

The Living Cell



Living cells are a complex network of interacting biopolymers, ions, and metabolites.

Complex cellular processes cannot be observed when the components are in their purified, isolated forms.

Thus, we are forced outside of the artificial confines of a test tube and into the dynamic living cell.

There is a burgeoning interest in developing methods where a reporter tag can be attached to a cellular component for aid in visualization and/or isolation.

Prescher, J. A.; Bertozzi, C. R. *Nat. Chem. Biol.* **2005**, *1*, 13-21.

David Goodshell, artist and biologist

Bioconjugation Chemistry

Bioconjugation chemistry - a selective coupling of two biomolecules together in a covalent linkage

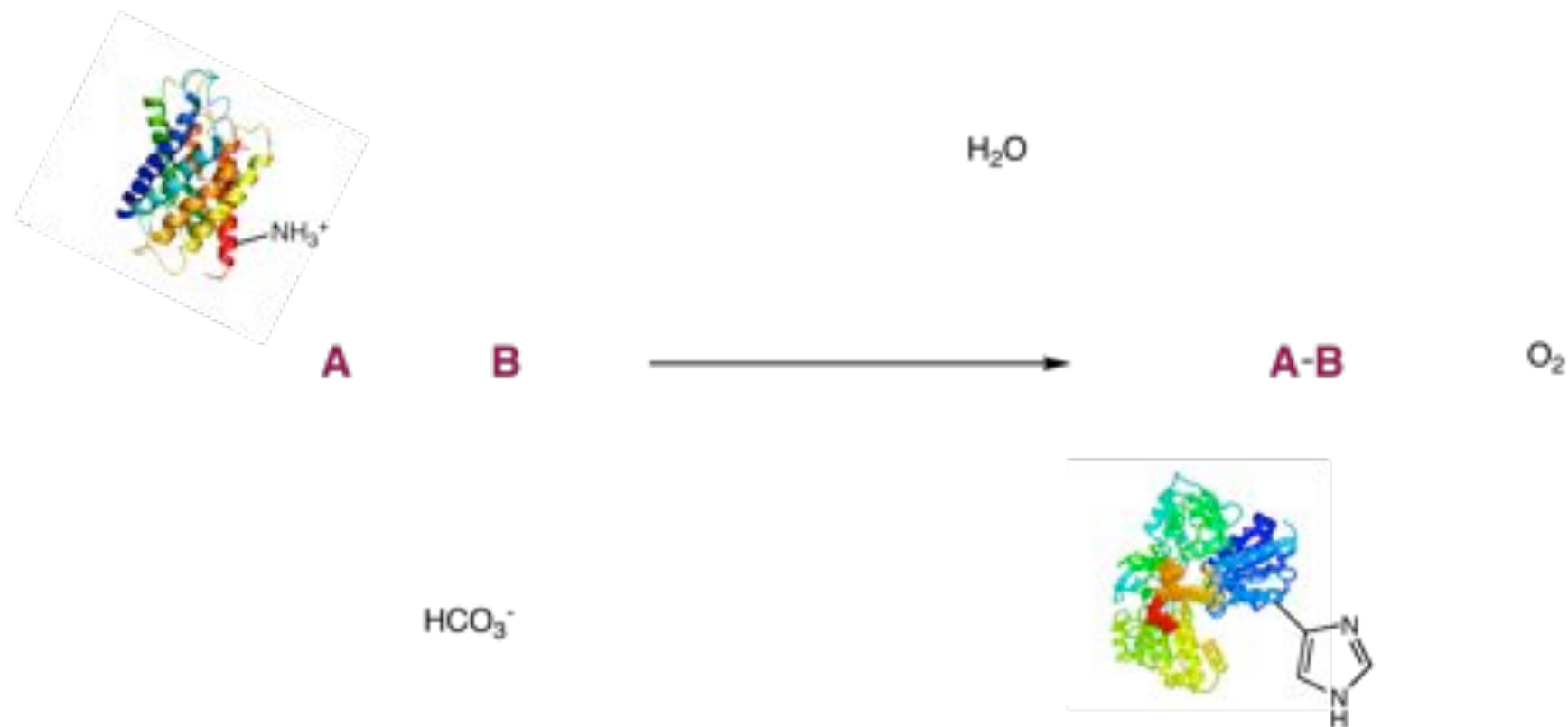


Antos, J. M.; McFarland, J. M.; Iavarone, A. T.; Francis, M. B. *J. Am. Chem. Soc.* **2009**, *131*, 6301-6308.

Sletten, E. M.; Bertozzi, C. R. *Angew. Chem. Int. Ed.* **2009**, *48*, 6974-6998.

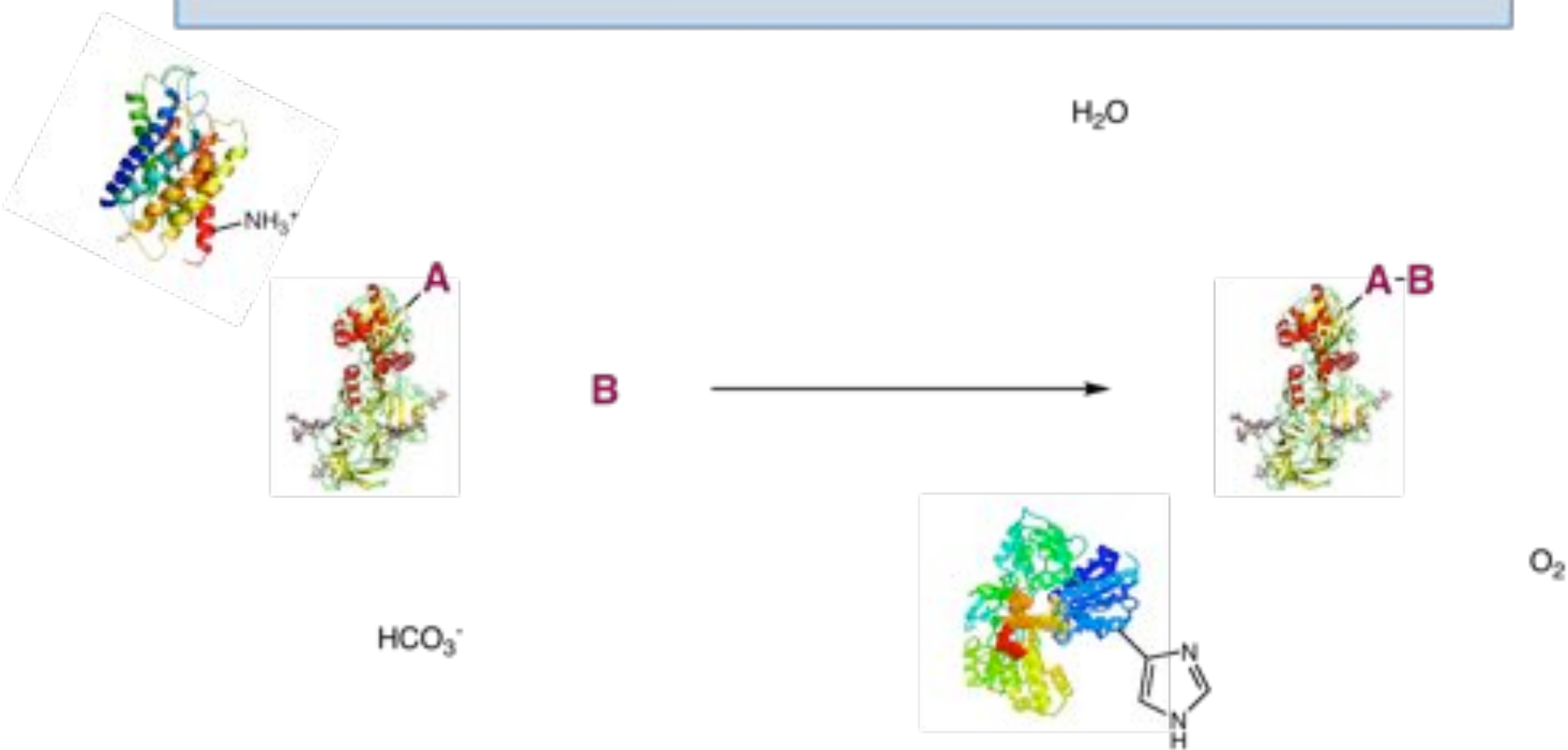
Bioorthogonal Chemistry

Bioorthogonal chemistry - a chemical reaction that can occur inside living systems without interfering with native biochemical processes



Bioorthogonal Chemistry

Bioorthogonal chemistry - a chemical reaction that can occur inside living systems without interfering with native biochemical processes



Bioorthogonal Chemistry - Requirements

water as the solvent

ambient temperature

non-toxic reagents

physiological pH

reaction product must be stable

no cross-reactivity

cell-permeable reagents

fast reaction kinetics

A Brief Consideration of Reaction Kinetics



most bioorthogonal reactions are bimolecular
in nature with second-order rate constants

[product] is approximately $k_2[A]_0[B]_0t$ k_2 is typically anywhere from 10^{-3} to $10^3 \text{ M}^{-1}\text{s}^{-1}$

