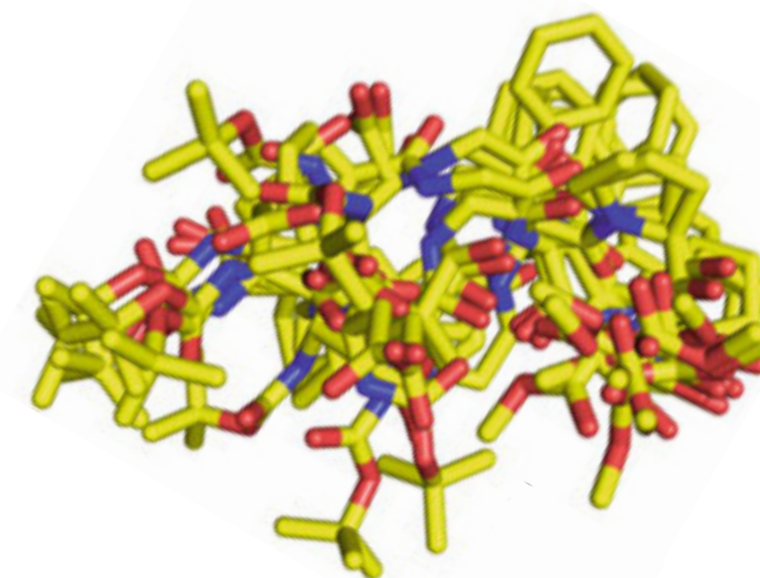
*ensemble of closely related
ground-state conformations*



epoxidation catalyst (Miller)

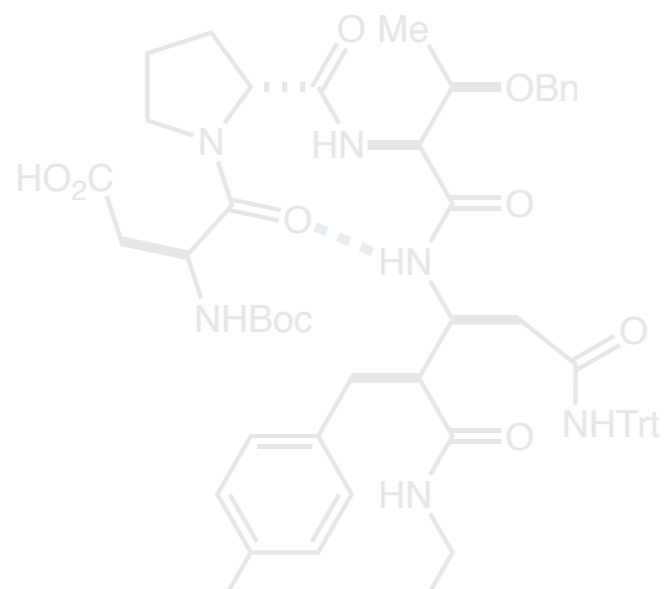
NMR &
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highly regio- and enantioselective
in the epoxidation of farnesol



heterogeneous ensemble

Design of the Secondary Structure



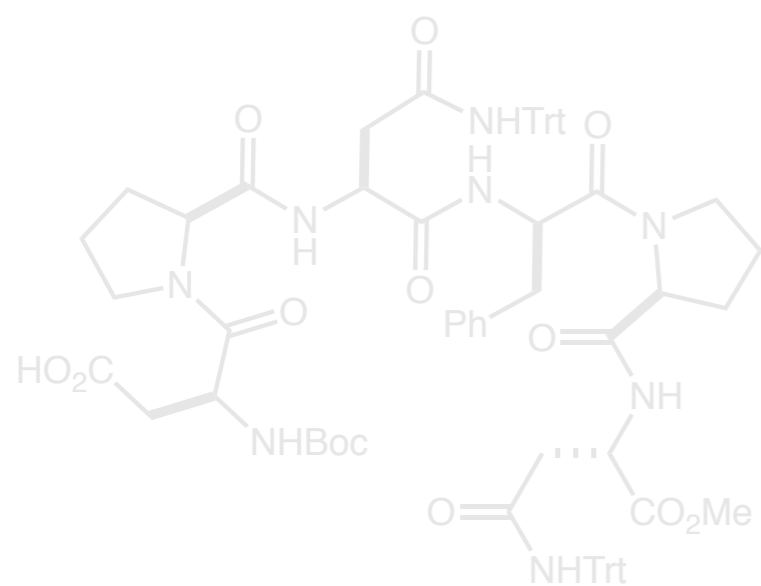
NMR &
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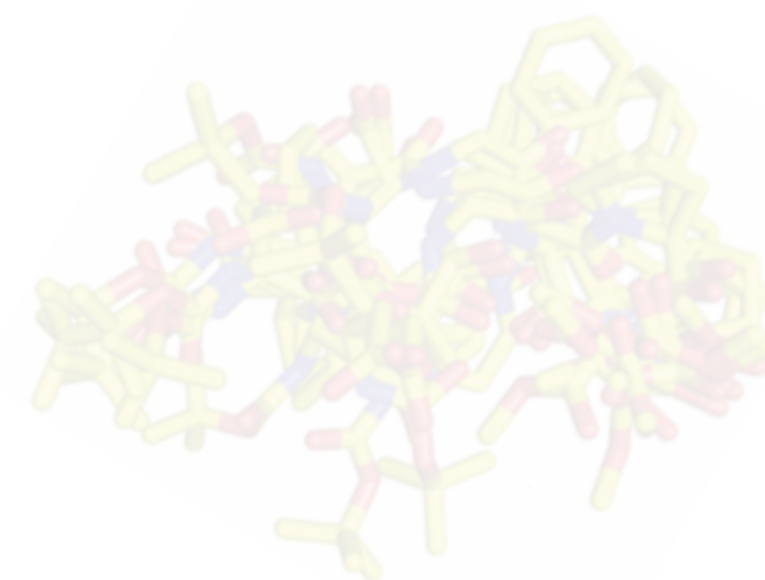
Even highly flexible, conformationally heterogeneous peptides may act as highly selective catalysts.

epoxidation catalyst (Miller)

ground-state conformations



NMR &
calculations

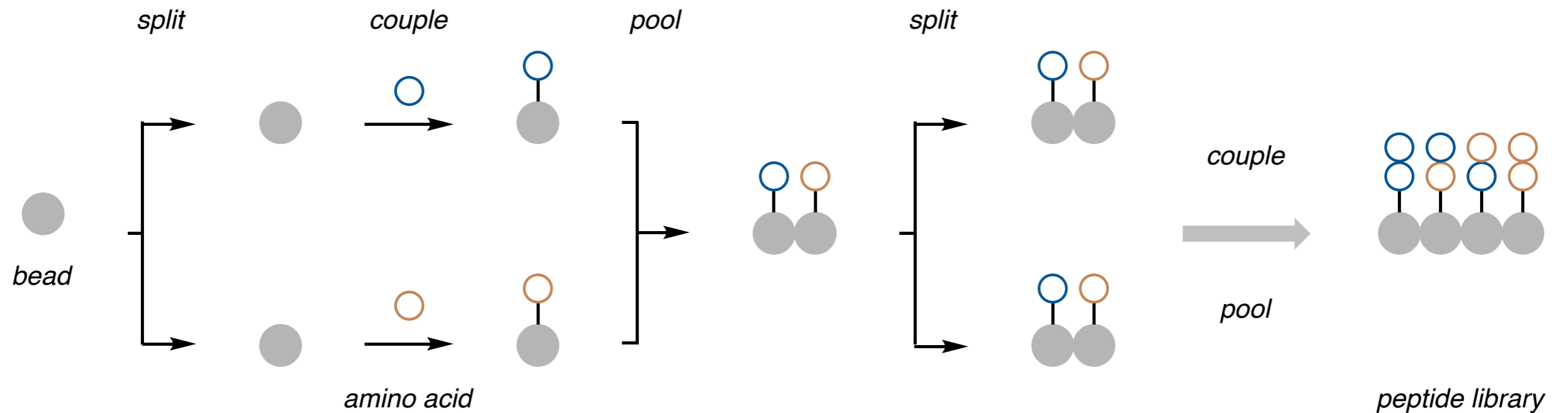


highly regio- and enantioselective
in the epoxidation of farnesol

heterogeneous ensemble

Strategies for Library Screening

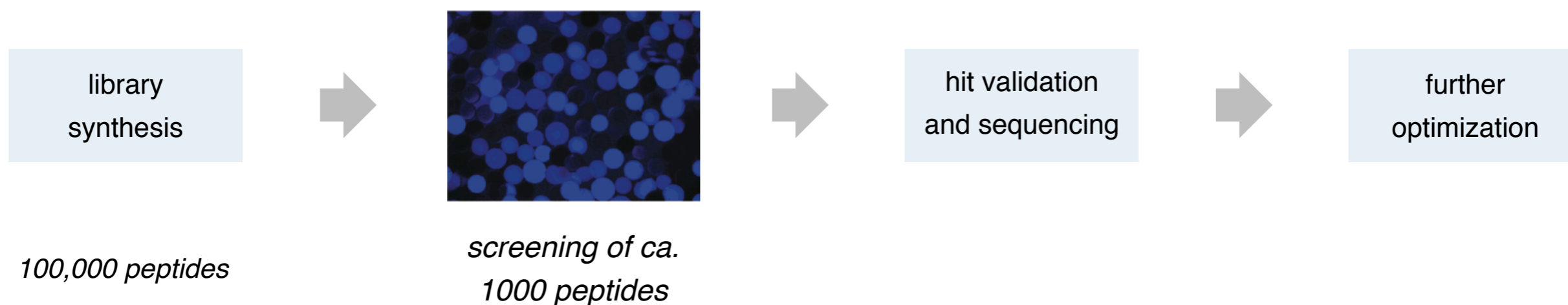
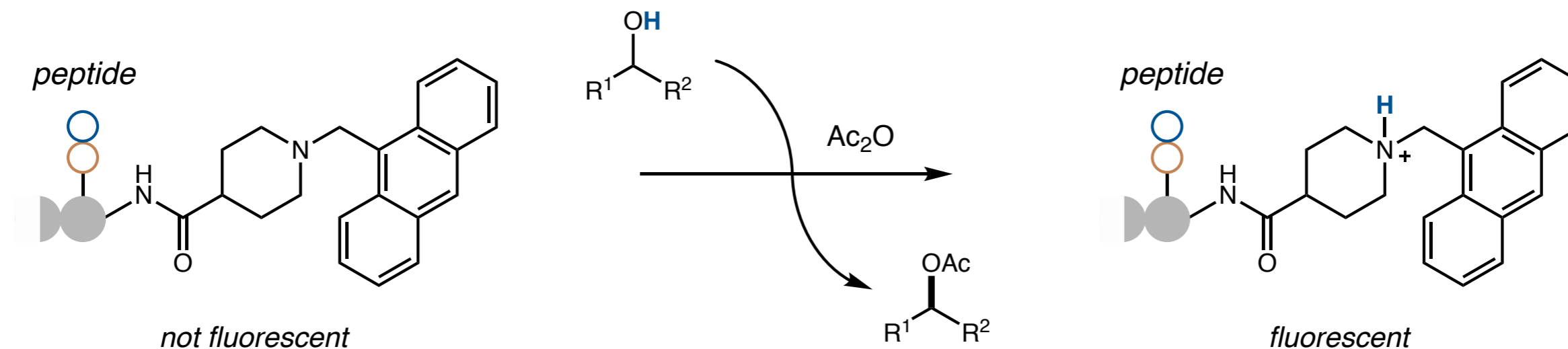
- Facile synthesis of large peptide libraries by split and pool synthesis



- ⦿ exponential increase of complexity with the number of cycles
- ⦿ biased libraries by control the introduction of specific amino acids in each cycle

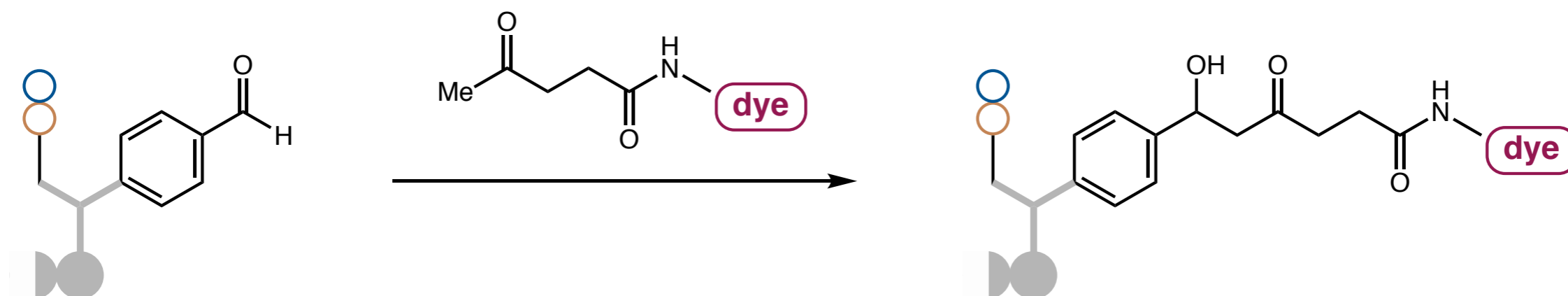
Strategies for Library Screening

- Visualization of hits by co-immobilized fluorescent tags



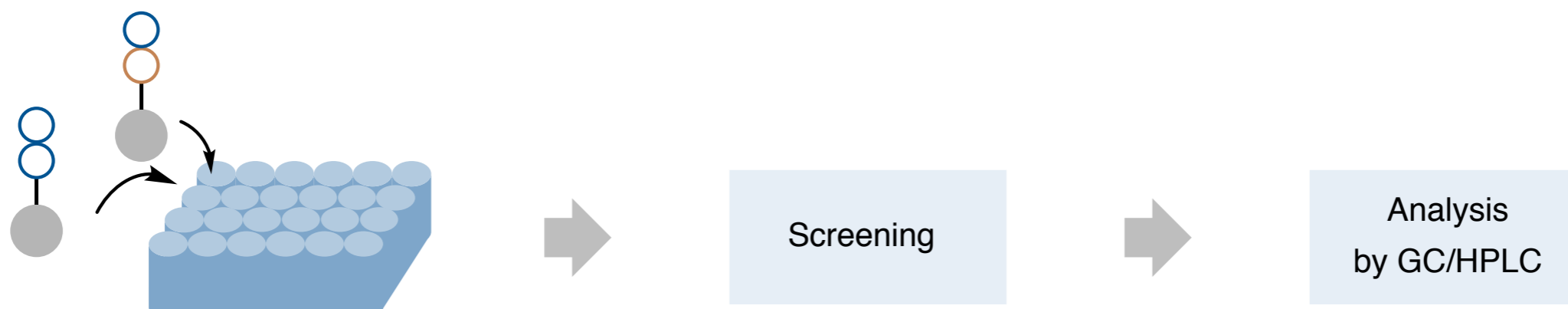
Strategies for Library Screening

■ Co-immobilization of substrates



Related work has been reported by Jacobsen, Bradley, Barbas, Davies, Berkessel, Snapper, Hoveyda, and others.

■ Sorting of individual beads into individual vials



Lichter, P.; Miller, S. J. *ACS Comb. Sci.* **2011**, *13*, 321.

Krattiger, P.; McCarthy, C.; Pfaltz, A.; Wennemers, H. *Angew. Chem., Int. Ed.* **2003**, *42*, 1722.

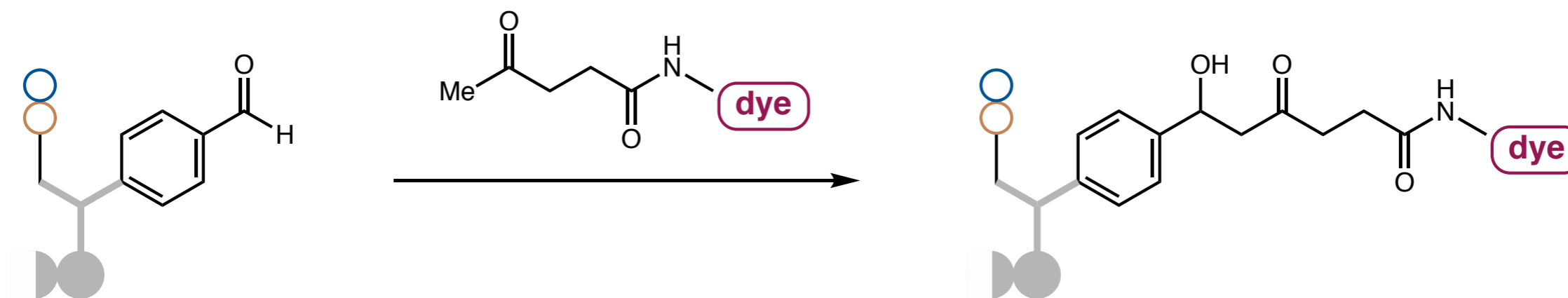
Small Peptides as Organocatalysts

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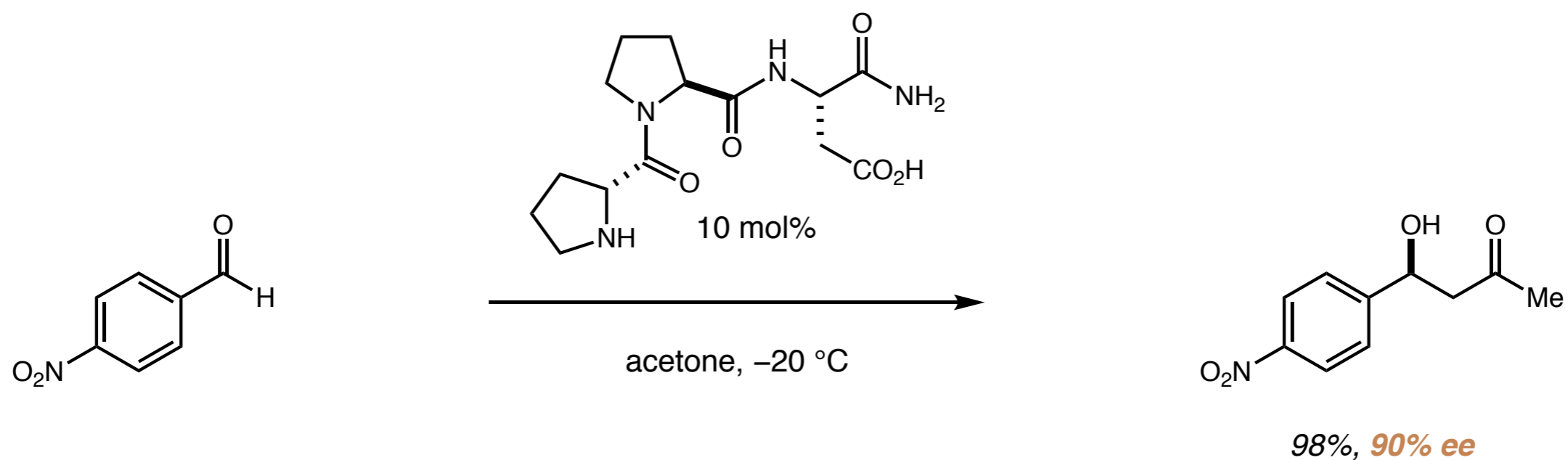
This group meeting does not cover the available literature comprehensively. Instead, selected key studies are discussed in order to explain the underlying concepts.