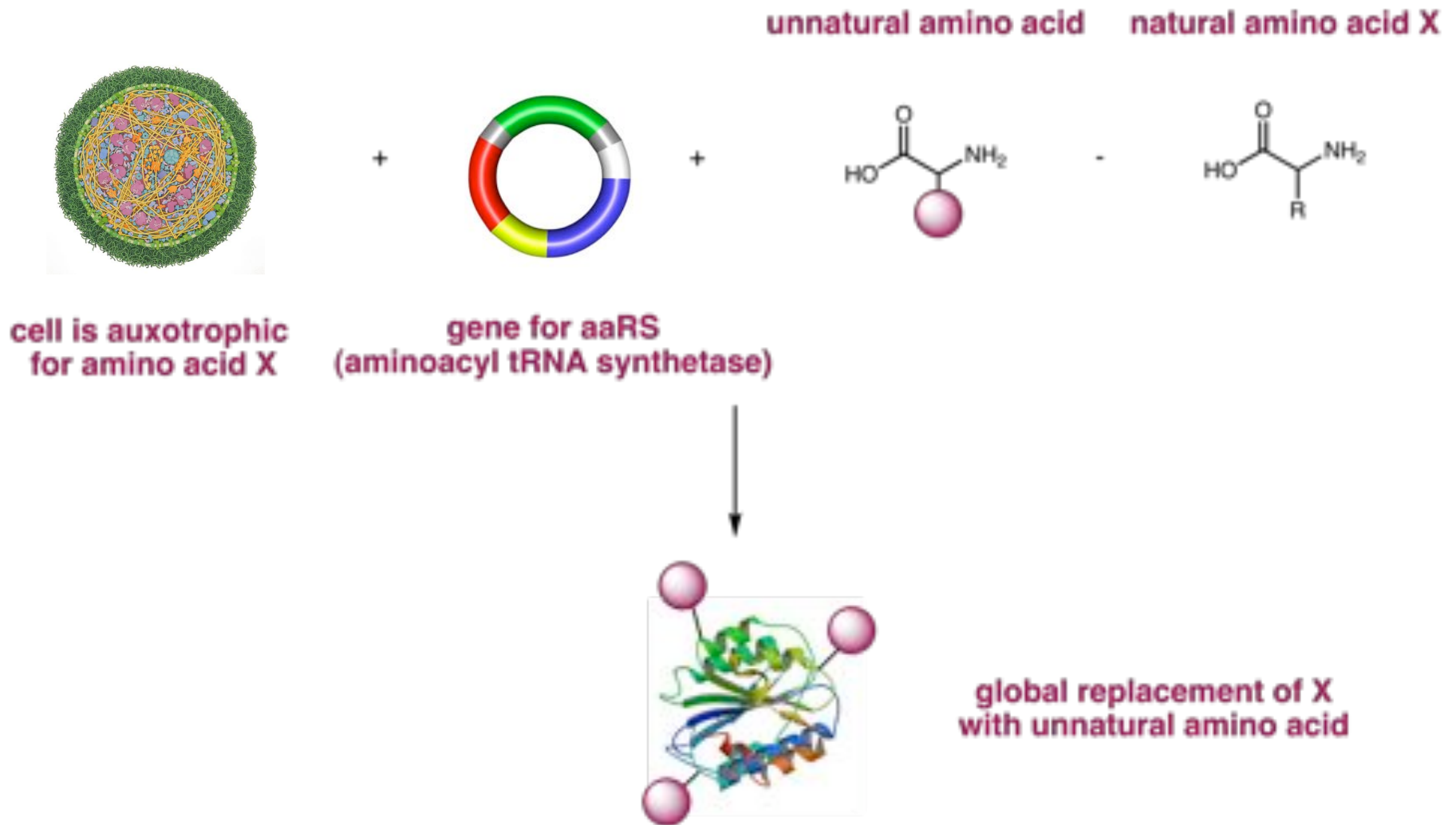
$$k_2 = 2.3 \text{ M}^{-1}\text{s}^{-1}, [A] = [B] = 1 \text{ } \mu\text{M}, \text{ time} = 1 \text{ hr}$$

~ 0.8 % labeling of product

Sletten, E. M.; Bertozzi, C. R. *Angew. Chem. Int. Ed.* **2009**, *48*, 6974-6998.

Lim, R. K. V.; Lin, Q. *Chem. Commun.* **2010**, *46*, 1589-1600.

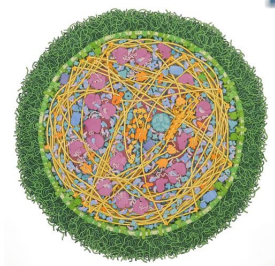
Incorporation of Unnatural Amino Acids



Link, A. J.; Mock, D. A.; Tirrell, D. A. *Curr. Opin. Biotechnol.* **2003**, *14*, 603.

Cowie, D. B.; Cohen, G. N. *Biochim. Biophys. Acta* **1957**, *26*, 252.

Incorporation of Unnatural Amino Acids



cell



gene for mutant aaRS



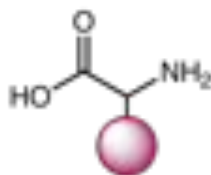
gene for tRNA



gene for protein



site-specific incorporation
of unnatural amino acid



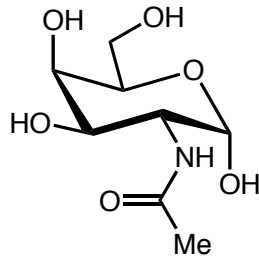
unnatural amino acid

Wang, L.; Schultz, P. G. *Angew. Chem. Int. Ed.* **2005**, *44*, 43.

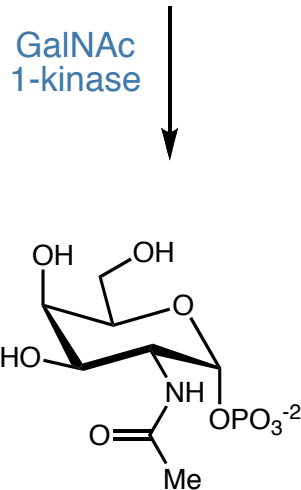
Noren, C. J.; Anthony-Cahill, S. J.; Griffith, M. C.; Schultz, P. G. *Science* **1989**, *244*, 182.

Bain, J. D.; Glabe, C. G.; Dix, T. A.; Chamberlin, A. R.; Diala, E. S. *J. Am. Chem. Soc.* **1989**, *111*, 8013.

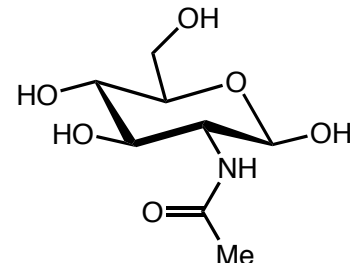
Biosynthesis of mucin-type O-linked glycoproteins



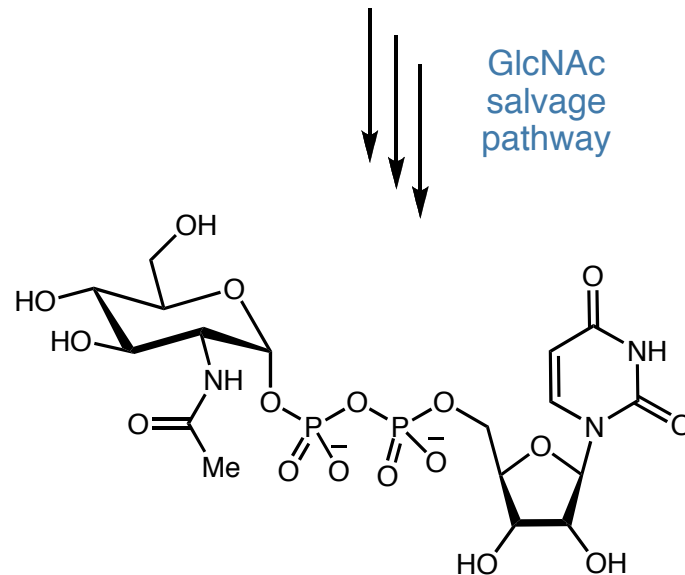
N-Acetylgalactosamine
GalNAc



GalNAc-1-P



N-Acetylglucosamine
GlcNAc

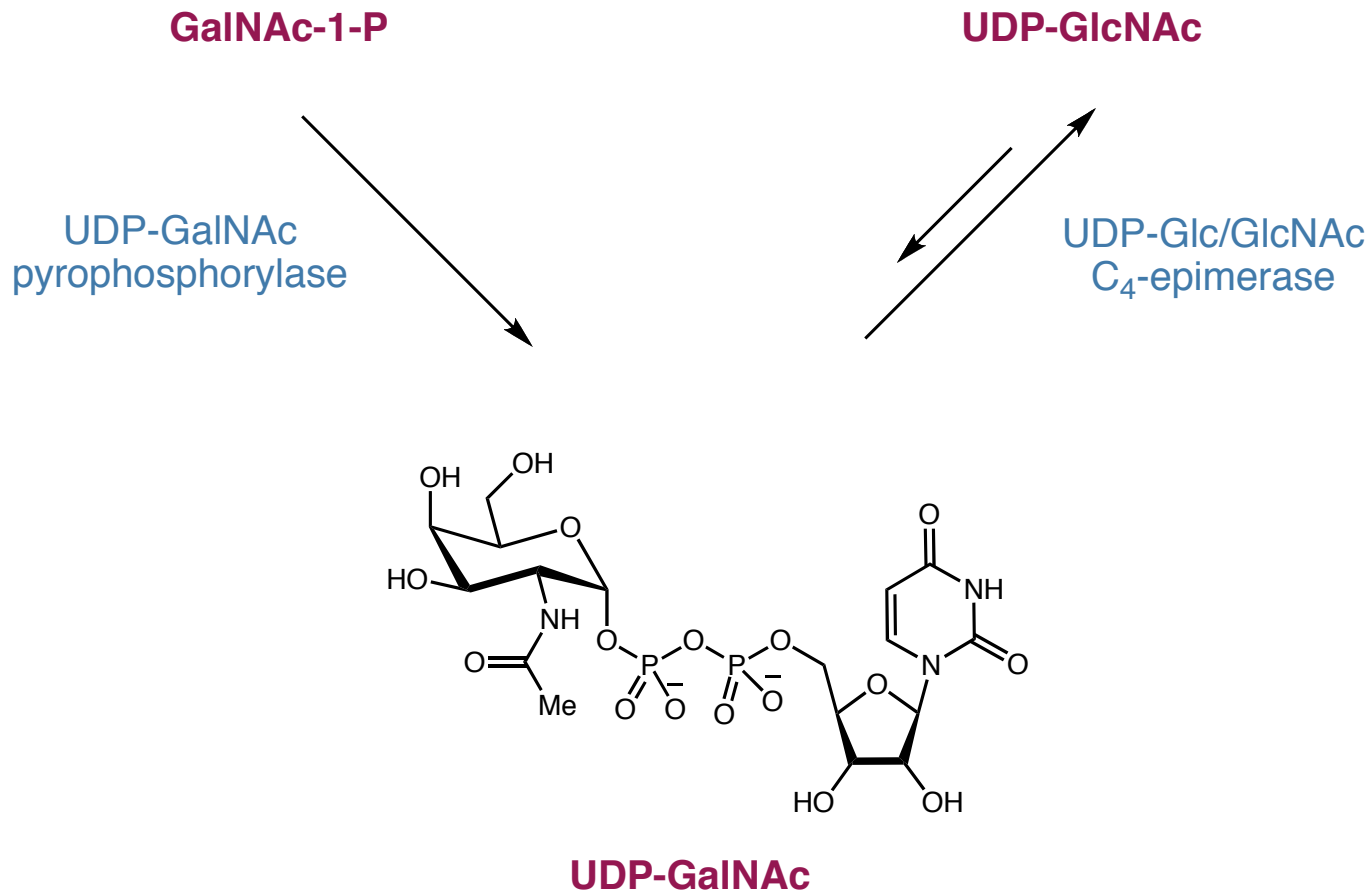


UDP-GlcNAc

Hang, H. C.; Yu, C.; Kato, D.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 14846-14851.