Wennemers' Tripeptide Catalyst

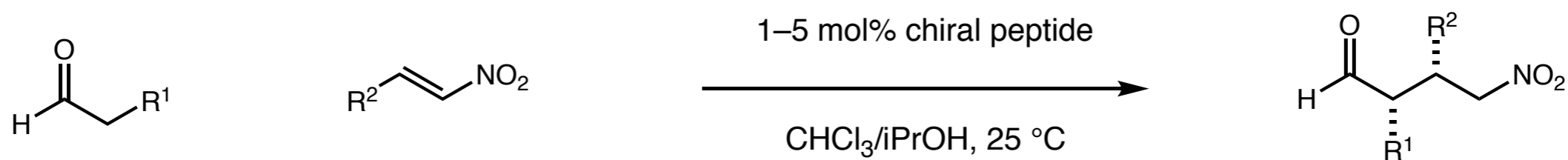
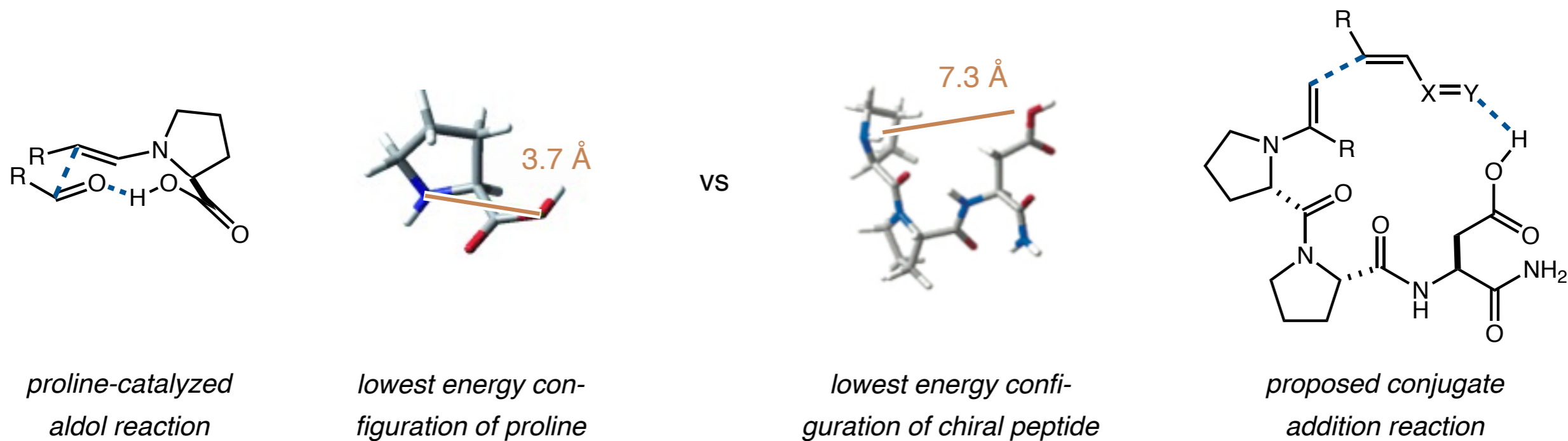
■ Hit identification by library screening



■ Optimization in solution



Wennemers' Tripeptide Catalyst

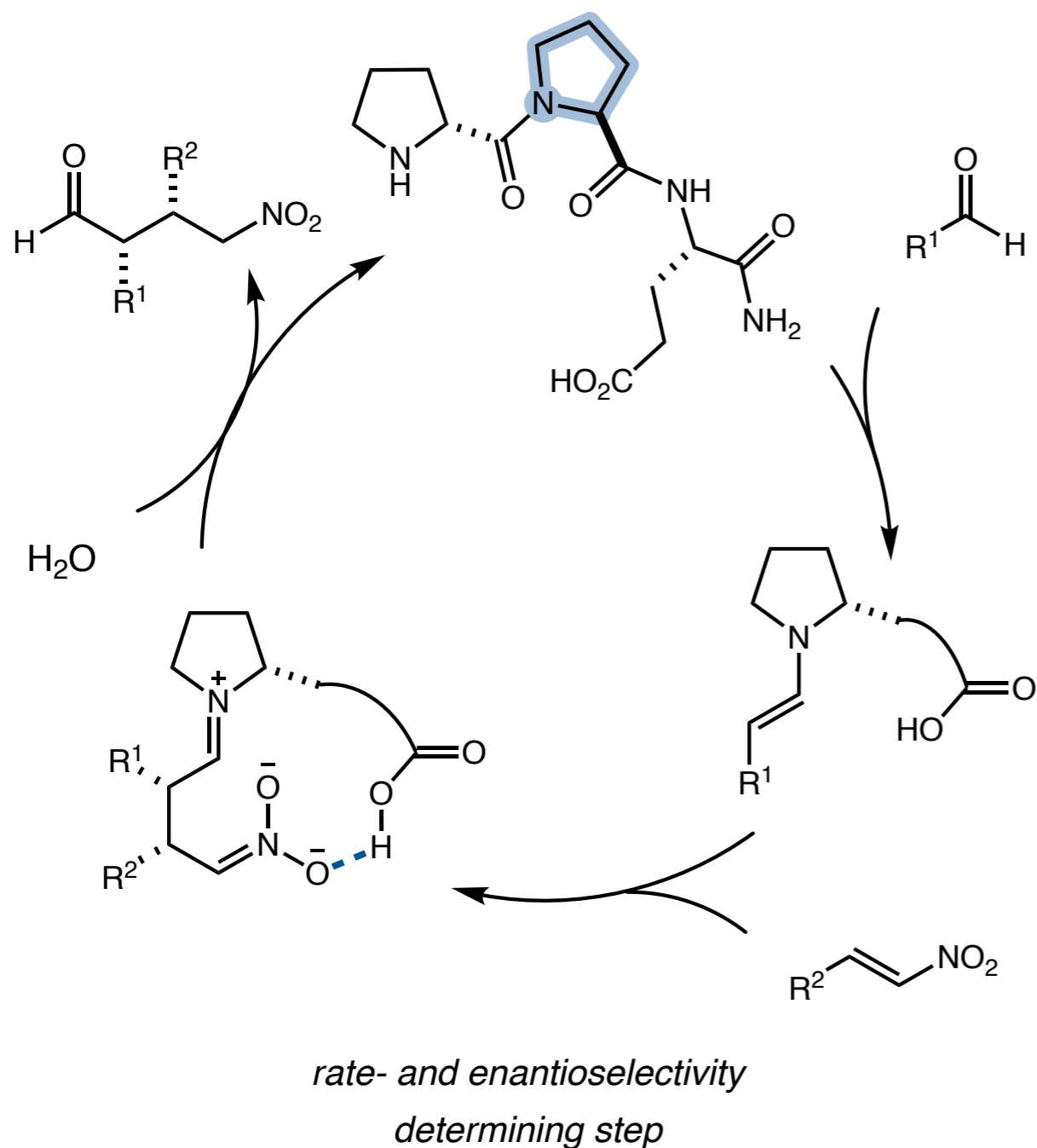


16 examples with **>90% ee**

reversal of selectivity by switching the enantiomer of a single amino acid

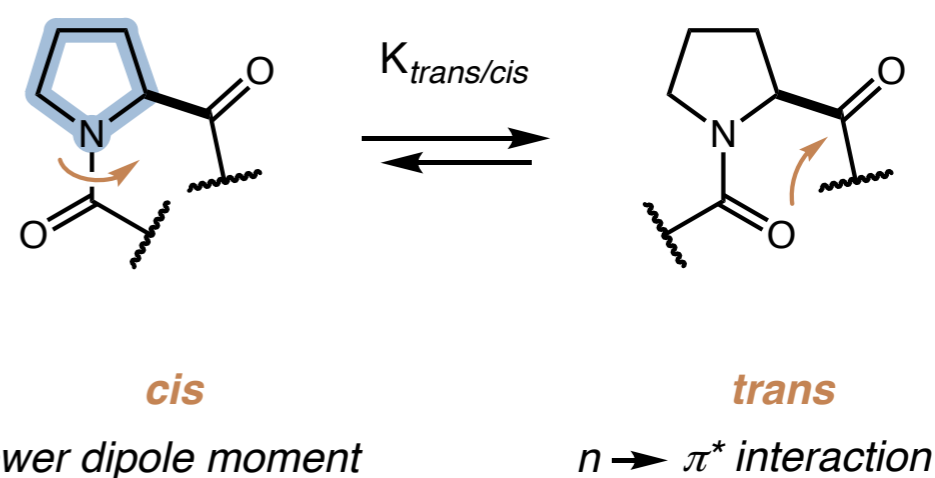
| chiral peptide | conversion | syn/anti | ee |
|--|------------|----------|-------------|
| H-Pro-Pro-Asp-NH ₂ | 96% | 10:1 | -85% |
| H- D -Pro-Pro-Asp-NH ₂ | 93% | 25:1 | 95% |

Wennemers' Tripeptide Catalyst



structure activity relationship

- free carboxylic acid required
- D-Pro-Pro moiety is critical
- conformational flexibility in the backbone tolerated
- $K_{trans/cis}$ ratio influences selectivity

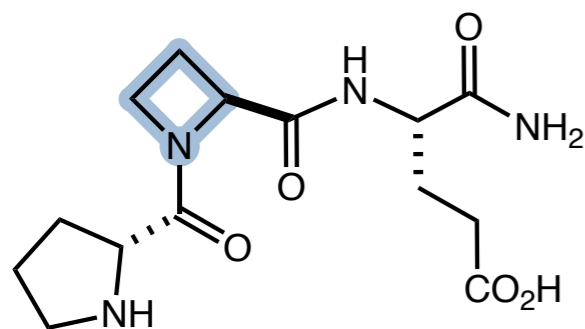


Siebler, C.; Maryasin, B.; Kuemin, M.; Erdmann, R. S.; Rigling, C.; Grünenfelder, C.; Ochsenfeld, C.; Wennemers, H. *Chem. Sci.* **2015**, *6*, 6725.

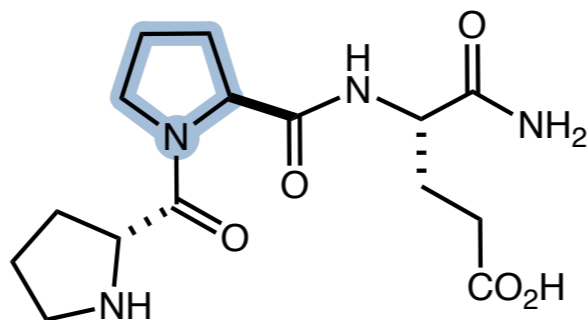
Bächle, F.; Duschmalé, J.; Ebner, C.; Pfaltz, A.; Wennemers, H. *Angew. Chem., Int. Ed.* **2013**, *52*, 12619.

Wiesner, M.; Neuburger, M.; Wennemers, H. *Chem. Eur. J.* **2009**, *15*, 10103.

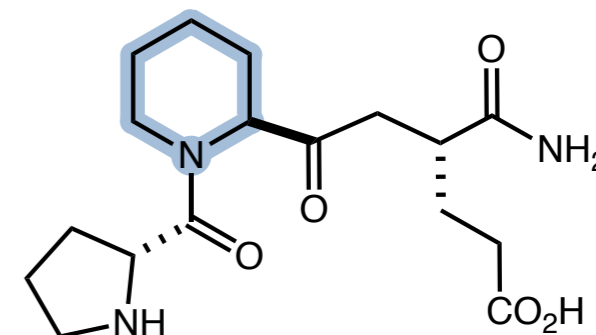
Wennemers' Tripeptide Catalyst



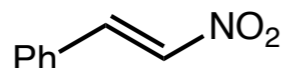
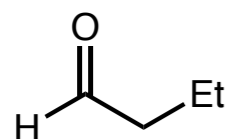
10:1 *trans/cis*



46:1 *trans/cis*

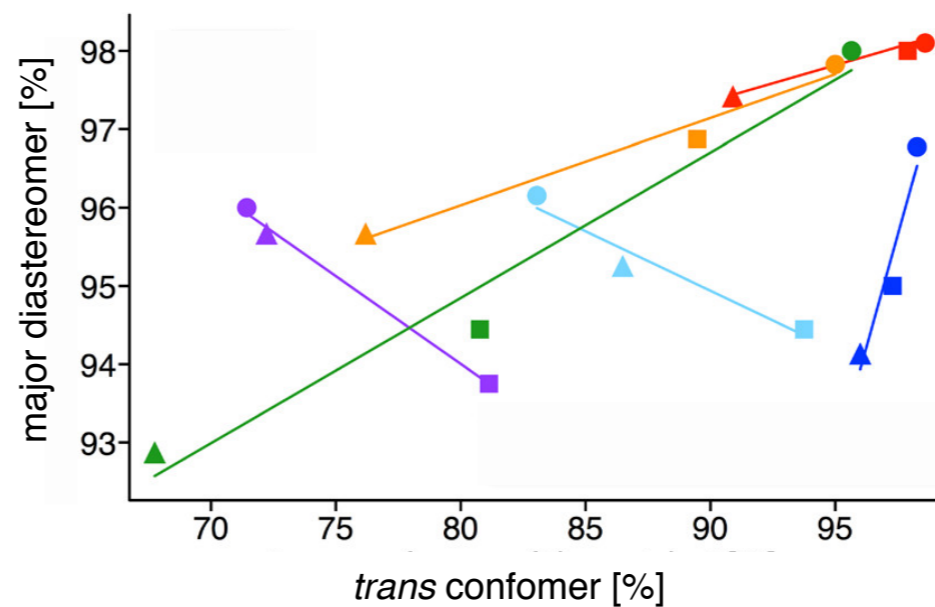
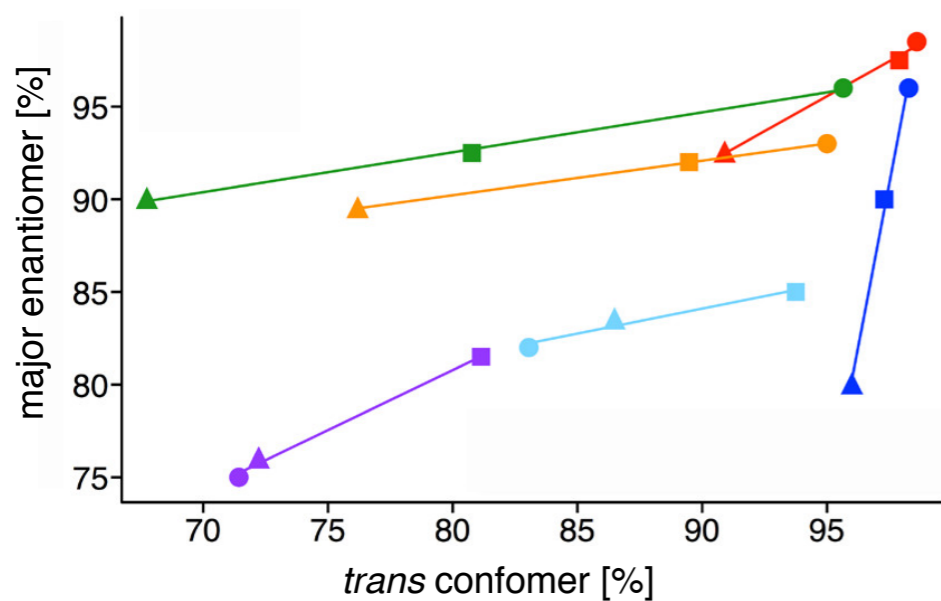
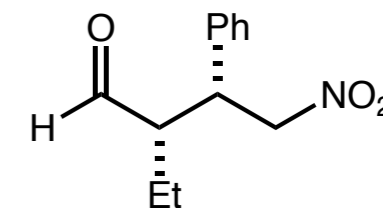


71:1 *trans/cis*



1 mol% chiral peptide

1 mol% NMM, 20 °C, 2 h



9:1 CHCl₃/MeOH

MeOH

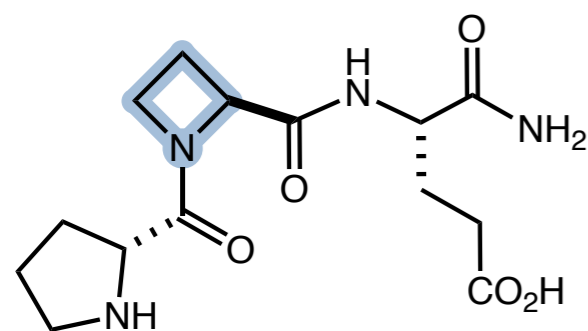
Dioxane

THF

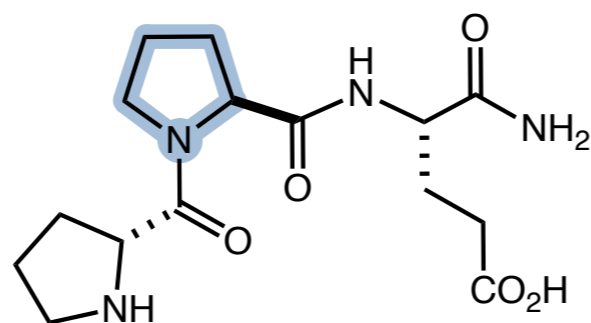
MeCN

DMF

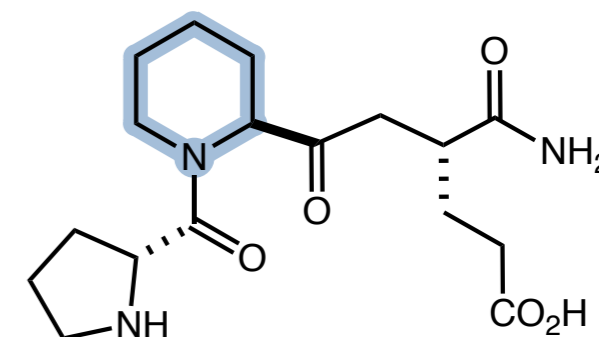
Wennemers' Tripeptide Catalyst



10:1 *trans/cis*



46:1 *trans/cis*



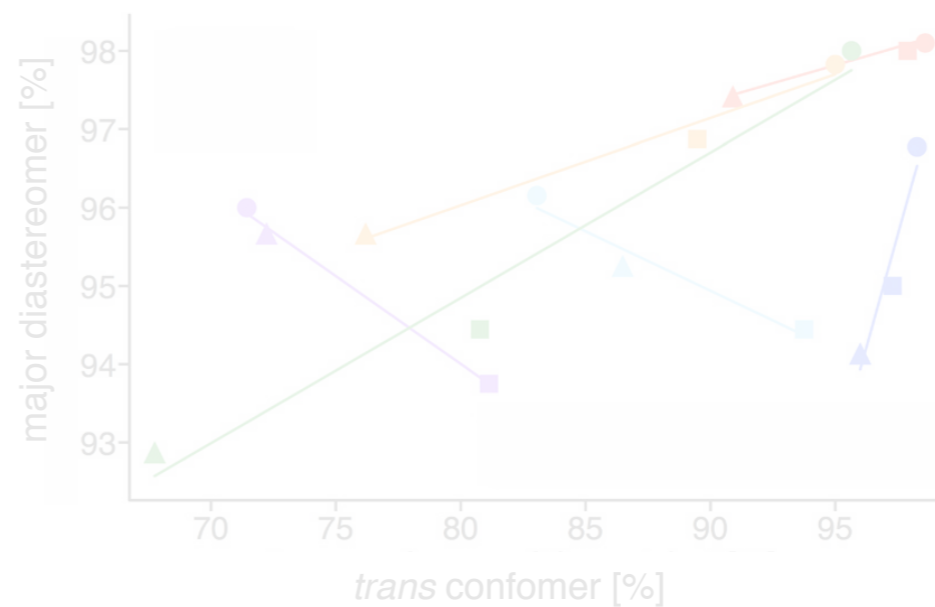
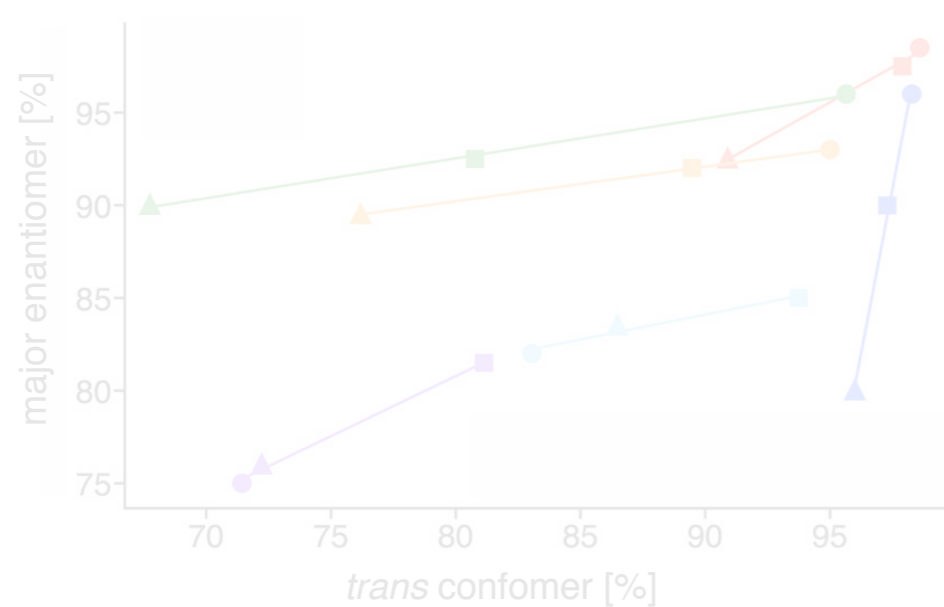
71:1 *trans/cis*



A higher proportion of the *trans* conformer leads to a higher enantio- and diastereoselectivity.

1 mol% NMM, 20 °C, 2 h

Et



9:1 CHCl₃/MeOH

MeOH

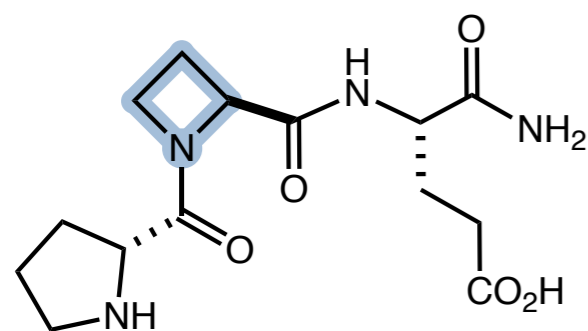
Dioxane

THF

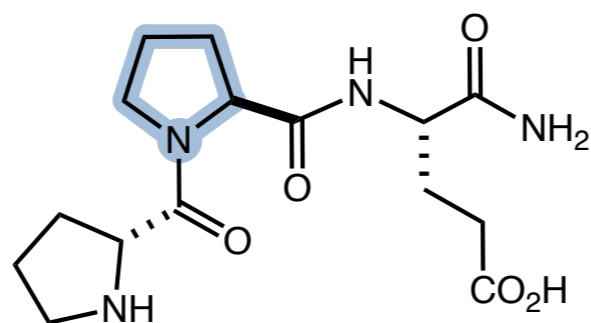
MeCN

DMF

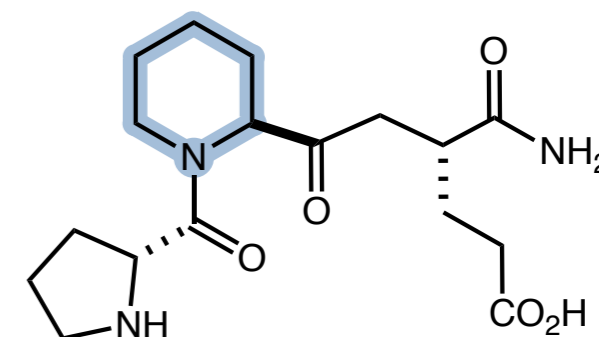
Wennemers' Tripeptide Catalyst



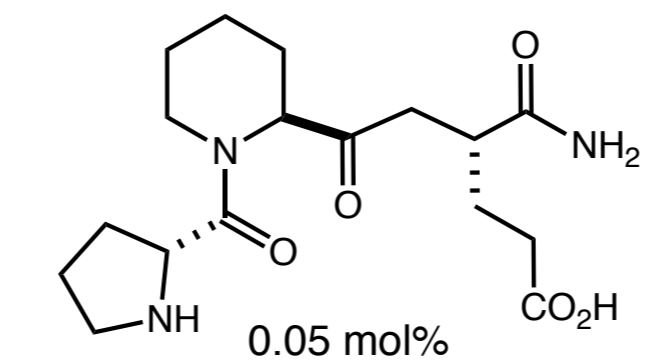
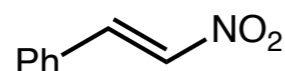
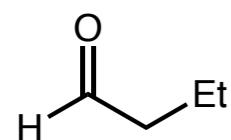
10:1 *trans/cis*



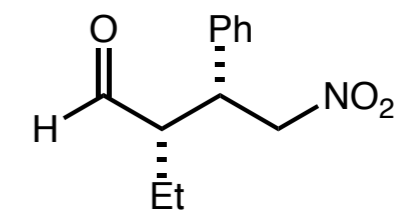
46:1 *trans/cis*



71:1 *trans/cis*



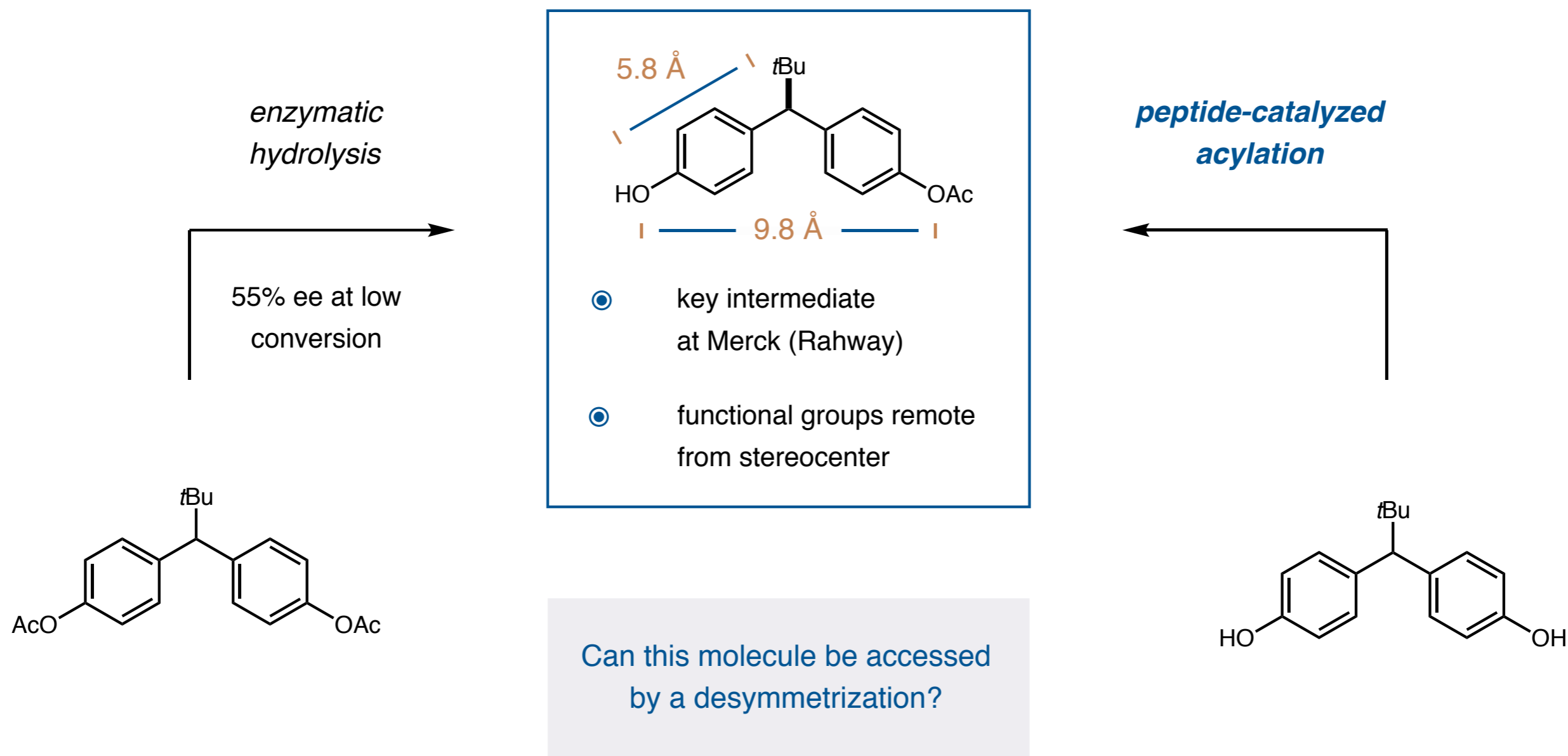
0.05 mol% NMM, 9:1
CHCl₃/iPrOH, 20 °C, 3 d



95%, 60.1 d.r., **99% ee**

Lowest catalyst loading of a secondary amine organocatalyst reported to date.

Remote Desymmetrization by Chiral Peptides

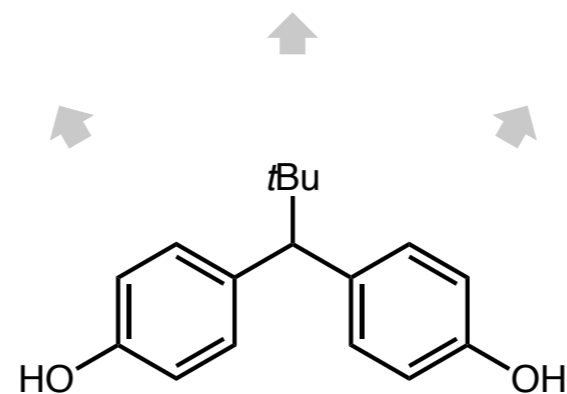
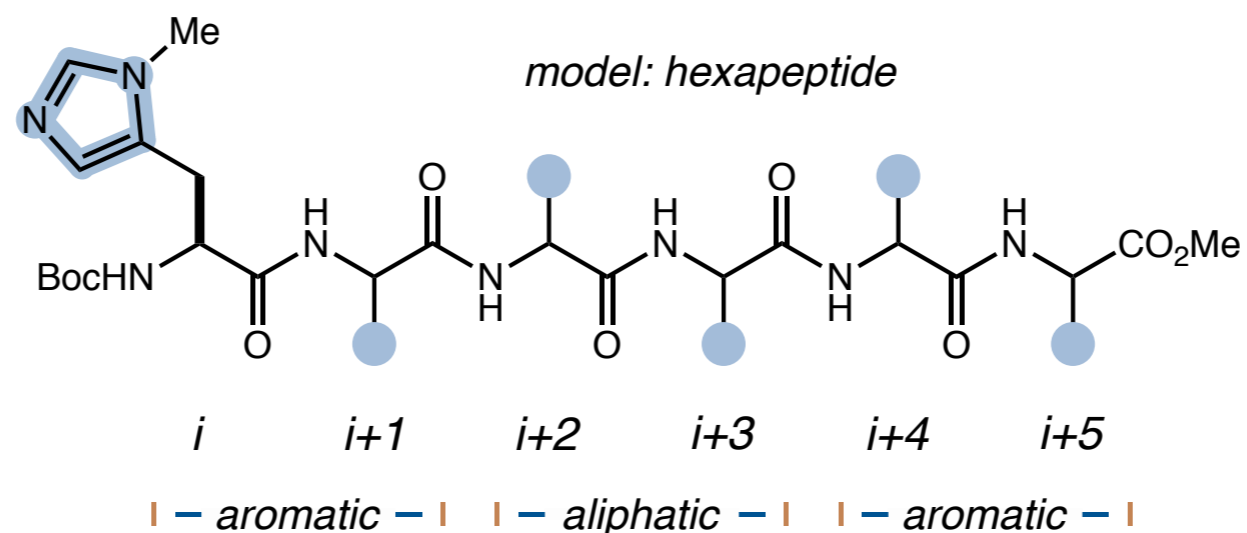


Lewis, C. A.; Gustafson, J. L.; Chiu, A.; Balsells, J.; Pollard, D.; Murry, J.; Reamer, R. A.; Hansen, K. B.; Miller, S. J., *T. J. Am. Chem. Soc.* **2008**, *130*, 16358. Lewis, C. A.; Chiu, A.; Kubryk, M.; Balsells, J.; Pollard, D.; Esser, C. K.; Murry, J.; Reamer, R. A.; Hansen, K. B.; Miller, S. J., *T. J. Am. Chem. Soc.* **2006**, *128*, 16454.