Dube, D. H.; Prescher, J. A.; Quang, C. N.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 4819-4824.

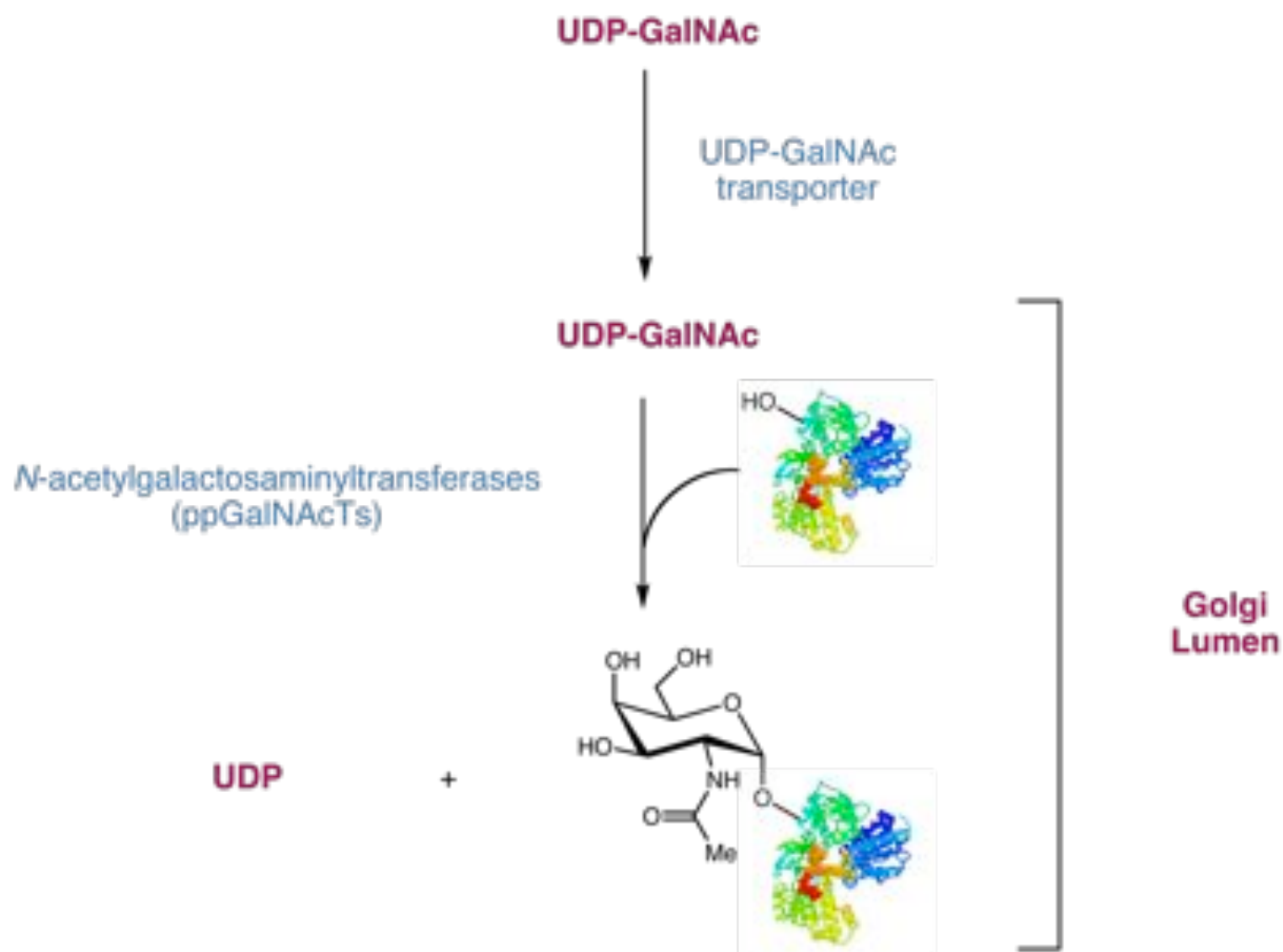
Biosynthesis of mucin-type O-linked glycoproteins



Hang, H. C.; Yu, C.; Kato, D.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 14846-14851.

Dube, D. H.; Prescher, J. A.; Quang, C. N.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 4819-4824.

Biosynthesis of mucin-type O-linked glycoproteins

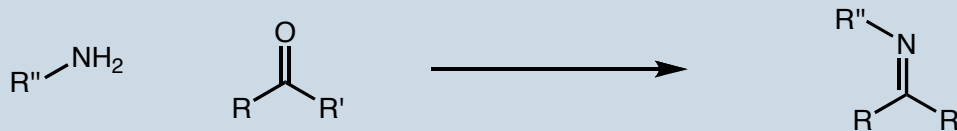


Hang, H. C.; Yu, C.; Kato, D.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 14846-14851.

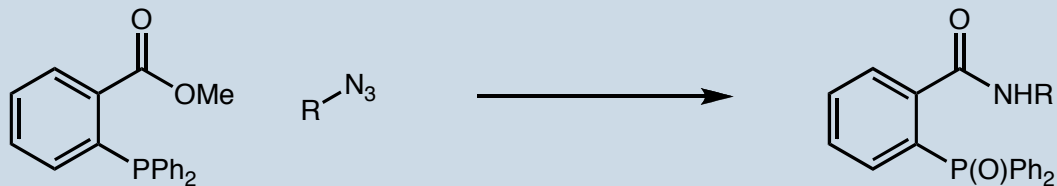
Dube, D. H.; Prescher, J. A.; Quang, C. N.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 4819-4824.