Remote Desymmetrization by Chiral Peptides

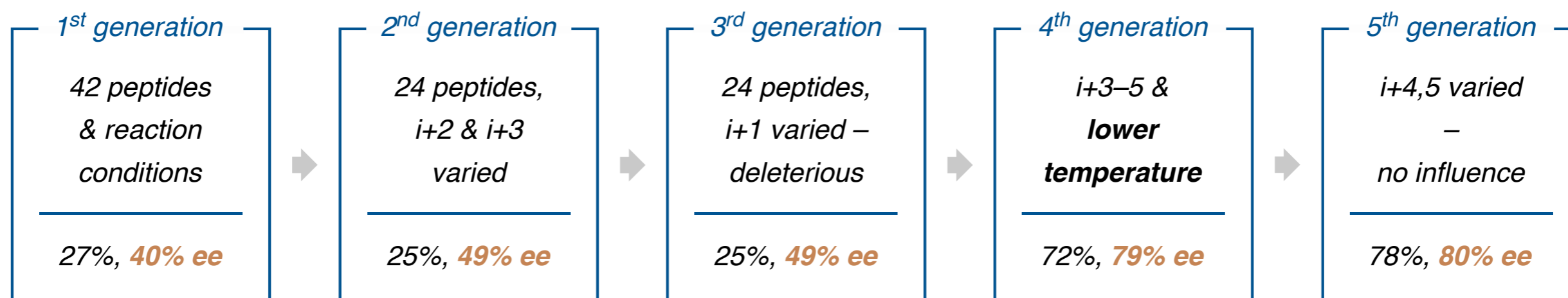
■ Design of the initial library

*active moiety
previously identified*



Case Study 2: Remote Desymmetrization by Chiral peptides

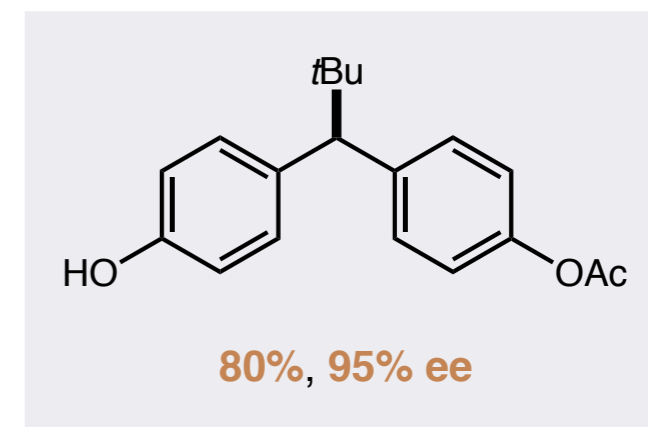
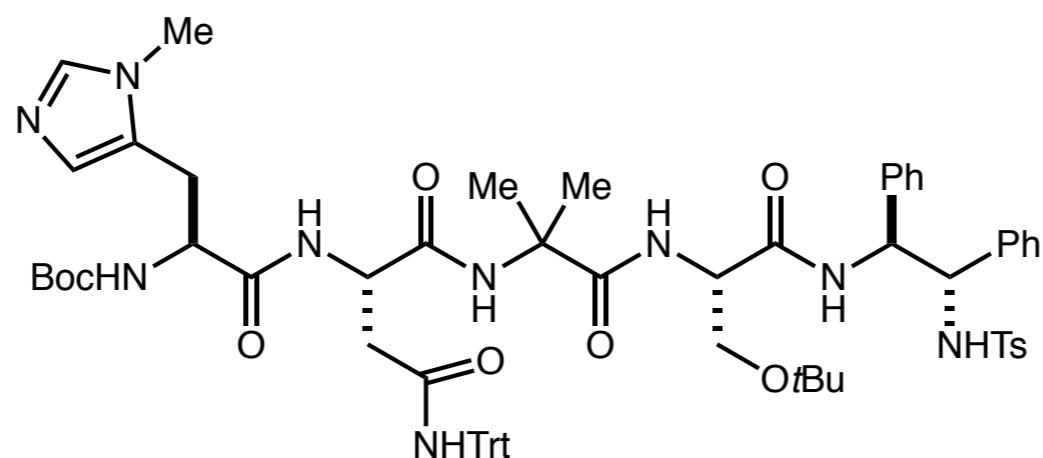
■ Optimization workflow



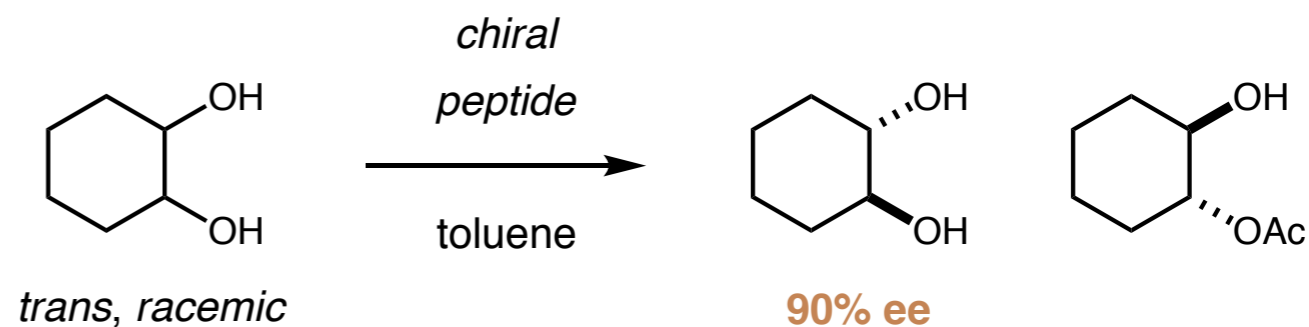
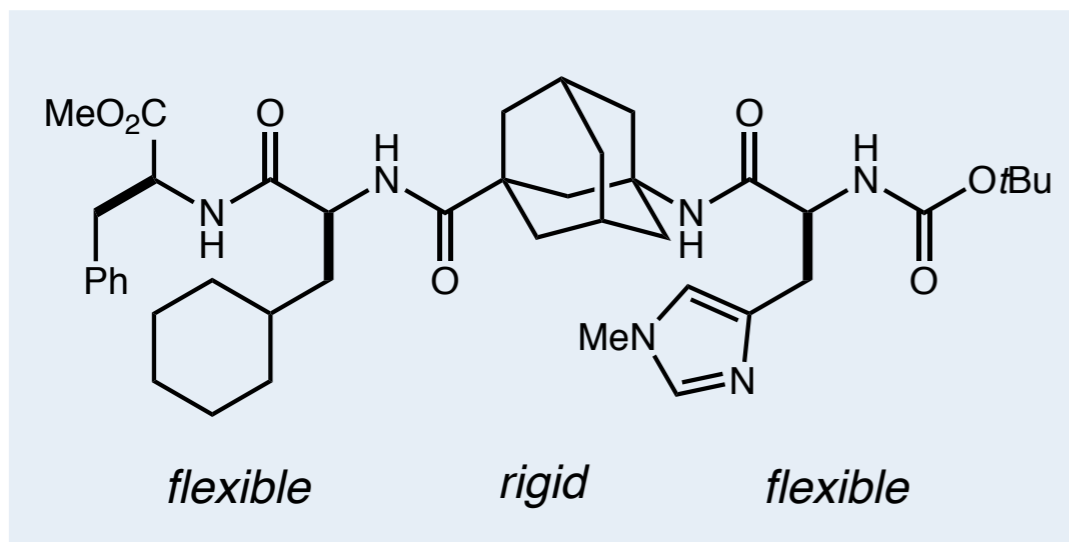
truncation to tetrapeptide



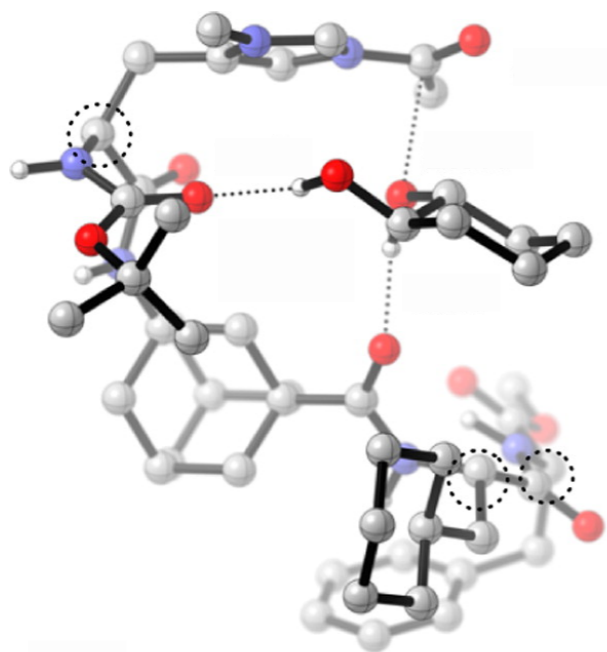
variation of C-terminus



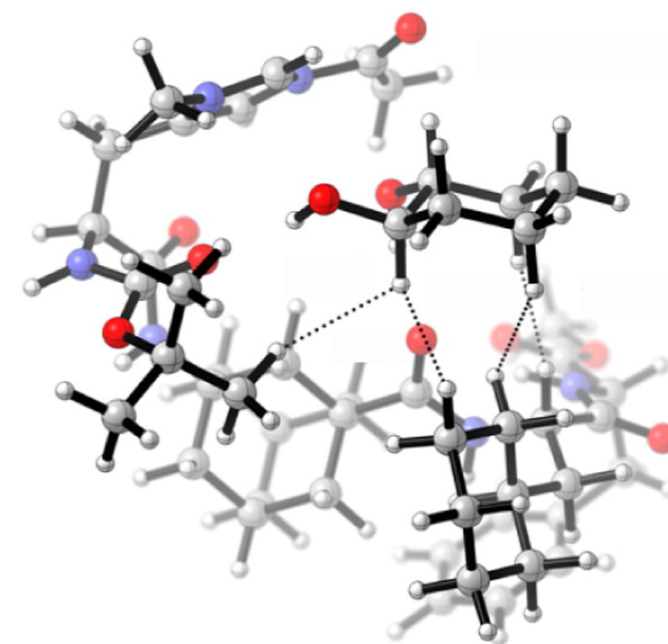
Schreiner's Lipophilic Oligopeptide Catalyst



Mechanistic information



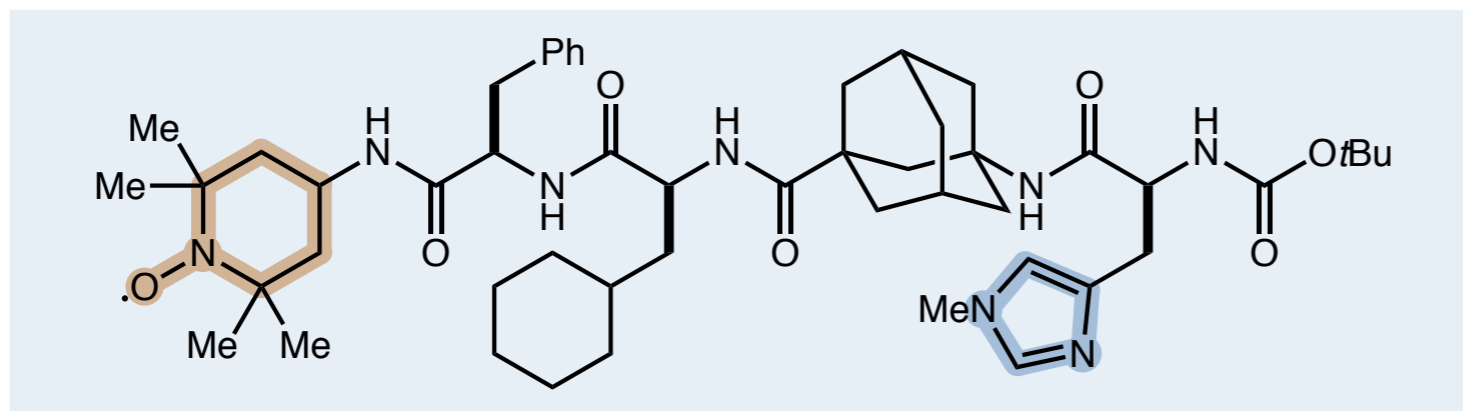
- no secondary structure - central adamantyl group separates both ends
- adamantyl group holds the 3 stereocenters that determine stereochemistry in place
- H-bonding to second hydroxyl group
- Dispersion interaction between hydrophobic substituents and substrate



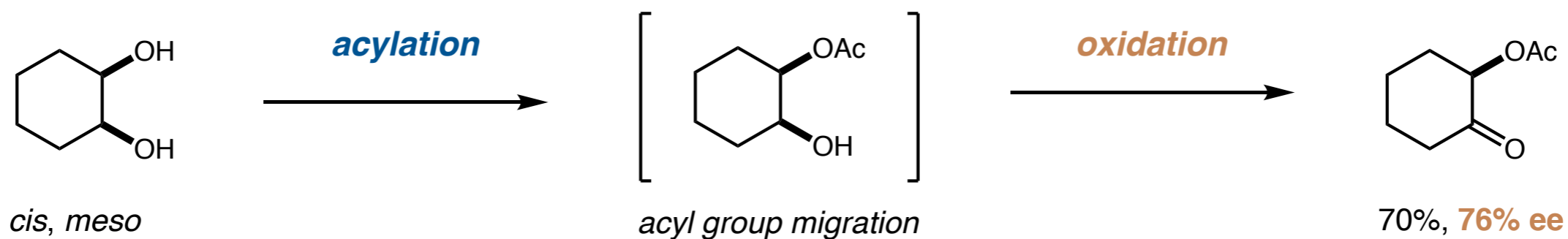
Müller, C. E.; Zell, D.; Hrdina, R.; Wende, R. C.; Wanka, L. Schuler, S. M. M.; Schreiner, P. R. *J. Org. Chem.* **2013**, *78*, 8465.

Müller, C. E.; Wanka, L.; Jewell, K.; Schreiner, P. R. *Angew. Chem., Int. Ed.* **2008**, *47*, 6180.

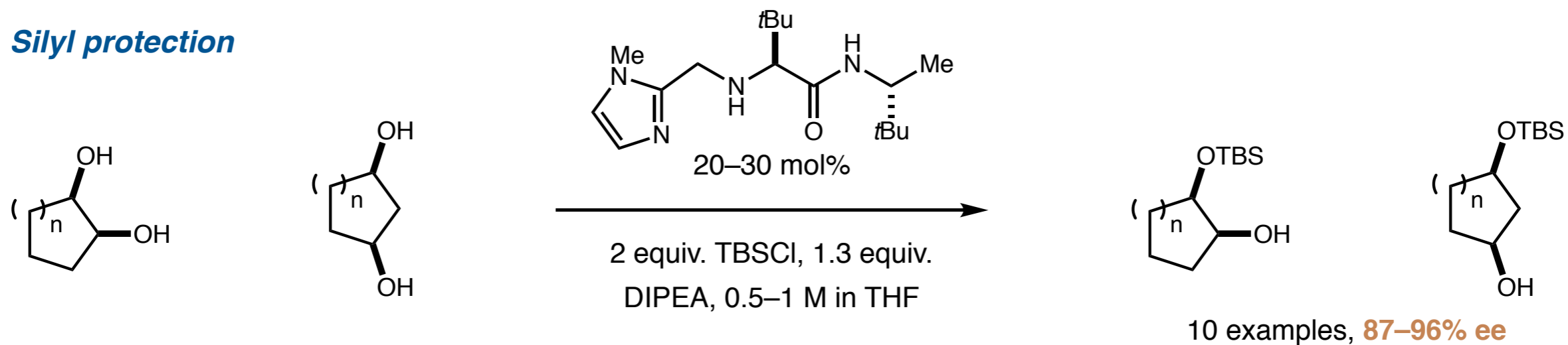
Desymmetrization of cis-Diols



two catalytically active species on a single peptide



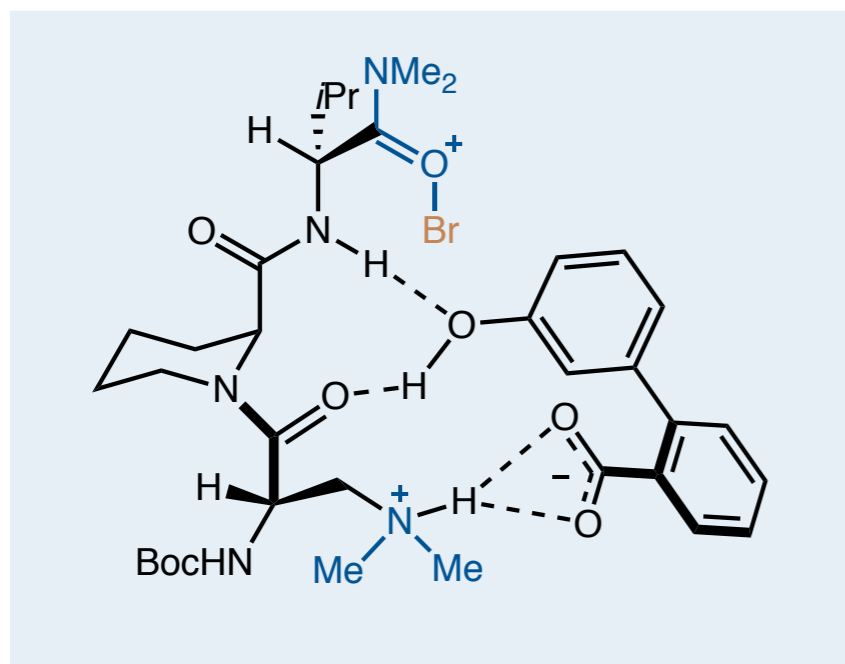
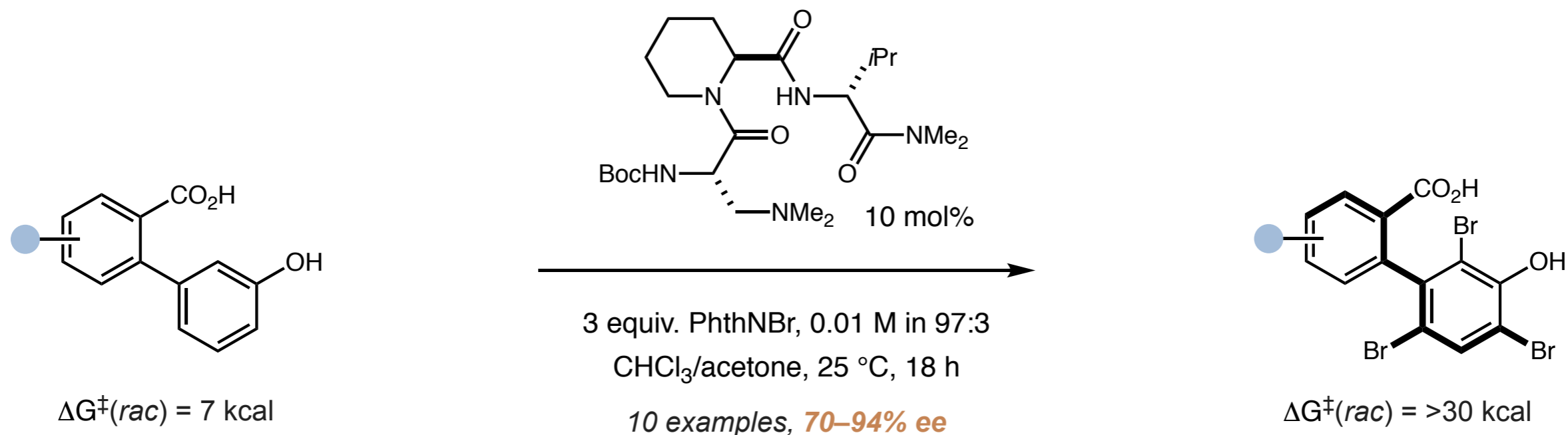
Silyl protection



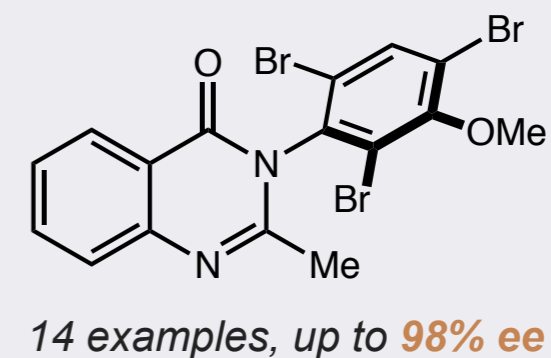
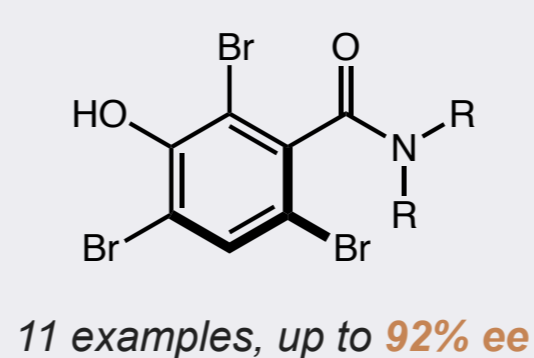
Zhao, Y.; Rodrigo, J.; Hoveyda, A. H.; Snapper, M. L. *Nature* **2006**, 443, 67.

Müller, C. E.; Hrdina, R.; Wende, R. C.; Schreiner, P. R. *Chem. Eur. J.* **2011**, 17, 6309.

Atroposelective Bromination



Related projects

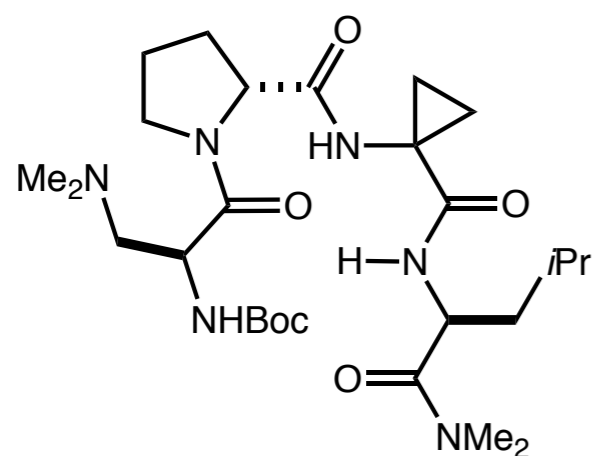


reoptimized peptide backbones used

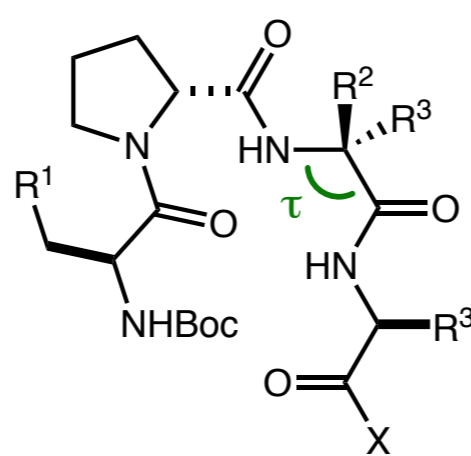
Diener, M. E.; Metrano, A. J.; Kusano, S.; Miller, S. J. *J. Am. Chem. Soc.* **2015**, *137*, 12369. Barrett, K. T.; Miller, S. J. *J. Am. Chem. Soc.* **2013**, *135*, 2963. Garand, E.; Kamrath, M. Z.; Jordan, P. A.; Wolk, A. B.; Leavitt, C. M.; McCoy, A. B.; Miller, S. J.; Johnson, M. A. *Science* **2012**, *335*, 694.

Gustafson, J. L.; Lim, D.; Miller, S. J. *Science* **2010**, *328*, 1251.

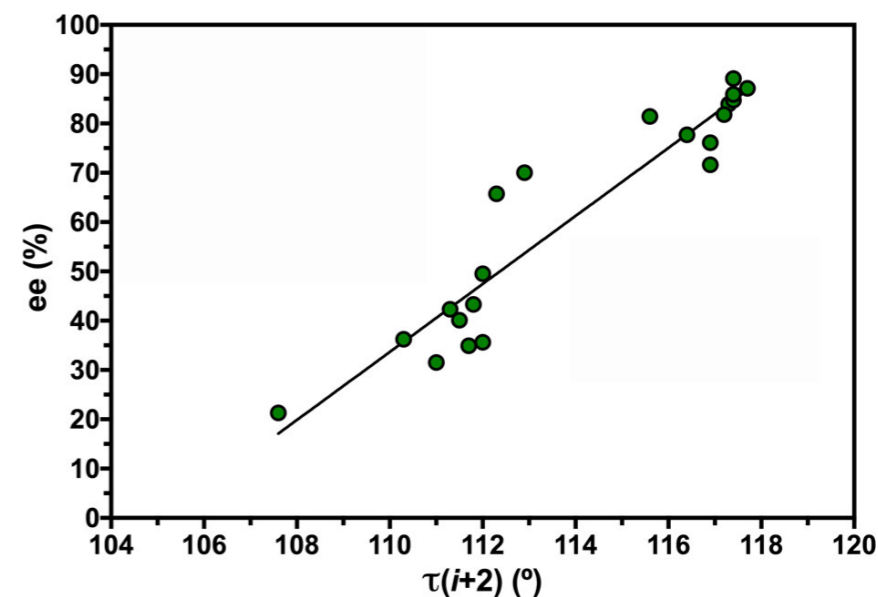
Atroposelective Bromination



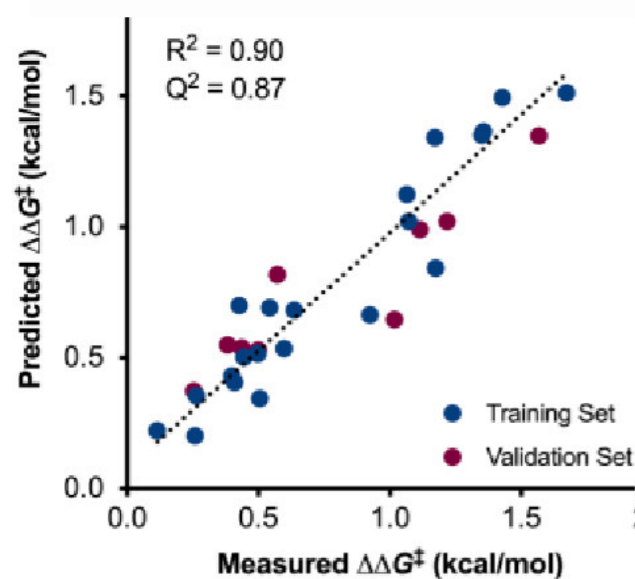
3 different conformers observed
by x-ray crystallography



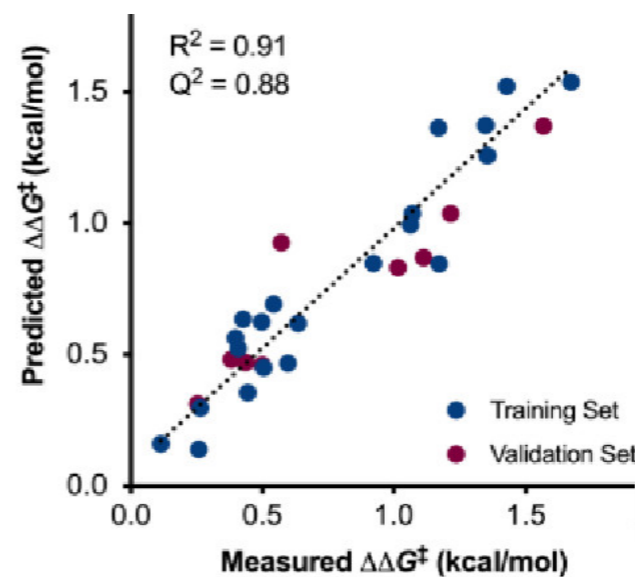
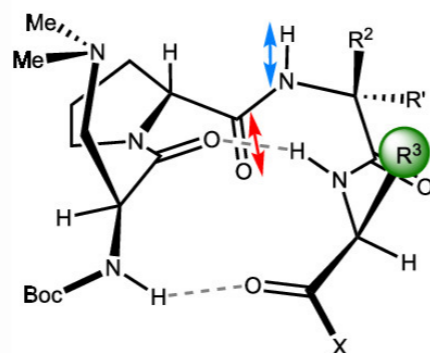
35 distinct peptide –
53 crystal structures



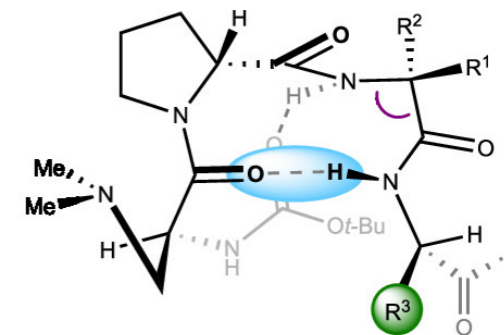
wider τ -angle \rightarrow more flexible & **more selective**



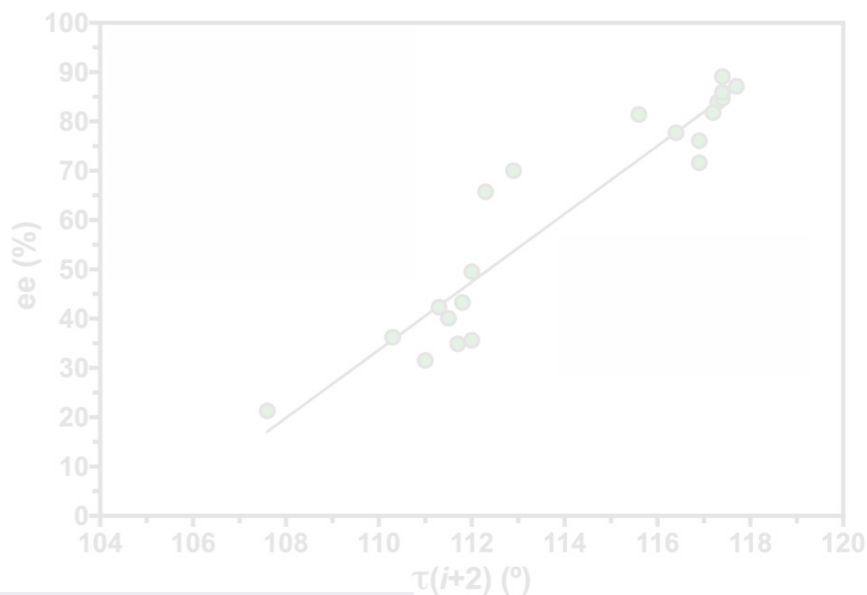
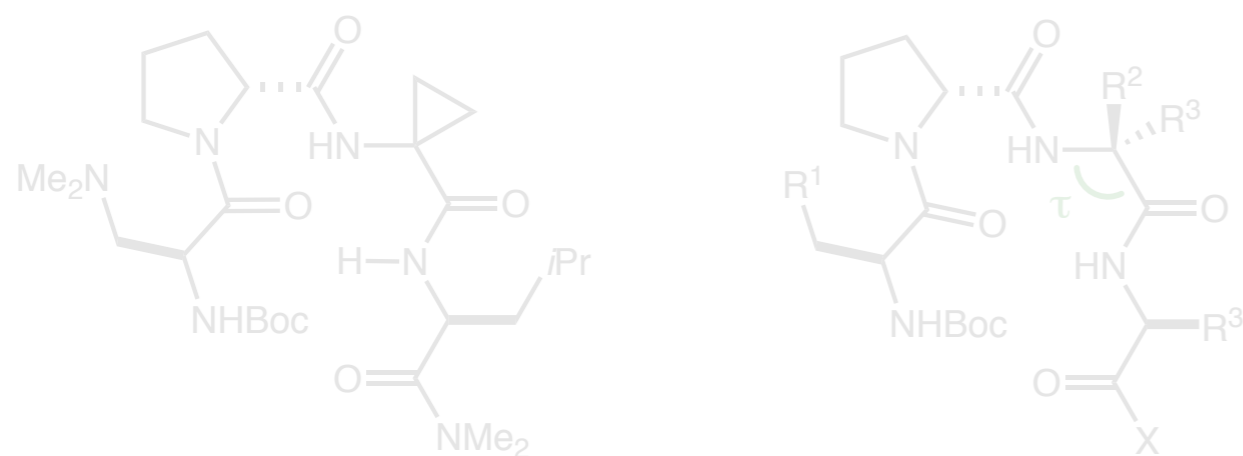
type II' β -turns



type I' β -turns



Atroposelective Bromination

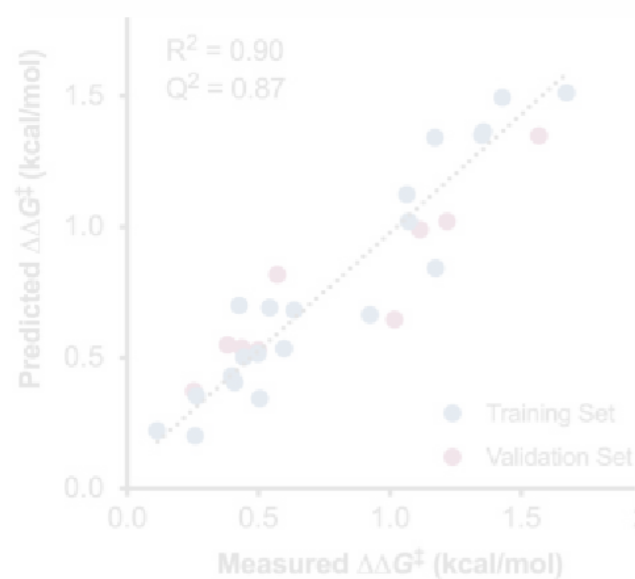


3 different conformers observed
by x-ray crystallography

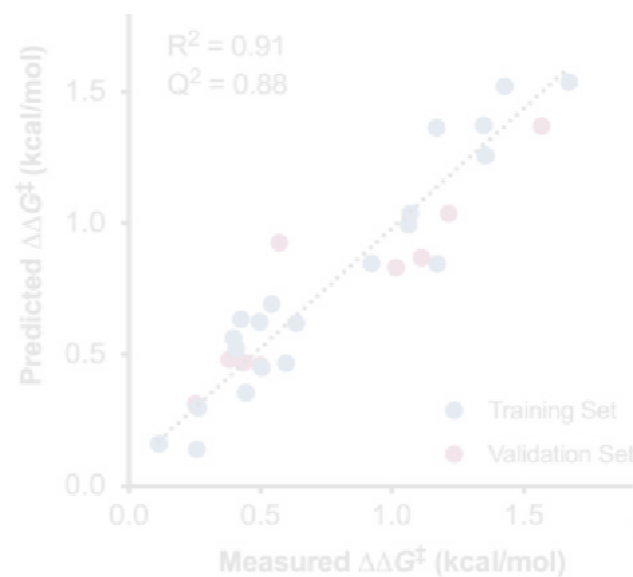
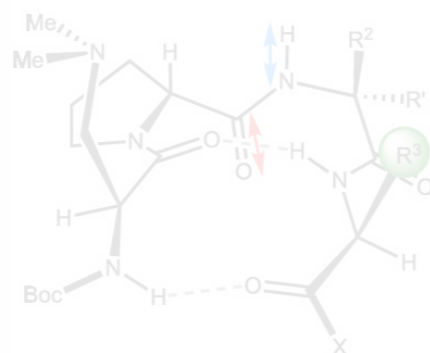
35 distinct peptide

Multiple conformers contribute to a transition state ensemble.

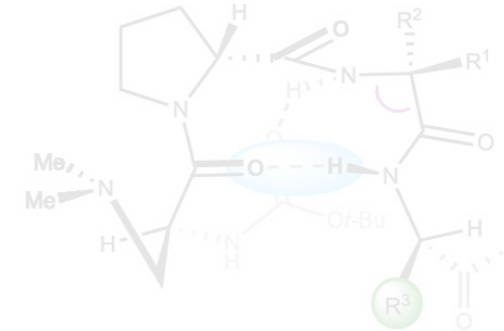
flexible & more selective



type II' β -turns

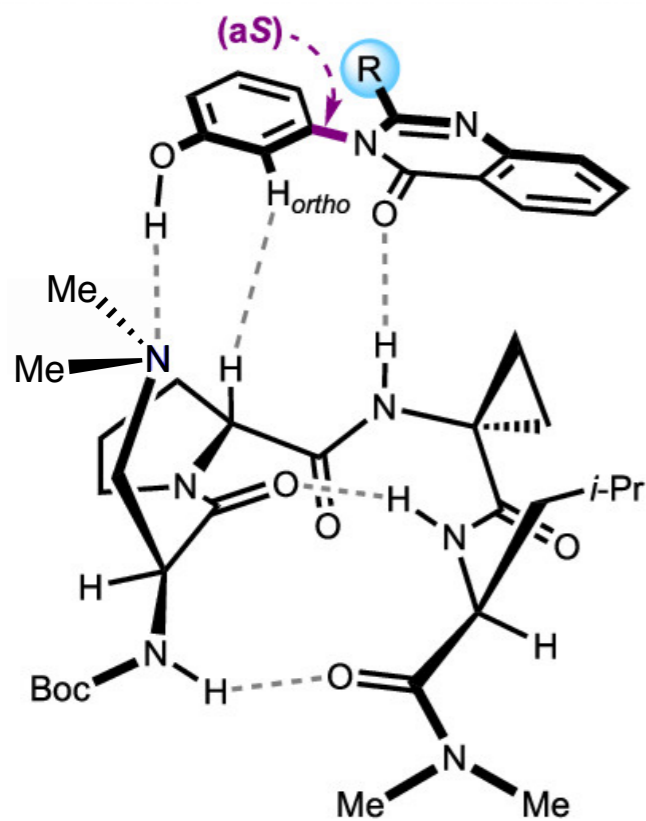


type I' β -turns



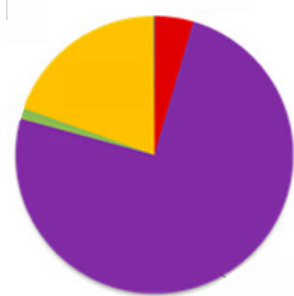
Crawford, J. M.; Stone, E. A.; Mertrano, A. J.; Miller, S. J.; Sigman, M. S. *J. Am. Chem. Soc.* **2018**, *140*, 868. Mertrano, A. J.; Abascal, N. C.; Mercado, B. Q.; Paulson, E. K.; Hurtley, A. E.; Miller, S. J. *J. Am. Chem. Soc.* **2017**, *139*, 491. Mertrano, A. J.; Abascal, N. C.; Mercado, B. Q.; Paulson, E. K.; Miller, S. J. *Chem. Commun.* **2016**, *52*, 4816.