Bioorthogonal Chemistry

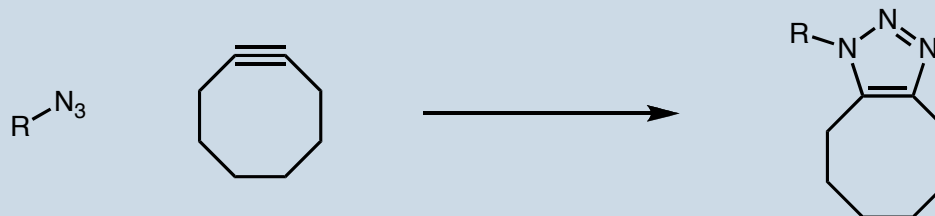
condensation
chemistry



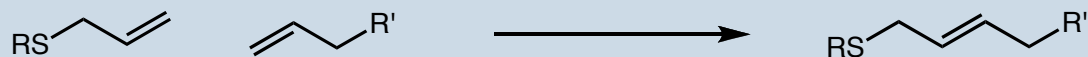
Staudinger
ligation



[3+2] cycloadditions

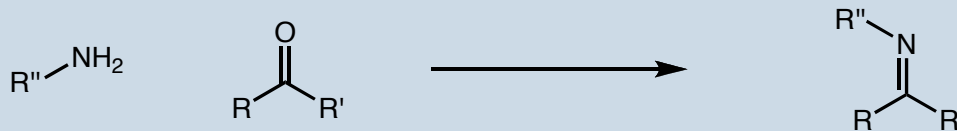


olefin chemistry

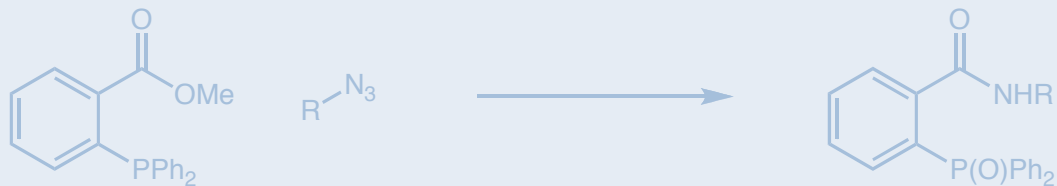


Bioorthogonal Chemistry

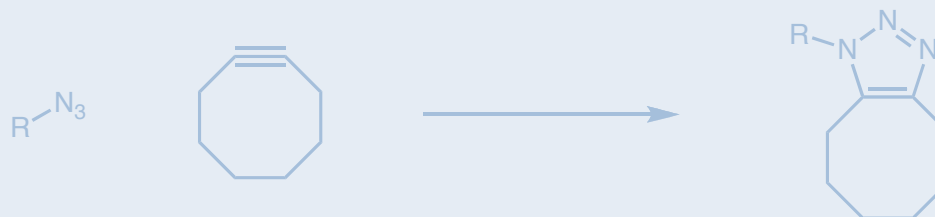
condensation
chemistry



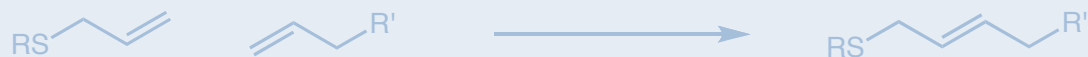
Staudinger
ligation



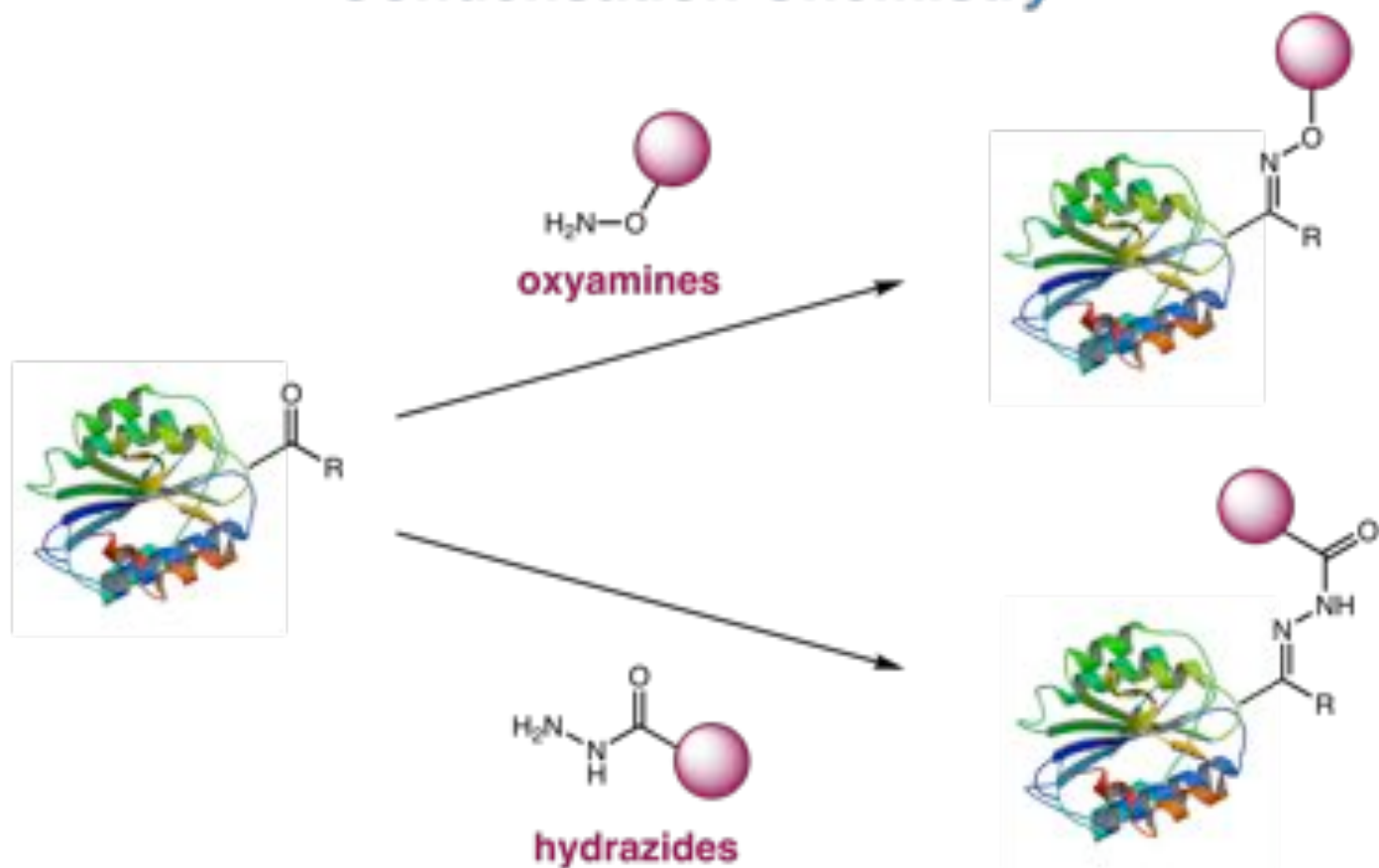
[3+2] cycloadditions



olefin chemistry

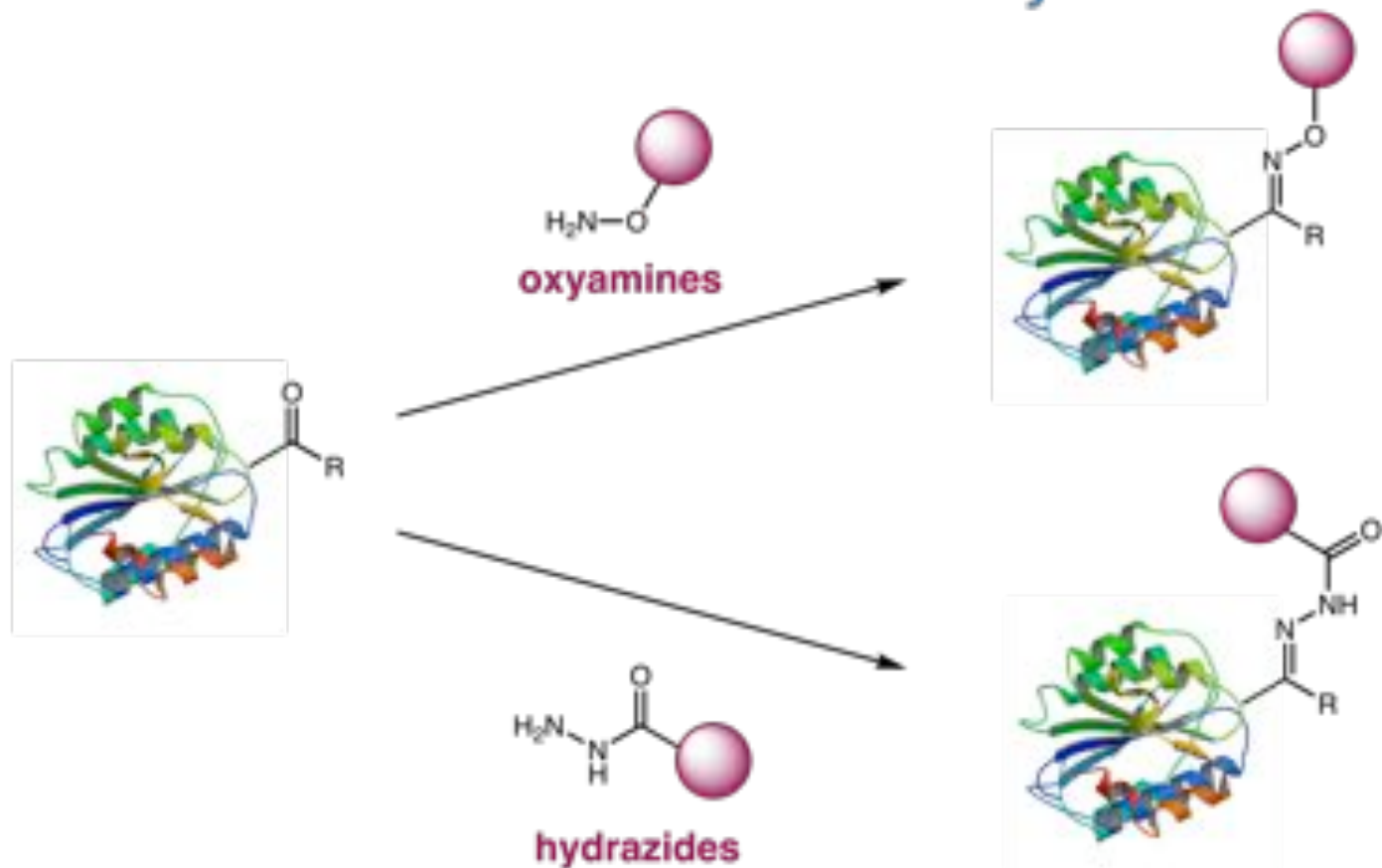


Condensation Chemistry



hydrazides and oxyamines are commonly used for this condensation chemistry
biological nucleophiles can condense but the equilibrium favors the starting material

Condensation Chemistry



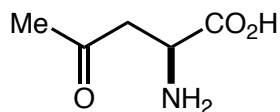
this chemistry is not widely employed inside cells or within live organisms

is applicable to cell surfaces for aldehydes and ketones are not present there

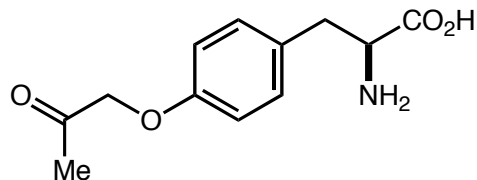
Condensation Chemistry

- Unnatural amino acid mutagenesis was performed on T4 lysozyme

incorporated at two solvent accessible sites - Ser⁴⁴ and Ala⁸²



5% efficiency



30% efficiency

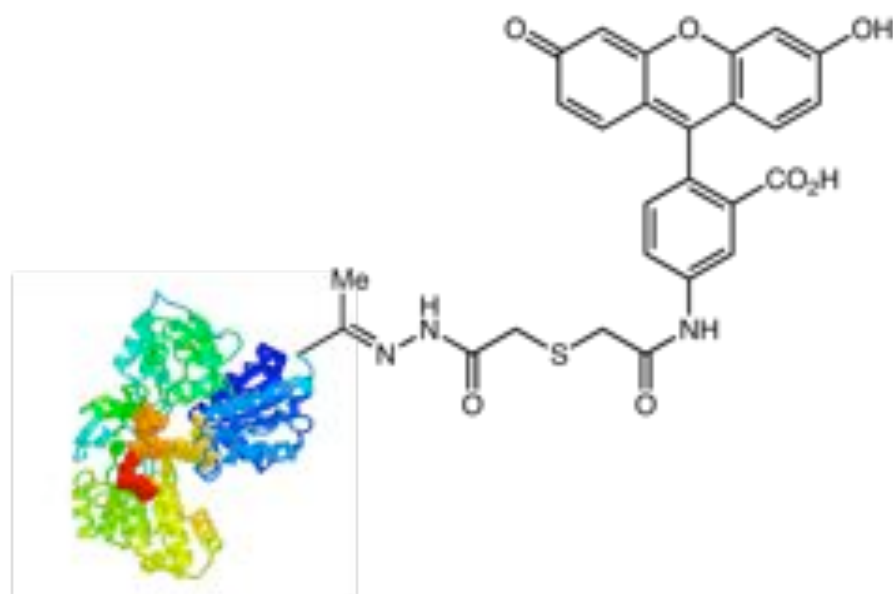
determined by catalytic activity, SDS-PAGE, and autoradiography

Cornish, V. W.; Hahn, K. M.; Schultz, P. G. *J. Am. Chem. Soc.* **1996**, *118*, 8150-8151.

Zhang, Z.; Smith, B. A. C.; Wang, L.; Brock, A.; Cho, C.; Schultz, P. G. *Biochemistry* **2003**, *42*, 6735-6746.

Condensation Chemistry

Hydrazone Formation



condensation proceeds in roughly 50% efficiency

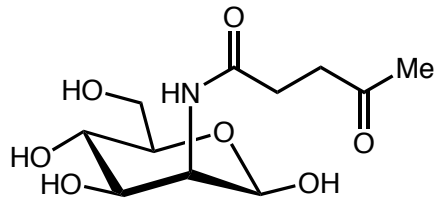
wild-type displays no fluorescence while the mutant does

Cornish, V. W.; Hahn, K. M.; Schultz, P. G. *J. Am. Chem. Soc.* **1996**, *118*, 8150-8151.

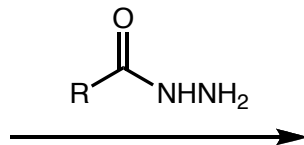
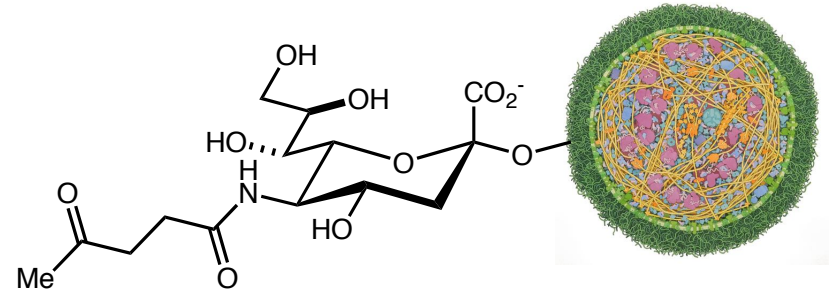
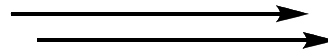
Zhang, Z.; Smith, B. A. C.; Wang, L.; Brock, A.; Cho, C.; Schultz, P. G. *Biochemistry* **2003**, *42*, 6735-6746.

Condensation Chemistry

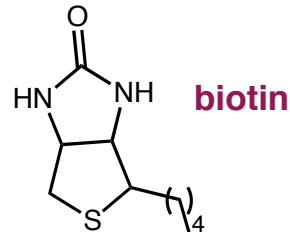
■ Cell surface remodeling



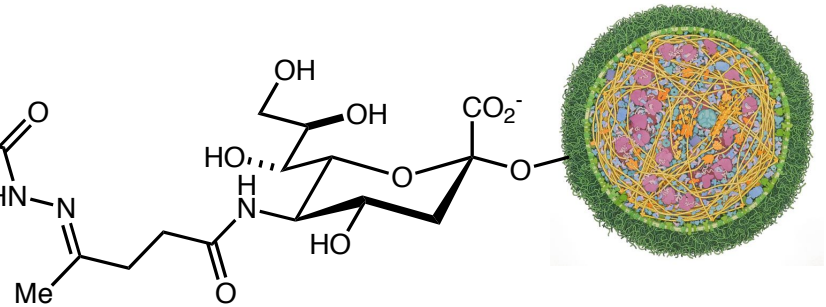
ManLev
a sialic acid analogue



Hydrazide



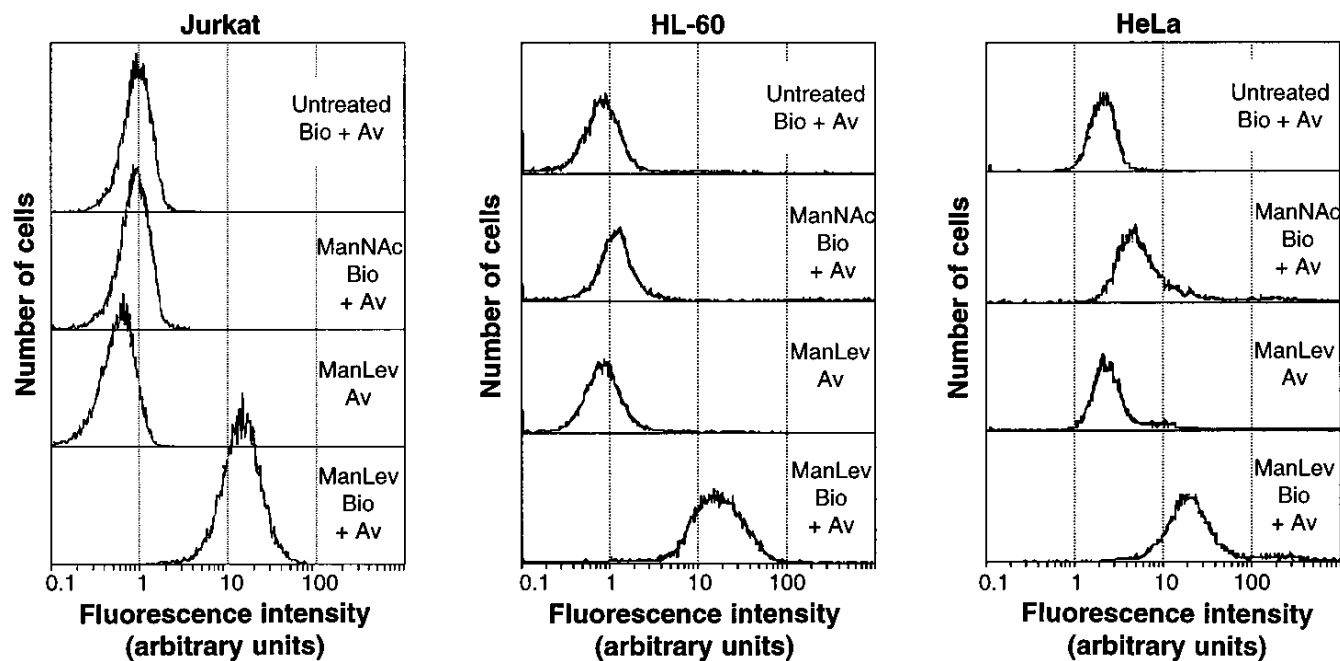
biotin



Keppler, O. T.; Horstkorte, R.; Pawlita, M.; Schmidts, C.; Reutter, W. *Glycobiology* **2001**, *11*, 11R- 18R.
Mahal, L. K.; Yarema, K. J.; Bertozzi, C. R. *Science* **1997**, *276*, 1125-1128.

Condensation Chemistry

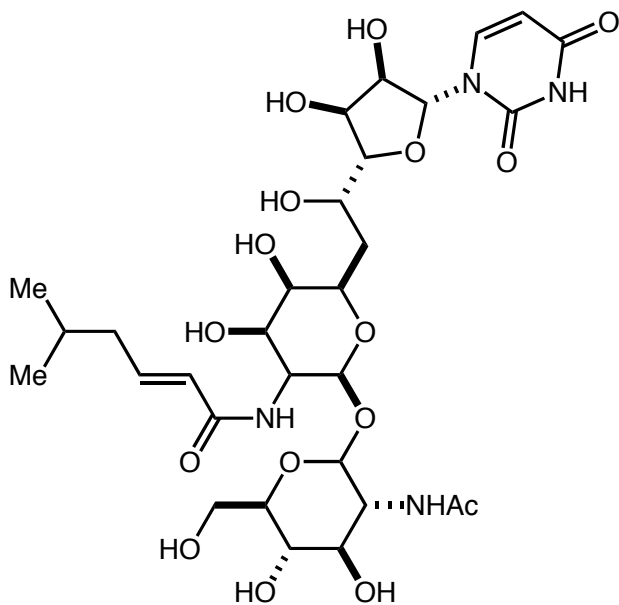
■ Cell surface remodeling



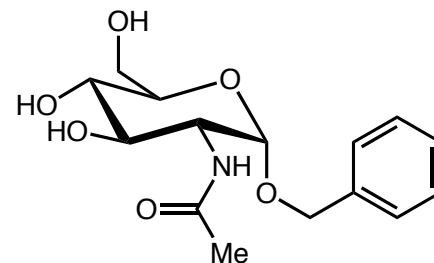
cells were analyzed by flow cytometry after staining with FITC (fluorescein isothiocyanate) avidin