Atroposelective Bromination

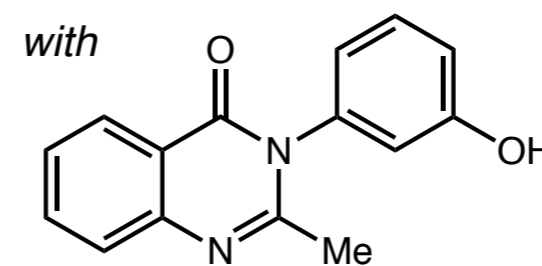


type II' β -turn

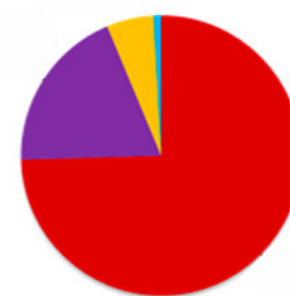
only peptide –
ground state



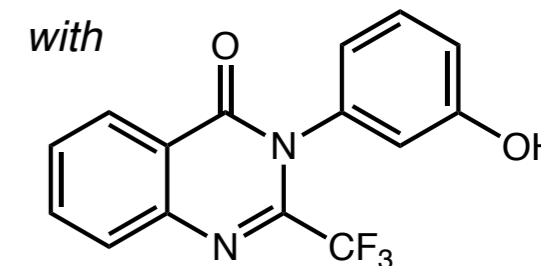
5% vs 75%



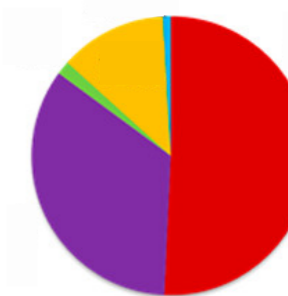
94% ee



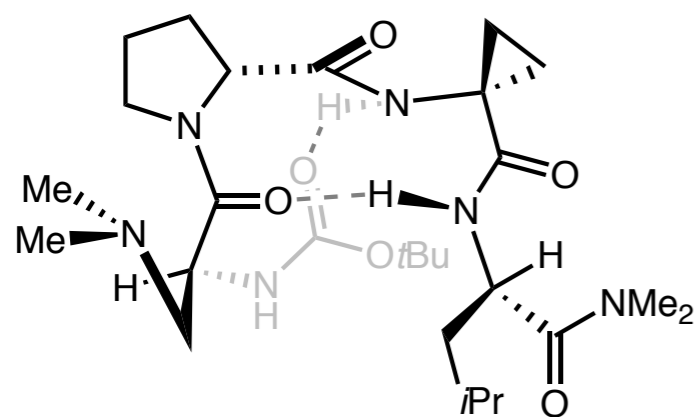
75% vs 19%



26% ee

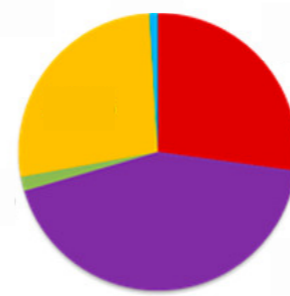


51% vs 34%



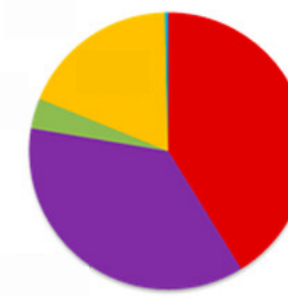
type I' β -turn

opposite enantiomer



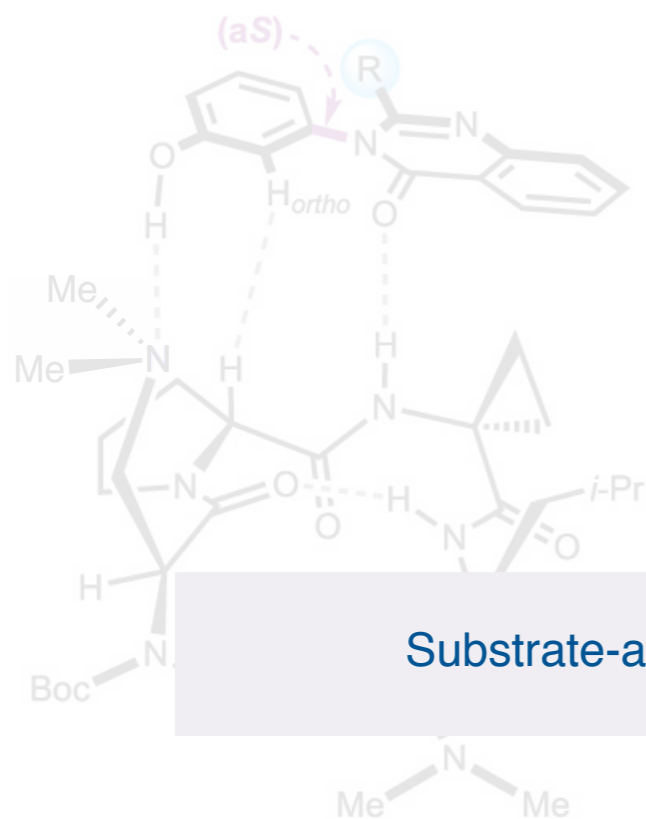
27% vs 43%

opposite enantiomer

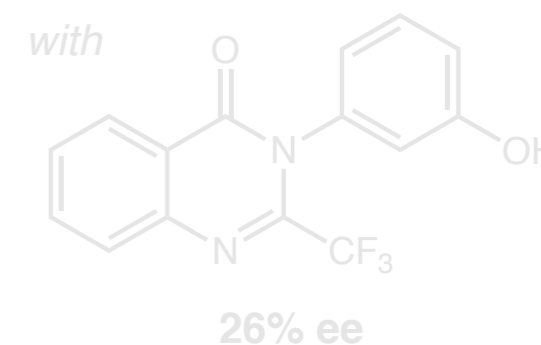
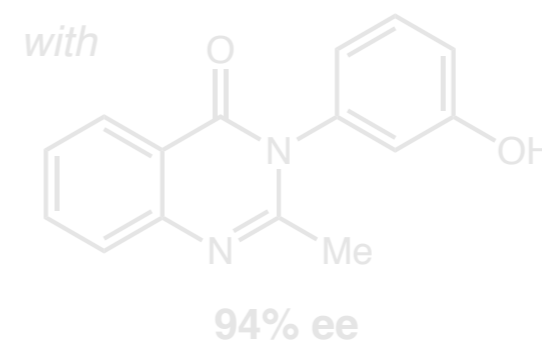


41% vs 36%

Atroposelective Bromination



only peptide –
ground state



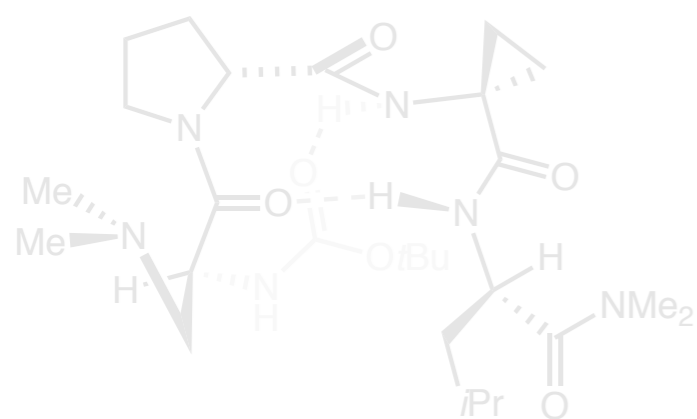
Substrate-association enforces a structural change of the peptide – induced fit.

type II' β -turn

5% vs 75%

75% vs 19%

51% vs 34%



type I' β -turn

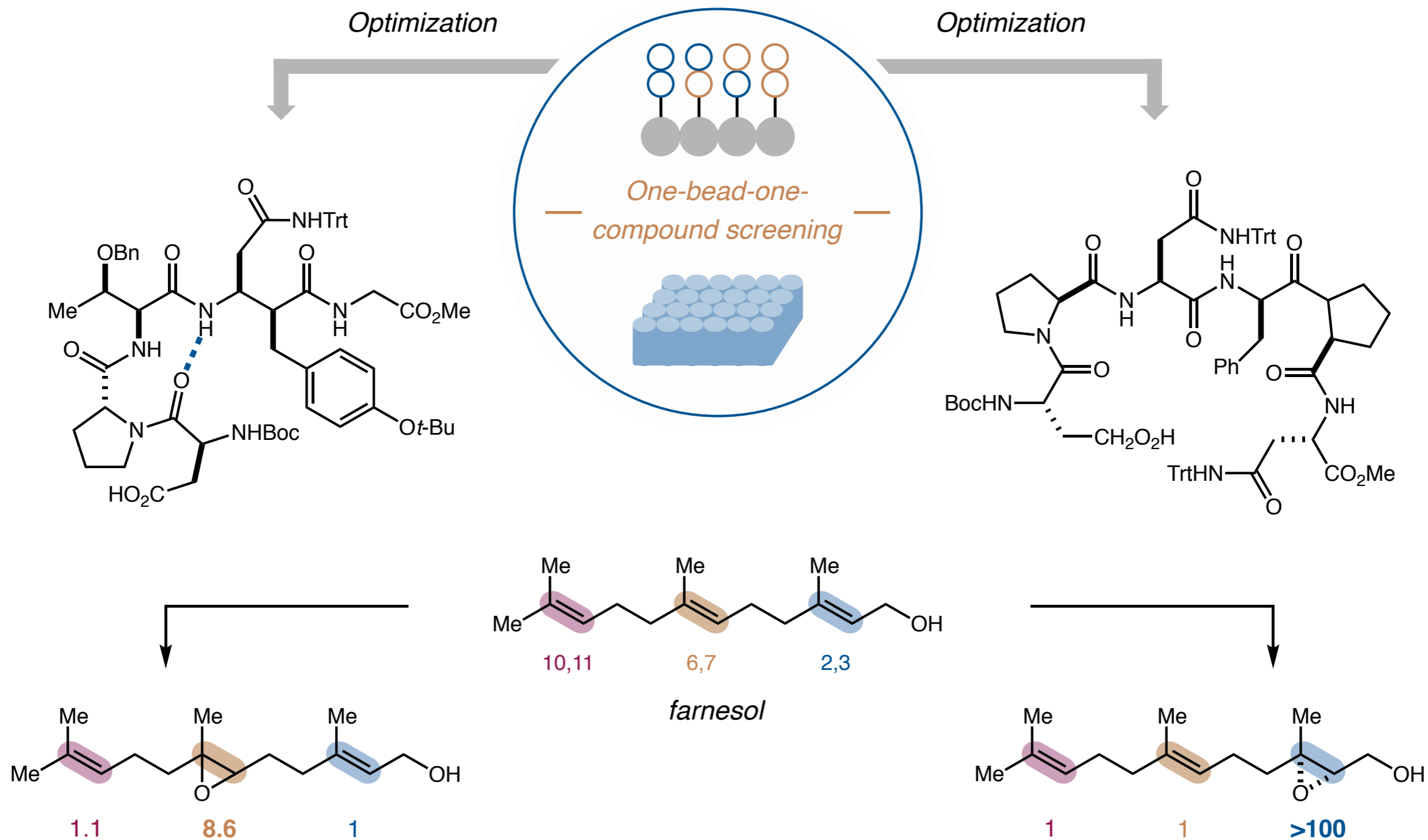
opposite enantiomer

27% vs 43%

opposite enantiomer

41% vs 36%

Site-Selective Polyene Oxidation



Abascal, N. C.; Lichtor, P. A.; Giuliano, M. W.; Miller, S. J. *Chem. Sci.* **2014**, *5*, 4504. Lichtor, P. A.; Miller, S. J. *J. Am. Chem. Soc.* **2014**, *136*, 5301.

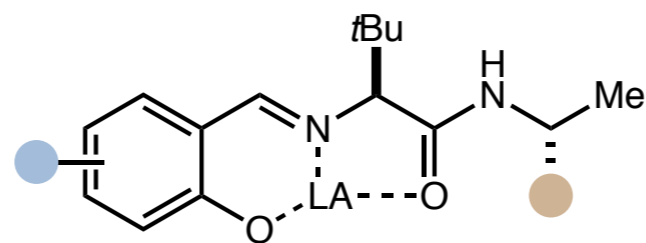
Lichtor, P. A.; Miller, S. J. *Nat. Chem.* **2012**, *4*, 990.

Small Peptides as Organocatalysts

- Introduction
- Catalytically relevant properties of small peptides
- Small peptides as organocatalyst
- **Small peptides in Lewis acid and transition metal catalysis**
- Small peptides in photoredox catalysis
- Summary

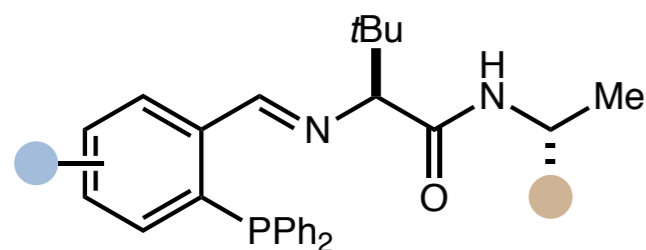
This group meeting does not cover the available literature comprehensively. Instead, selected key studies are discussed in order to explain the underlying concepts.

Pioneering Work on Peptides in Lewis Acid and Transition Metal Catalysis



LA = Ti, Zr, Al

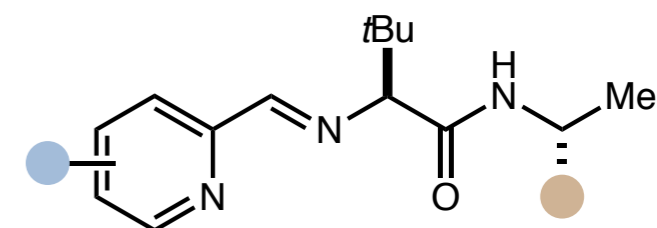
Snapper & Hoveyda: cyanation, alkylation



*Hoveyda: copper-catalyzed
conjugate additions*

Role of the peptide

creation of a chiral
environment around
the metal catalyst

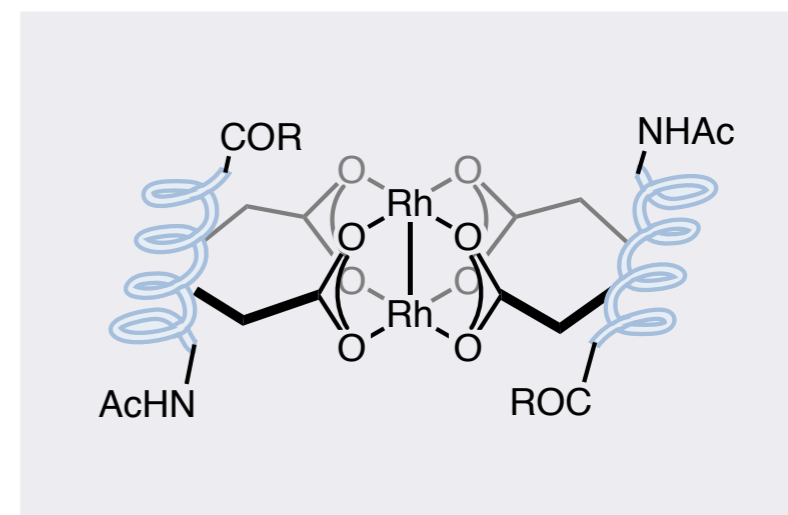
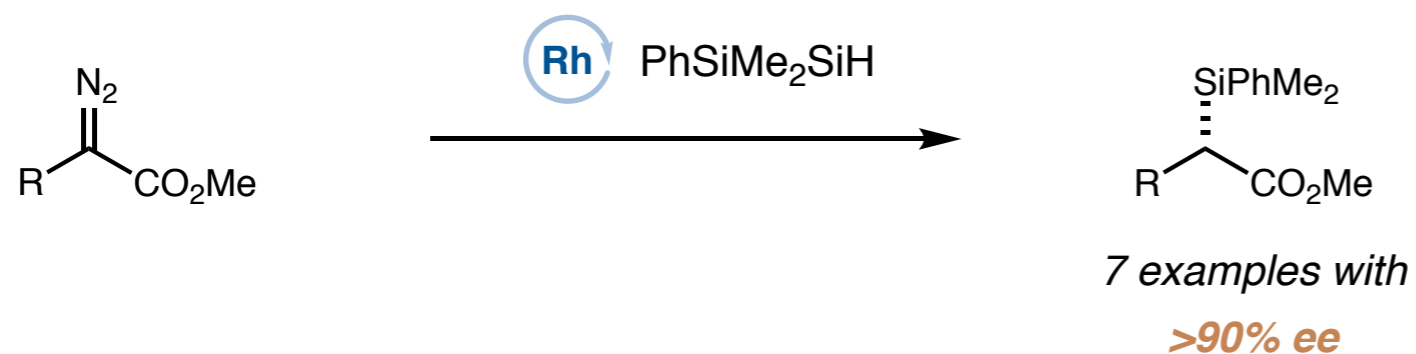


*Hoveyda: copper-catalyzed
allylic alkylation*

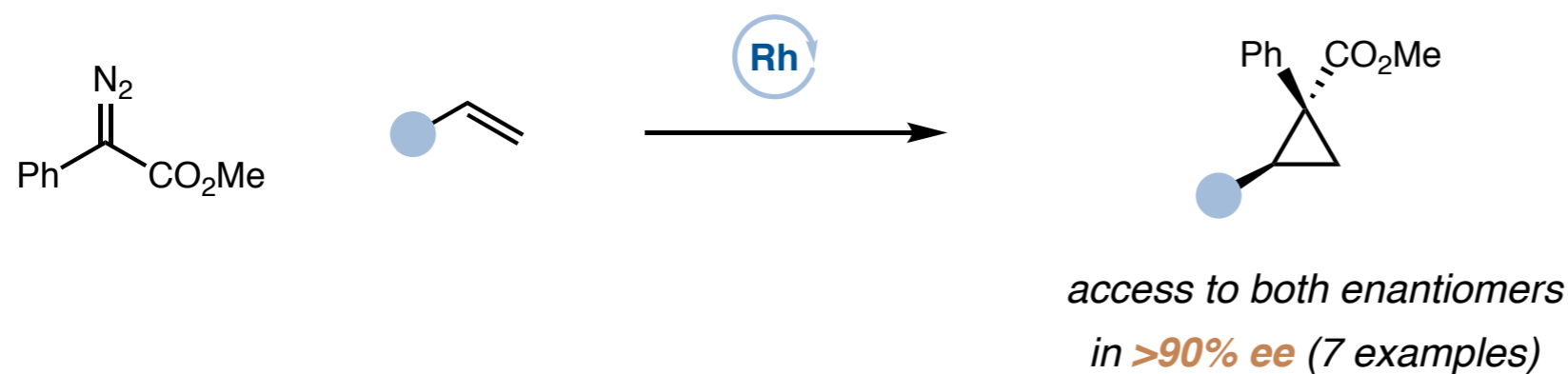
Key references: Deng, H.; Isler, M. P.; Snapper, M. L.; Hoveyda, A. H. *Angew. Chem., Int. Ed.* **2002**, *41*, 1009. Degrado, S. J.; Mizutani, H.; Hoveyda, A. H. *J. Am. Chem. Soc.* **2001**, *123*, 755. Porter, J. R.; Traverse, J. F.; Hoveyda, A. H.; Snapper, M. L. *J. Am. Chem. Soc.* **2001**, *123*, 984. Luchaco-Cullis, C. A.; Mizutani, H.; Murphy, K. E.; Hoveyda, A. H. *Angew. Chem., Int. Ed.* **2001**, *40*, 1456. Krueger, C. A.; Kuntz, K. W.; Dzierba, C. D.; Wirschun, W. G.; Gleason, J. D.; Snapper, M. L.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1999**, *121*, 4284.