Condensation Chemistry

tunicamycin



alpha-benzyl N-acetylglucosamine

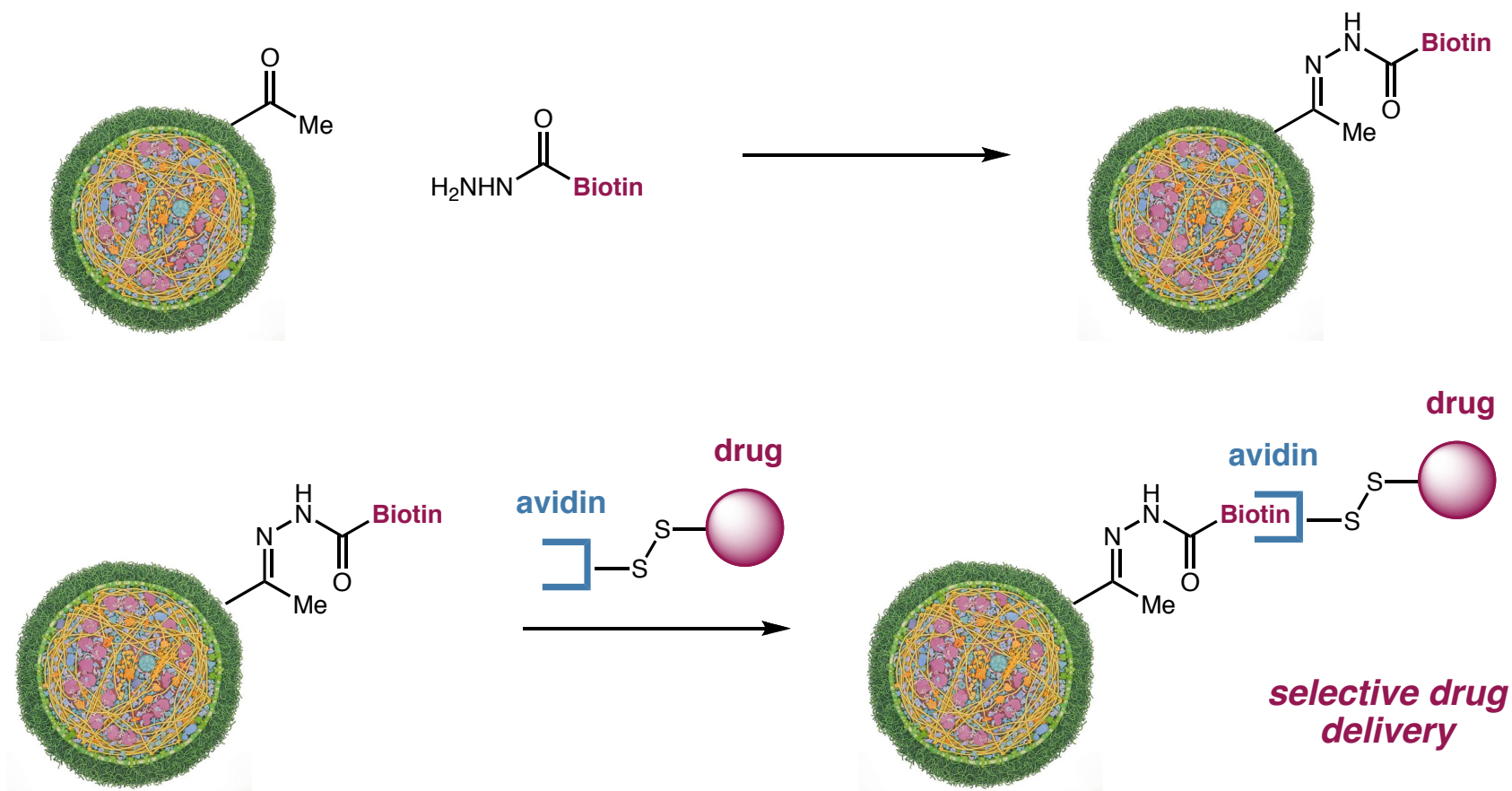


Addition of tunicamycin, a known inhibitor of N-linked protein glycosylation, inhibits ketone expression with ManLev treatment in Jurkat cells.

Ketone expression was blocked on HL-60 and HeLa cells via alpha-benzyl N-acetylgalactosamine, an inhibitor of O-linked glycosylation.

Condensation Chemistry

■ Selective drug delivery?



Condensation Chemistry

■ Selective drug delivery?



ricin
inhibits protein synthesis

toxicity of the conjugate was dependent on the expression of ketones

cells with high ketone expression (~700,000 ketones per cell) were sensitive to lethal doses of ricin

LD₅₀ between 1 to 10 nM

cells with low ketone expression (~50,000 ketones per cell) showed no toxicity

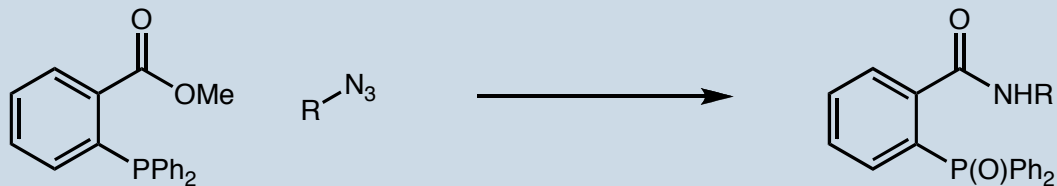
indicates the potential for cell surface engineering to support selective drug delivery

Bioorthogonal Chemistry

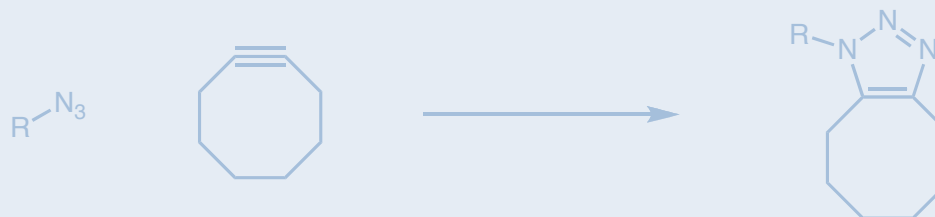
condensation
chemistry



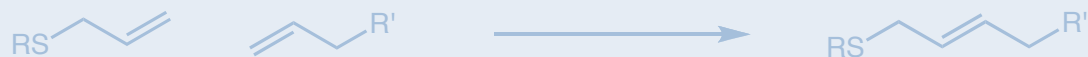
**Staudinger
ligation**



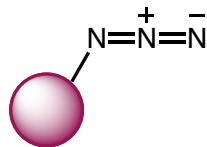
[3+2] cycloadditions



olefin chemistry



Azide - A Powerful Chemical Reporter



absent from biological systems

possesses orthogonal reactivity to most biological functional groups

the azide is small, so biological perturbation is minimal

first used as a chemical reporter in 2000 in the Staudinger Ligation

Griffin, R. J. *Prog. Med. Chem.* **1994**, 31, 121.

Hendricks, S. B.; Pauling, L. *J. Am. Chem. Soc.* **1925**, 47, 2904.

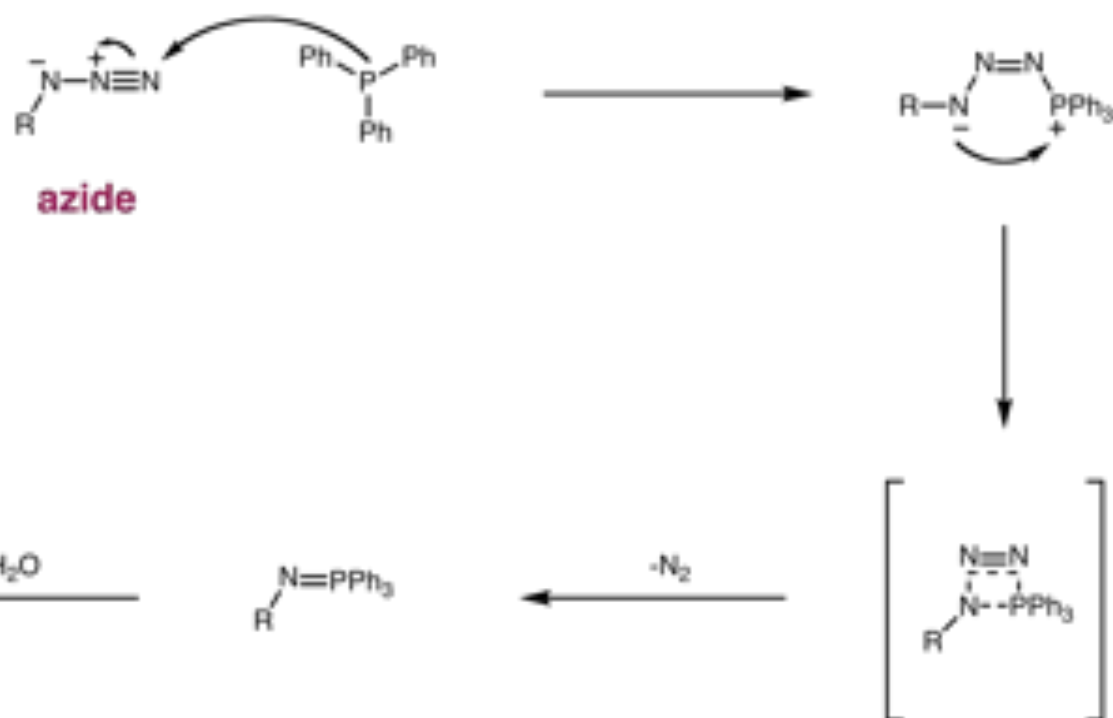
Saxon, E.; Bertozzi, C. R. *Science* **2000**, 287, 2007.