Helical Peptide-Ligated Rhodium-“Paddlewheel” Complexes

■ Enantioselective insertion into Si–H bonds



■ Enantioselective cyclopropanation



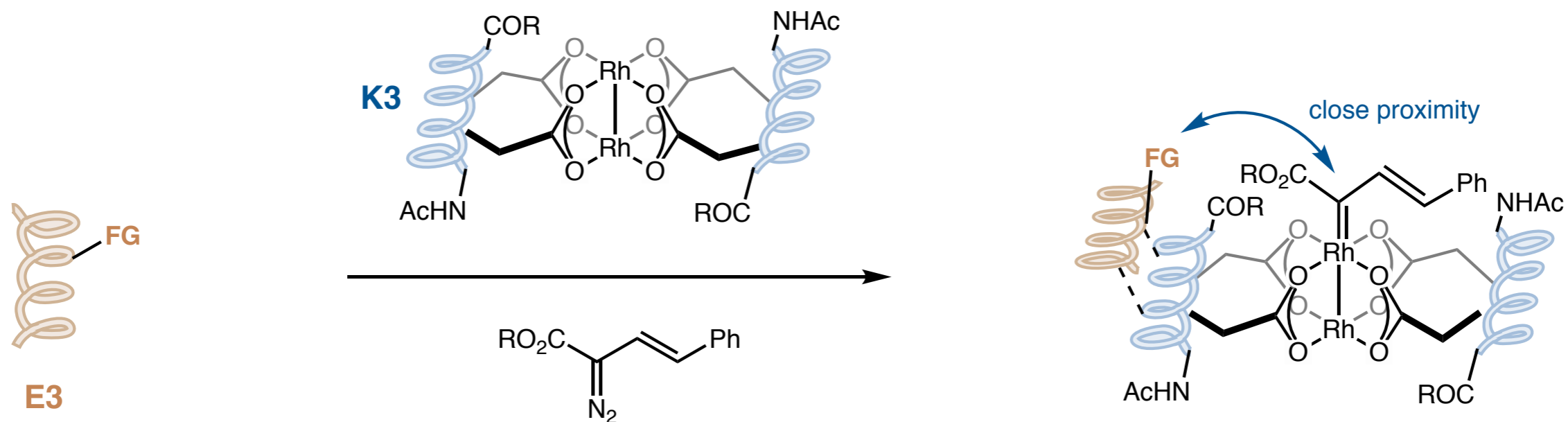
Role of the peptide

creation of a chiral environment around the metal catalyst

Samasivan, R.; Ball, Z. T. *Angew. Chem., Int. Ed.* **2012**, *51*, 8568.

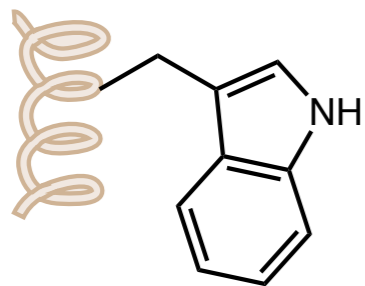
Samasivan, R.; Ball, Z. T. *J. Am. Chem. Soc.* **2010**, *132*, 9289.

Structure-Selective Protein Functionalization



helical peptide

E3/K3 coiled coil assembly places side chain in close proximity of the rhodium carbenoid



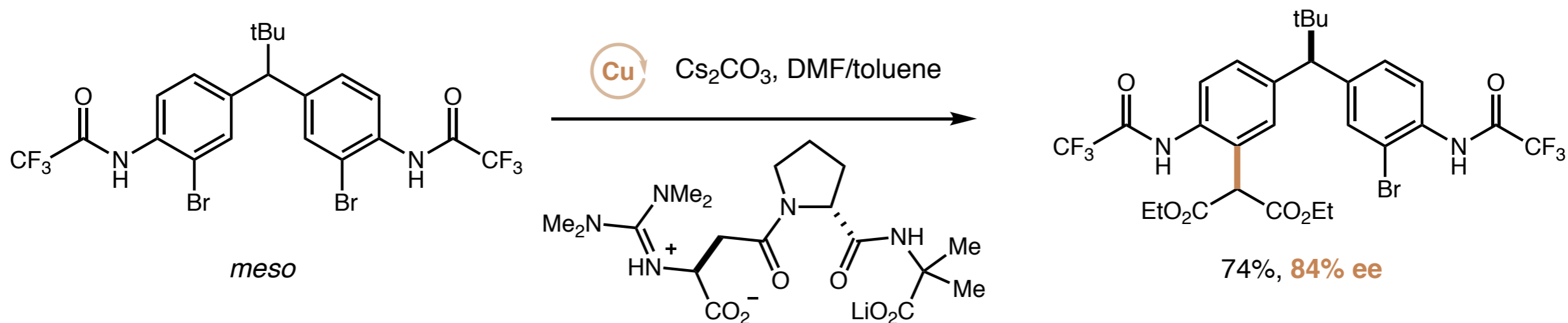
> 1000 rate enhancement compared to $\text{Rh}_2(\text{OAc})_4$

Although tryptophan is most reactive, selective functionalization of E3-tyrosine or phenylalanines occurs even in presence of random tryptophane-containing peptides.

Role of the peptide

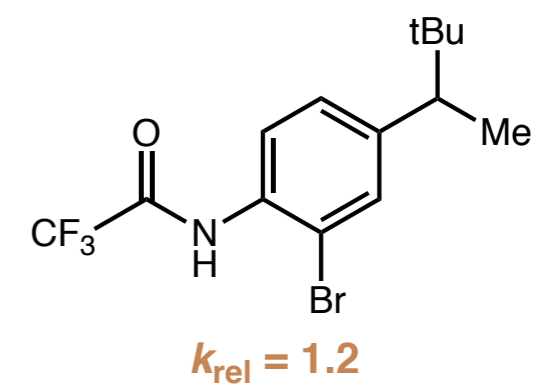
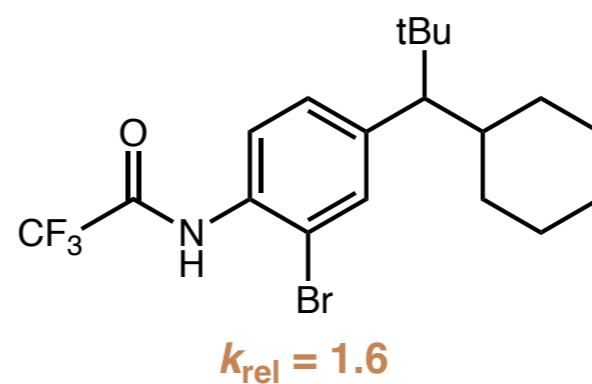
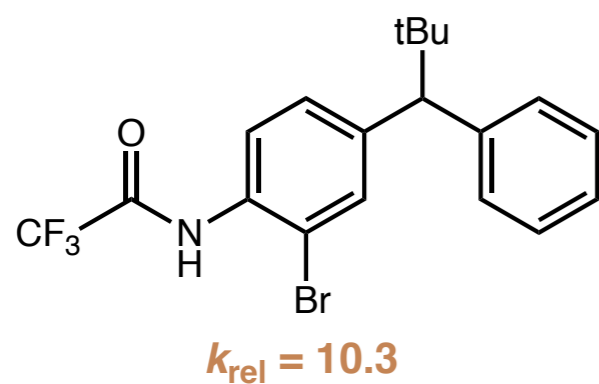
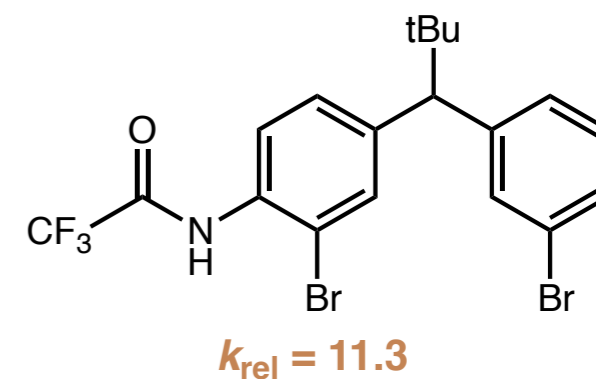
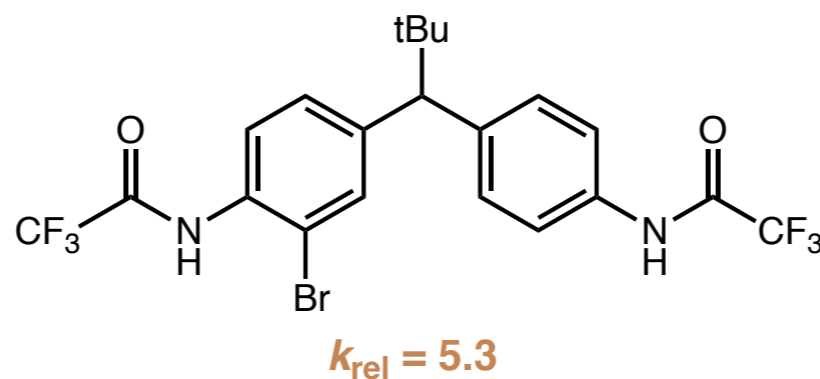
substrate recognition by non-covalent interactions

Copper-Peptide-Mediated Cross-Coupling of Diarylmethanes

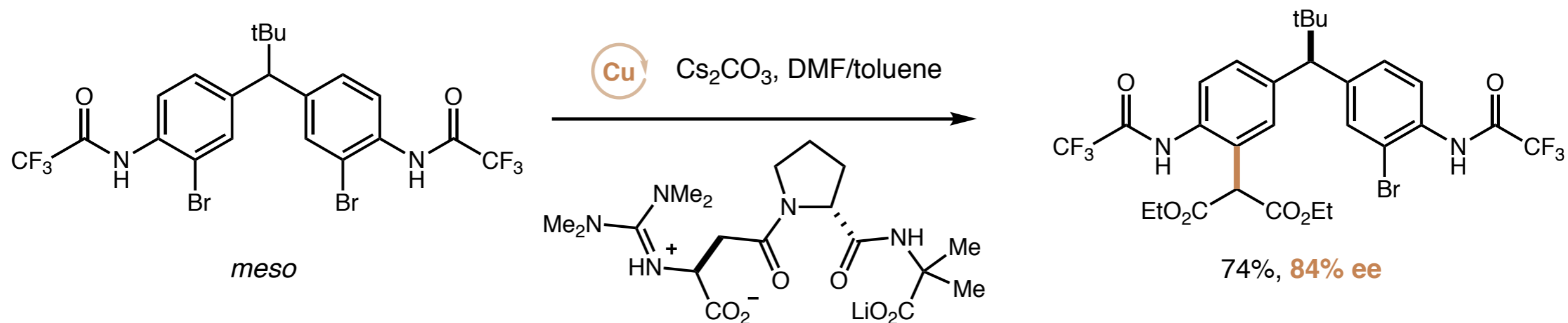


identification of the substrate-peptide interaction

A high k_{rel} in the kinetic resolution of unsymmetrical substrates indicates a substrate-peptide interaction

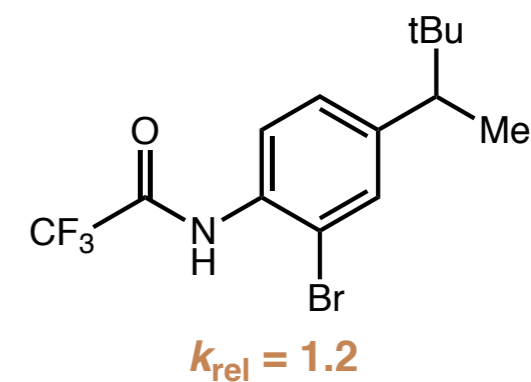
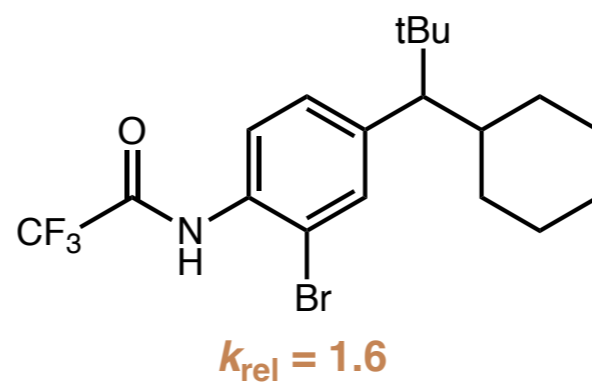
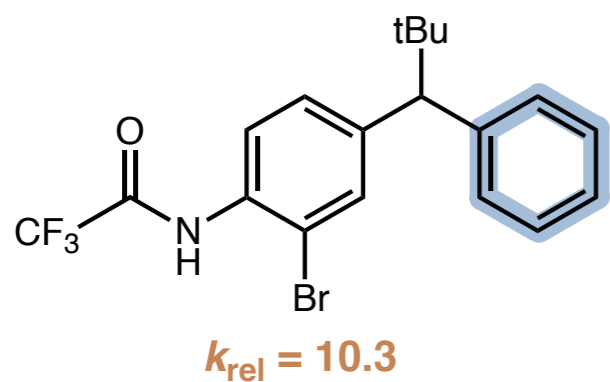
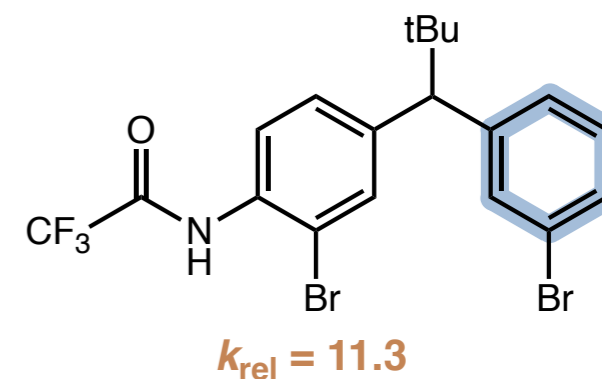
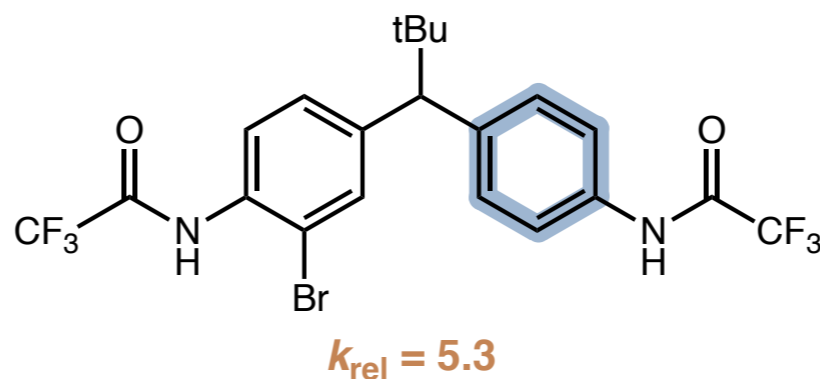