The Staudinger Reduction

- inspiration for the Staudinger ligation came from the Staudinger reduction



Hermann Staudinger

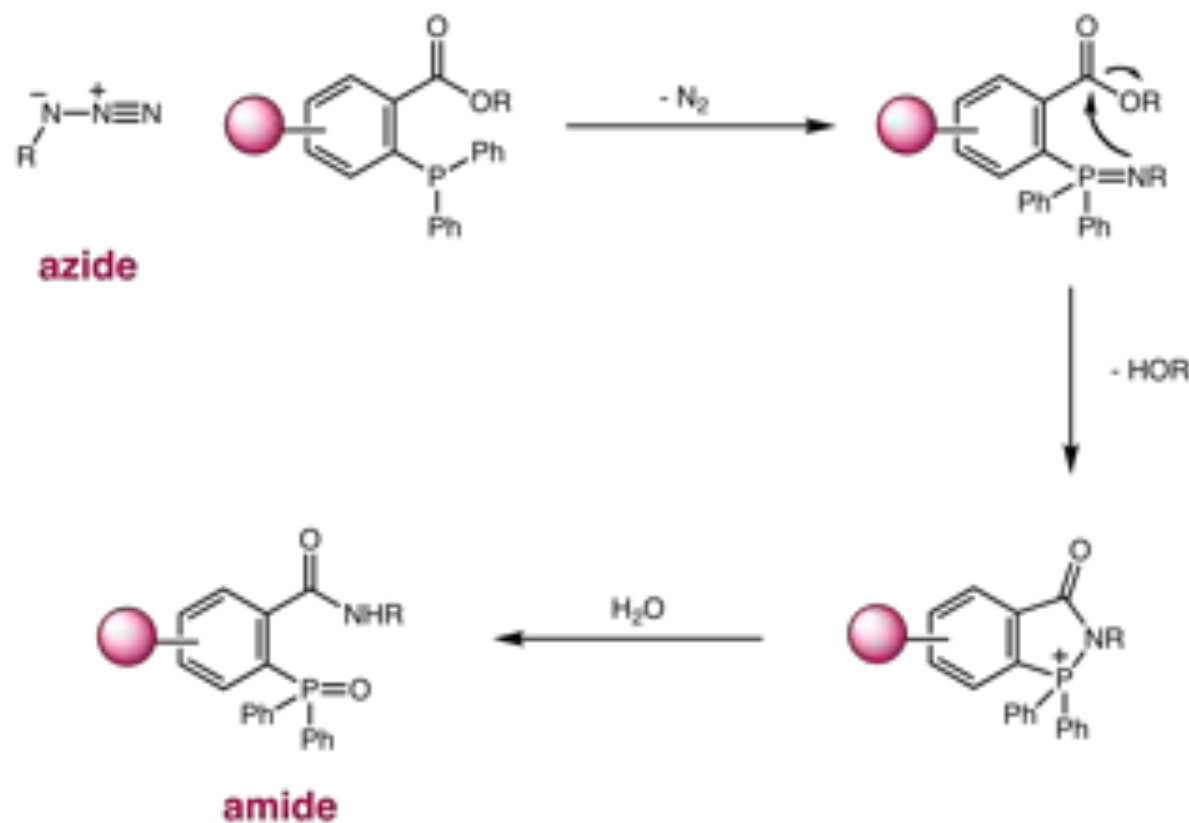


The Staudinger Ligation

- inspiration for the Staudinger ligation came from the Staudinger reduction



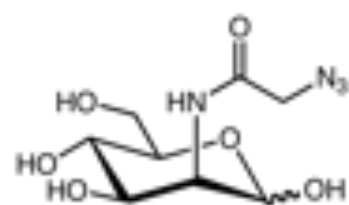
Carolyn Bertozzi



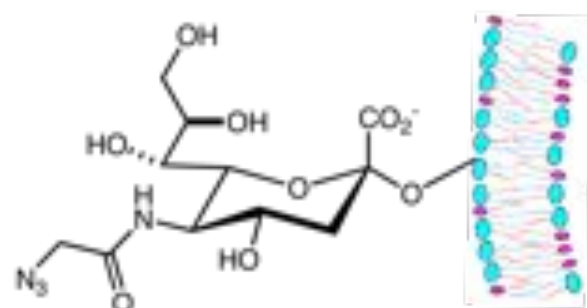
Saxon, E.; Bertozzi, C. R. *Science* **2000**, *287*, 2007.
Staudinger, H.; Meyer, J. *Helv. Chim. Acta.* **1919**, *2*, 635.

The Staudinger Ligation

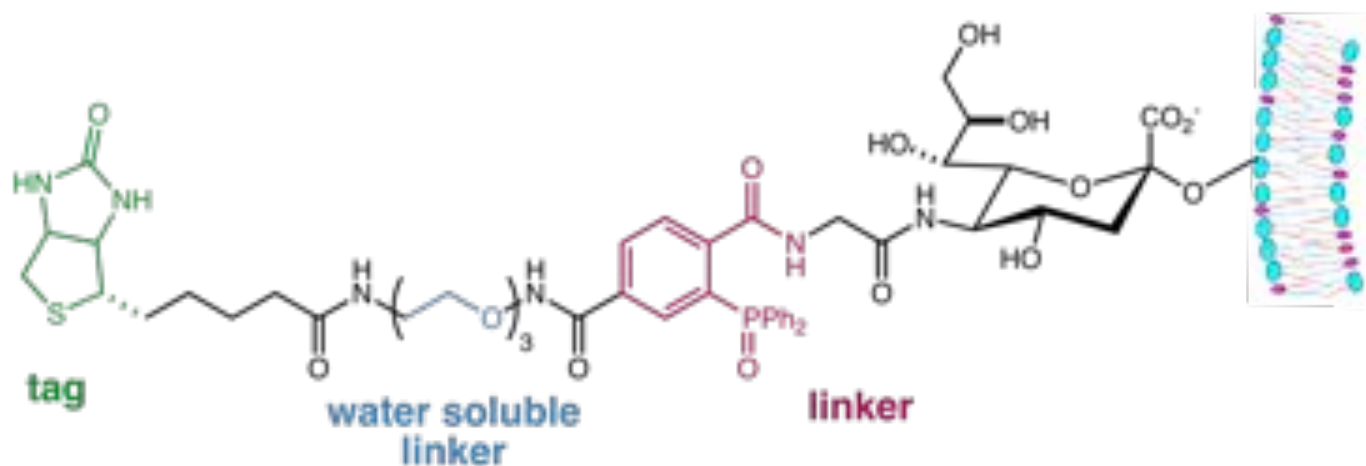
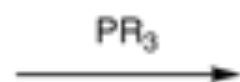
Cell surface engineering



azidosugar



sialic acid analogue



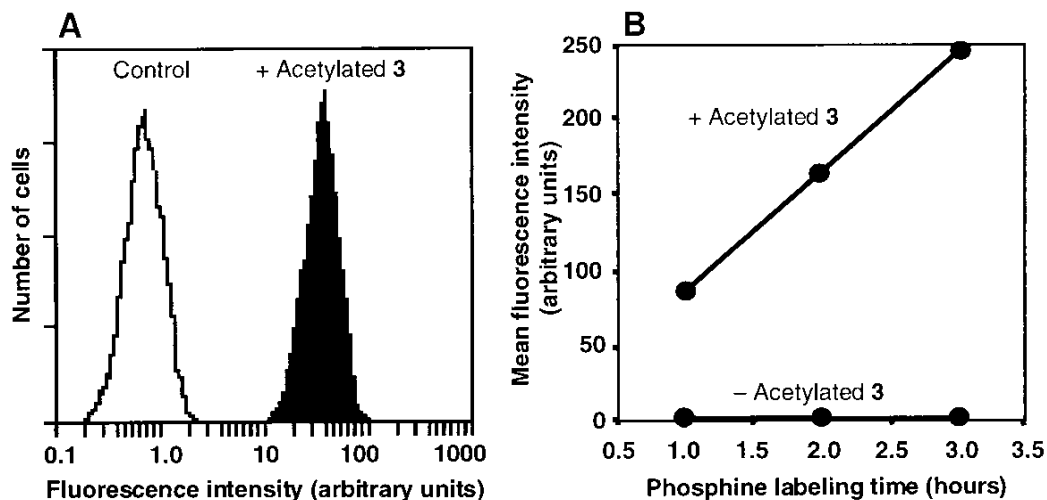
The Staudinger Ligation

Jurkat cells were incubated with azidoacetylmannosamine at a concentration 20 mM for three days.

Cell viability was tested with incubation for up to six days.

Cells were washed and reacted with phosphine for 1 hour at a concentration of 1 mM.

Stained with FITC-avidin and analyzed by flow cytometry.

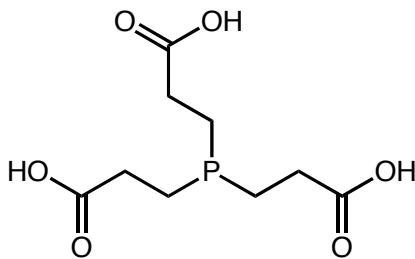


The Staudinger Ligation

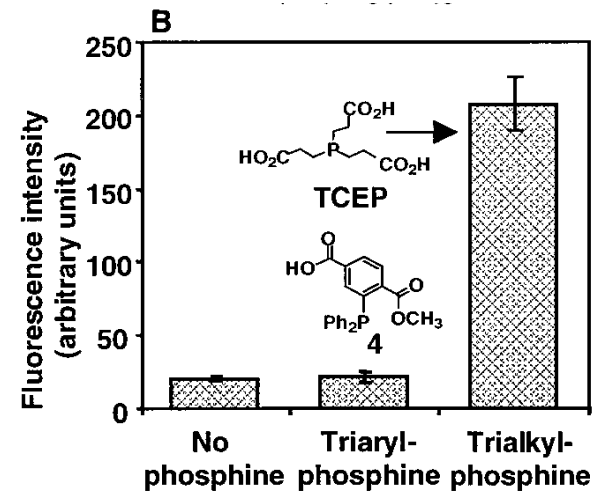
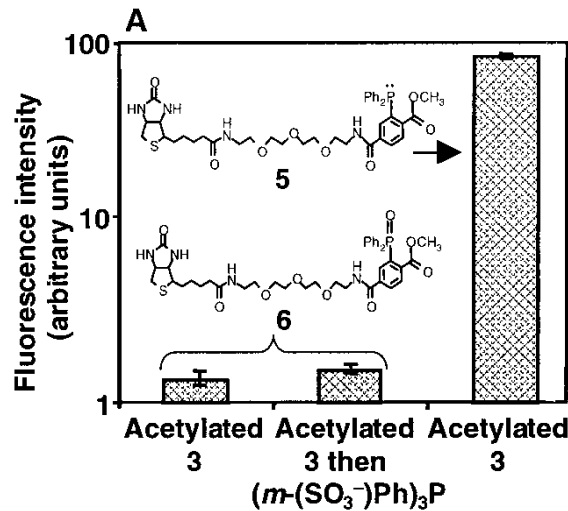
Biological Controls -

Could a Staudinger reduction be taking place and the phosphine oxide localizes in or outside the cell?