Copper-Peptide-Mediated Cross-Coupling of Diarylmethanes

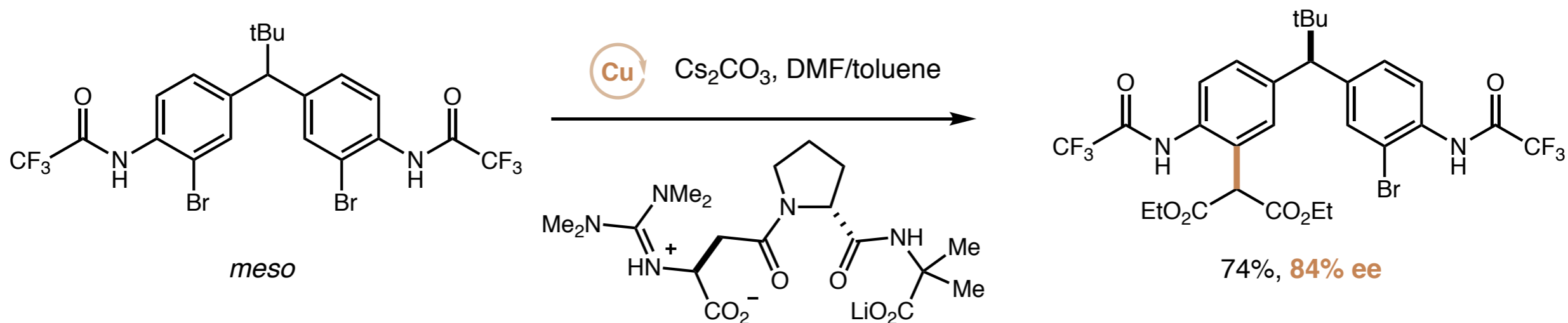


identification of the substrate-peptide interaction

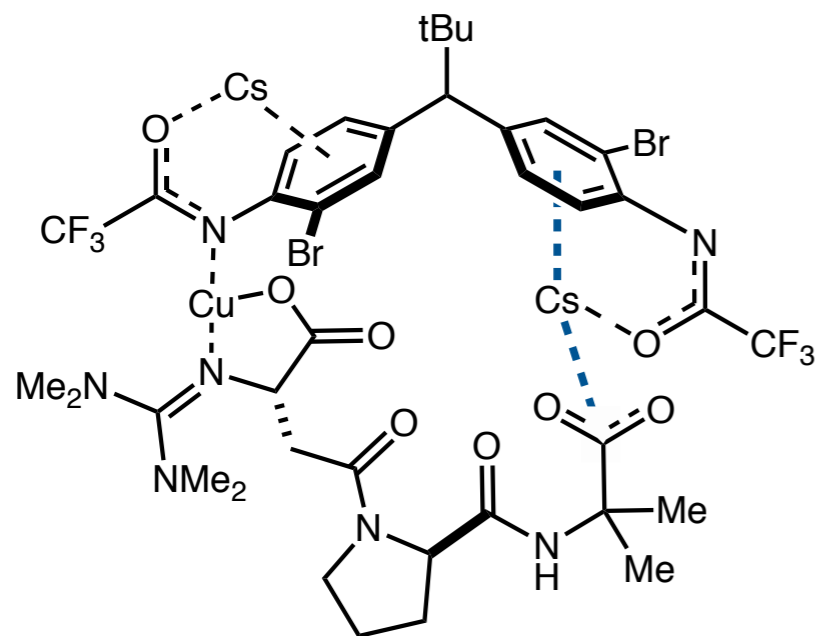
A high k_{rel} in the kinetic resolution of unsymmetrical substrates indicates a substrate-peptide interaction



Copper-Peptide-Mediated Cross-Coupling of Diarylmethanes



identification of the substrate-peptide interaction



Proposed model

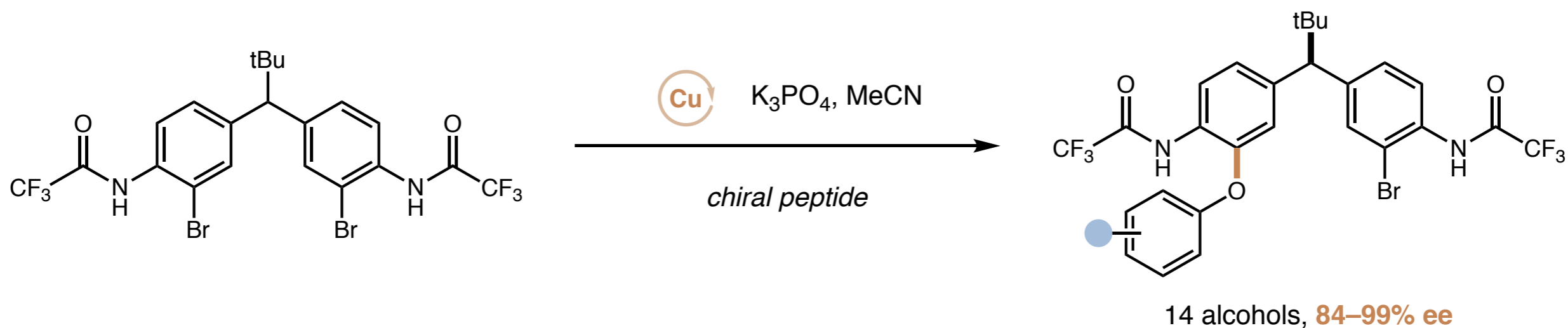
cation- π -interaction
between substrate
and ligated cesium

Role of the peptide

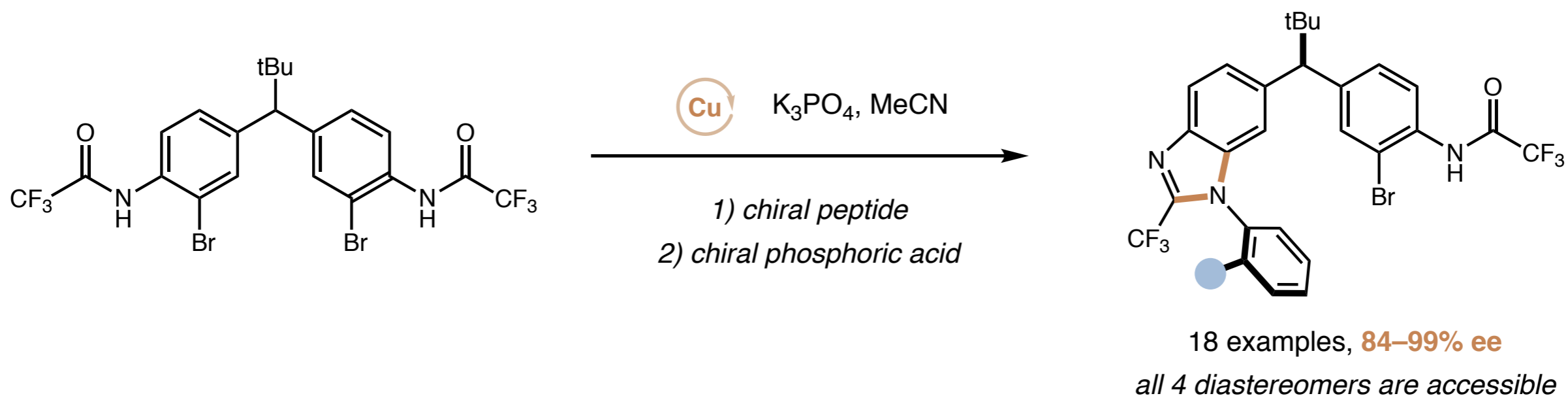
substrate recognition
by non-covalent
interaction

Copper-Peptide-Mediated Cross-Coupling of Diarylmethanes

■ C–O coupling



■ C–N coupling and catalyst-controlled cyclodehydration



Kwon, Y.; Chinn, A. J.; Kim, B.; Miller, S. J. *J. Am. Chem. Soc.* **2018**, *140*, 6251.

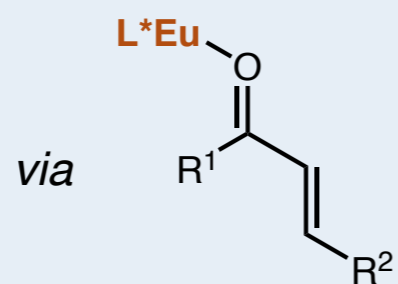
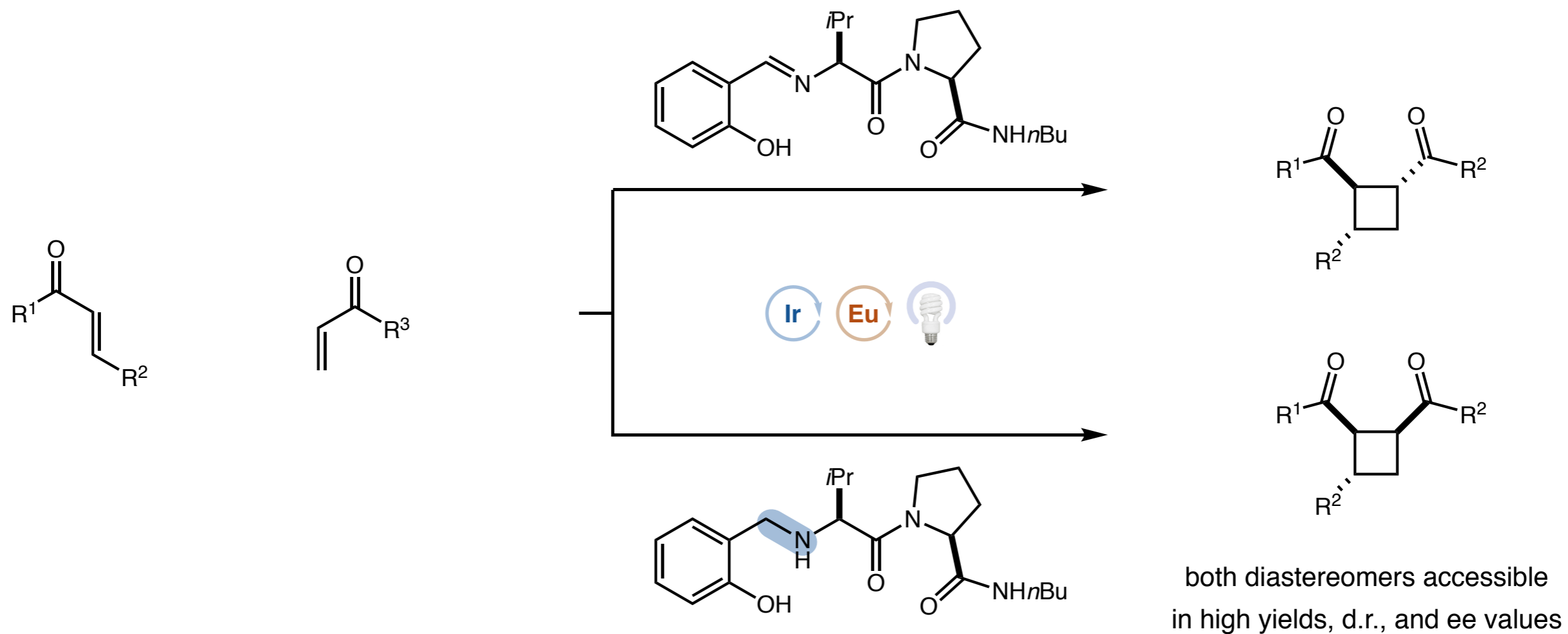
Chinn, A. J.; Kim, B.; Kwon, Y.; Miller, S. J. *J. Am. Chem. Soc.* **2017**, *139*, 18107.

Small Peptides as Organocatalysts

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- Summary

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Enantioselective [2+2] Photocycloaddition

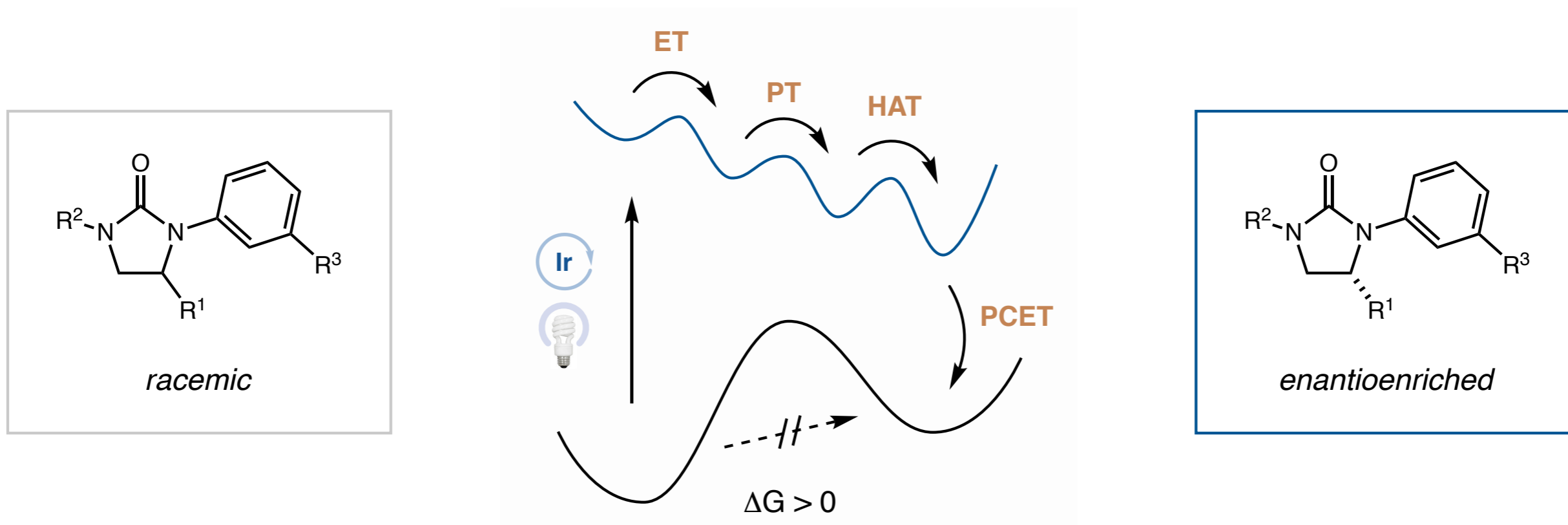


Role of the peptide

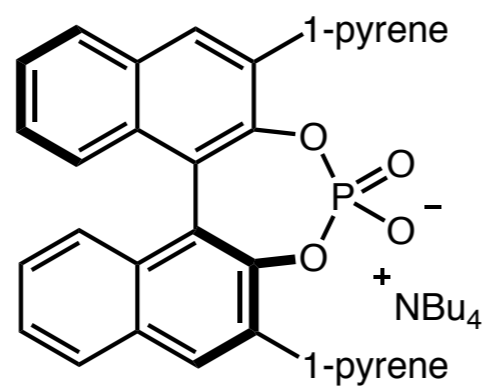
creation of a chiral environment around the metal catalyst

Light-Driven Deracemization

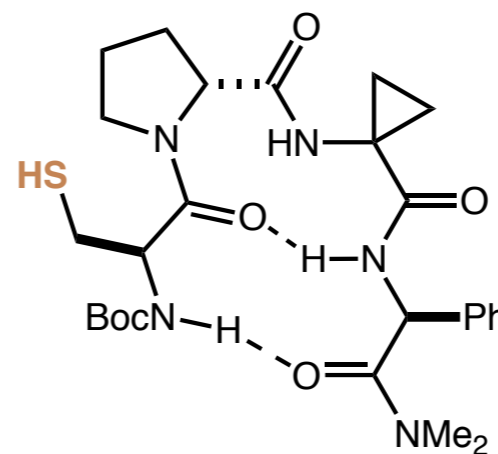
- Photoredox strategy for an out-of-equilibrium deracemization



two chiral
co-catalysts



chiral phosphate

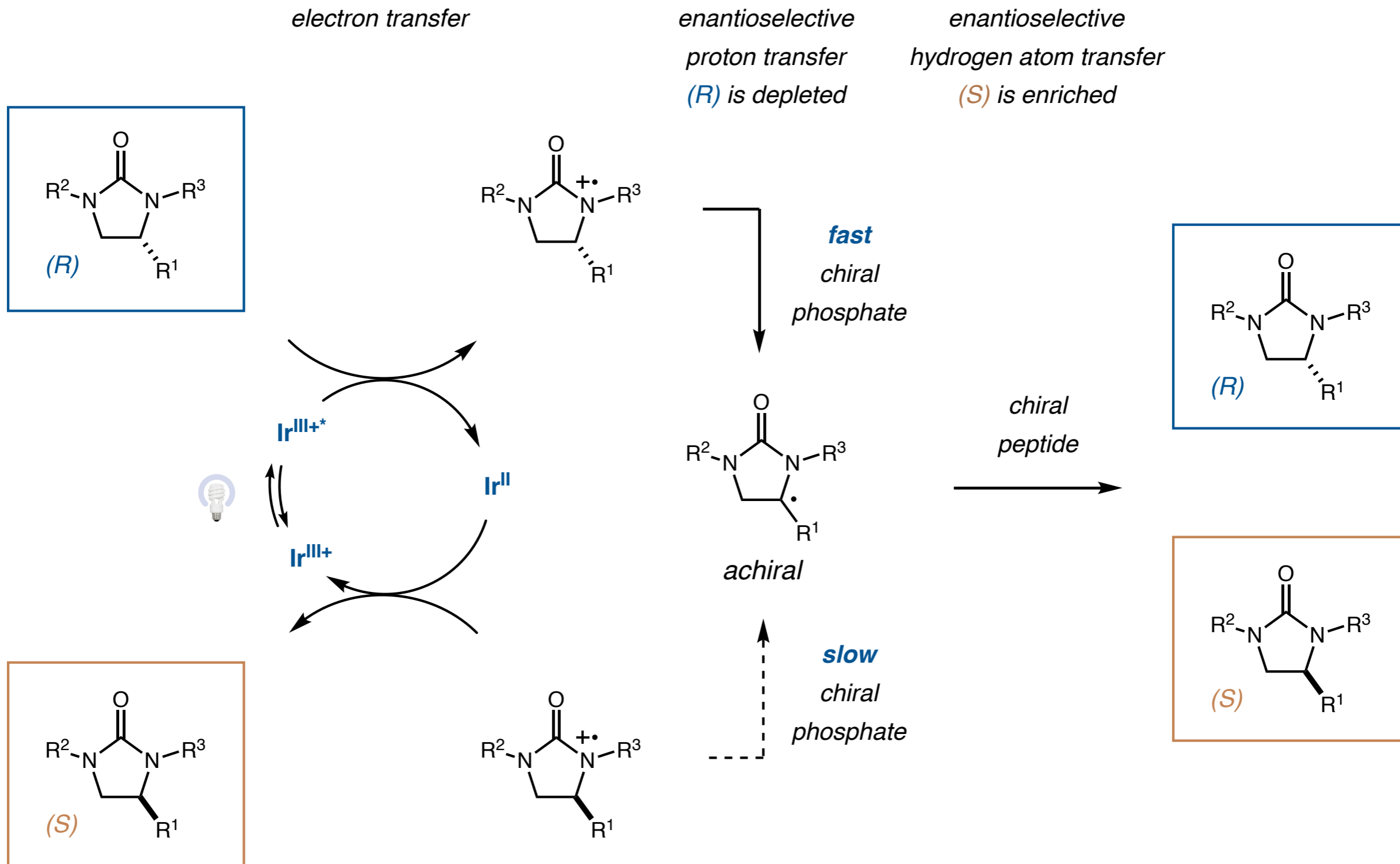


chiral peptide

Role of the peptide

chiral hydrogen
atom donor

Light-Driven Deracemization



Summary

- A diverse set of reactivity can be achieved using organocatalytic, organometallic, or merged activation modes.
- Peptides may control selectivity in catalytic transformations using non-covalent interactions.
- Control of the secondary structure is vital in order to place reactive groups in close proximity.
- A variety of screening technologies may assist in the discovery process.

Thank you for your attention.

Questions?