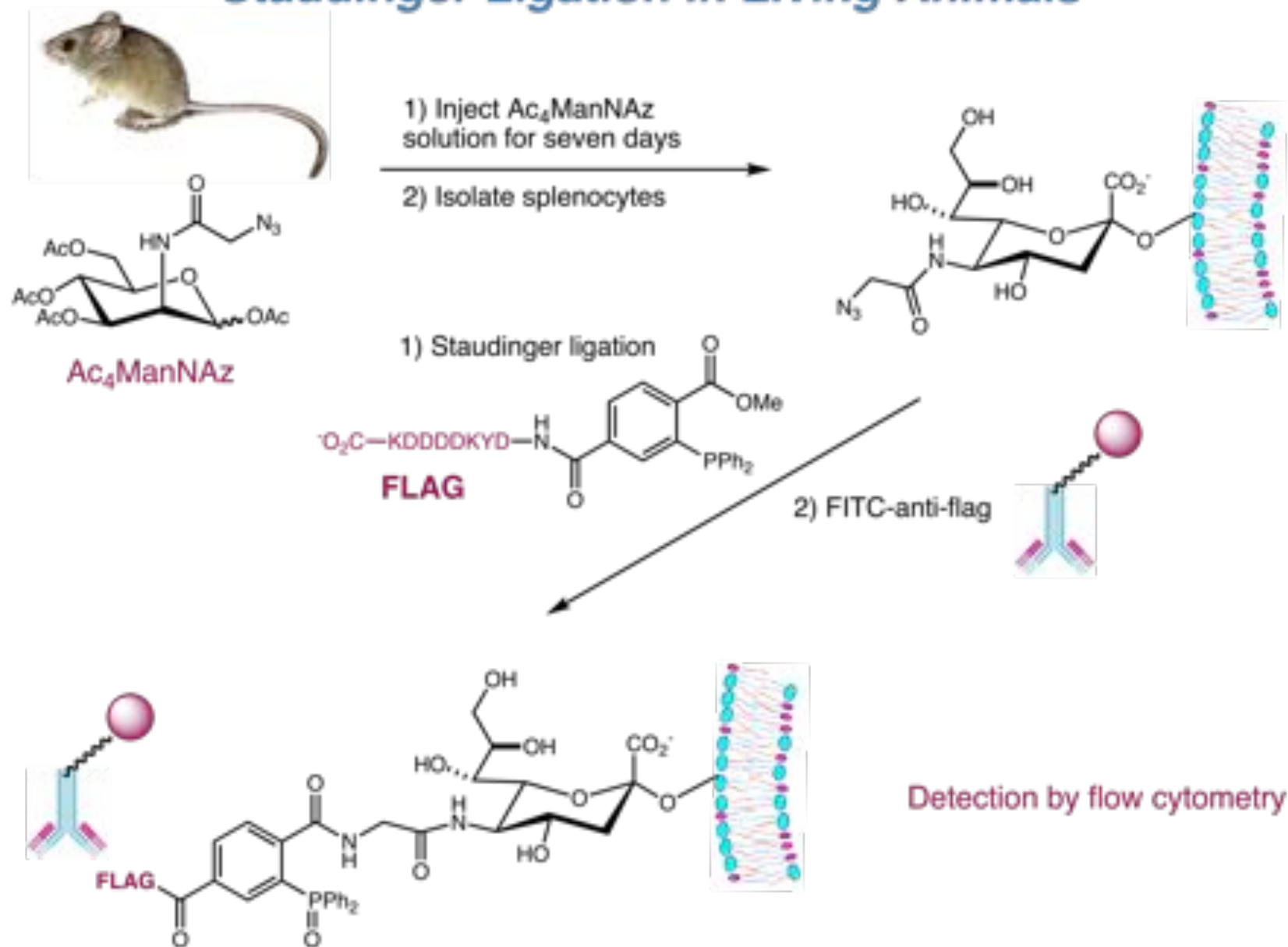
Could the phosphine be reducing disulfide bonds and not be completely bioorthogonal?



TCEP



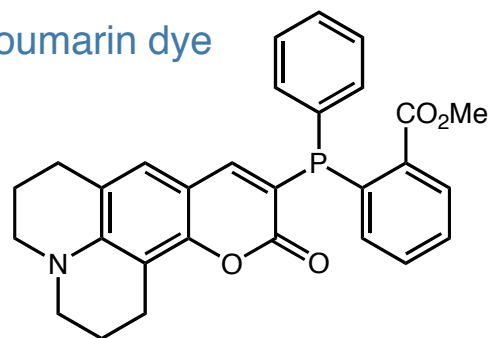
Staudinger Ligation in Living Animals



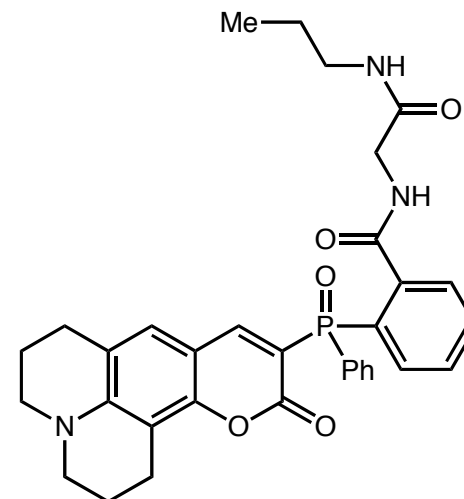
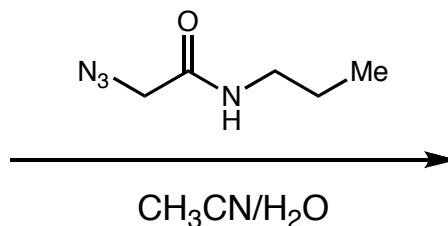
Activation of a Fluorogenic Dye via the Staudinger Ligation

The lone pair of electrons on phosphorus quench the excited fluorophore

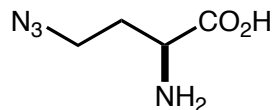
coumarin dye



not fluorescent



strongly fluorescent



L - azidohomoalanine

Phosphine dye was reacted with recombinant murine dihydrofolate reductase bearing azidohomoalanine residues.

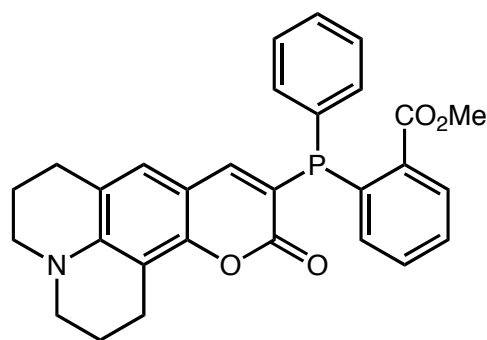
Unnatural amino acids incorporated during overexpression in a methionine auxotrophic *E. coli* strain.

Fluorescence was observed over background.

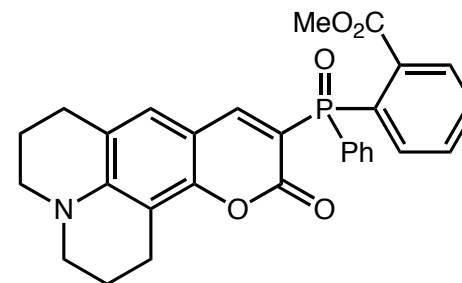
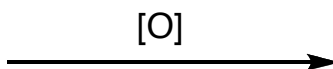
No washing or Western blotting necessary.

Activation of a Fluorogenic Dye via the Staudinger Ligation

A small problem



not fluorescent

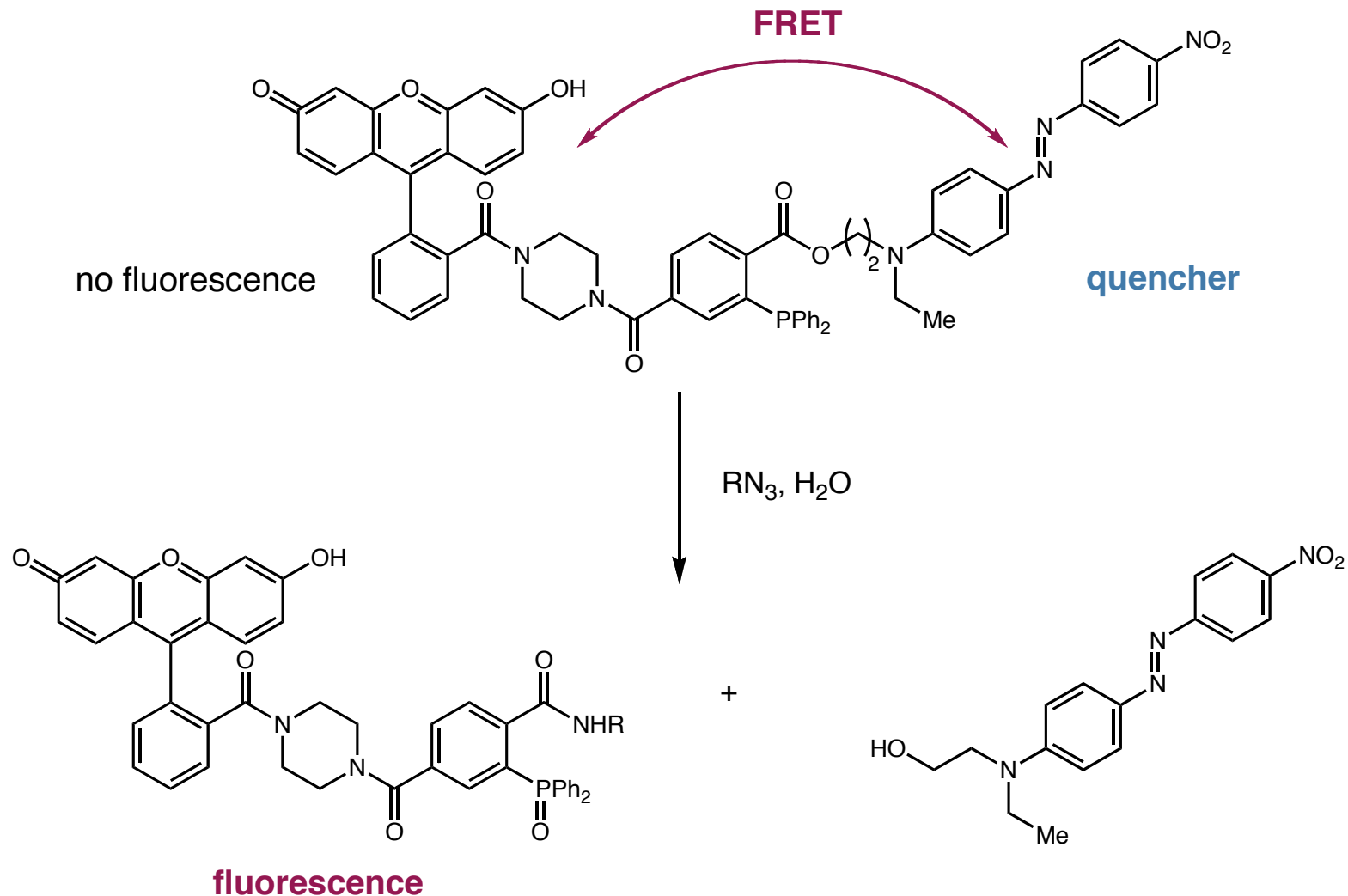


strongly fluorescent

Oxidation of the probe by air would provide background fluorescence.

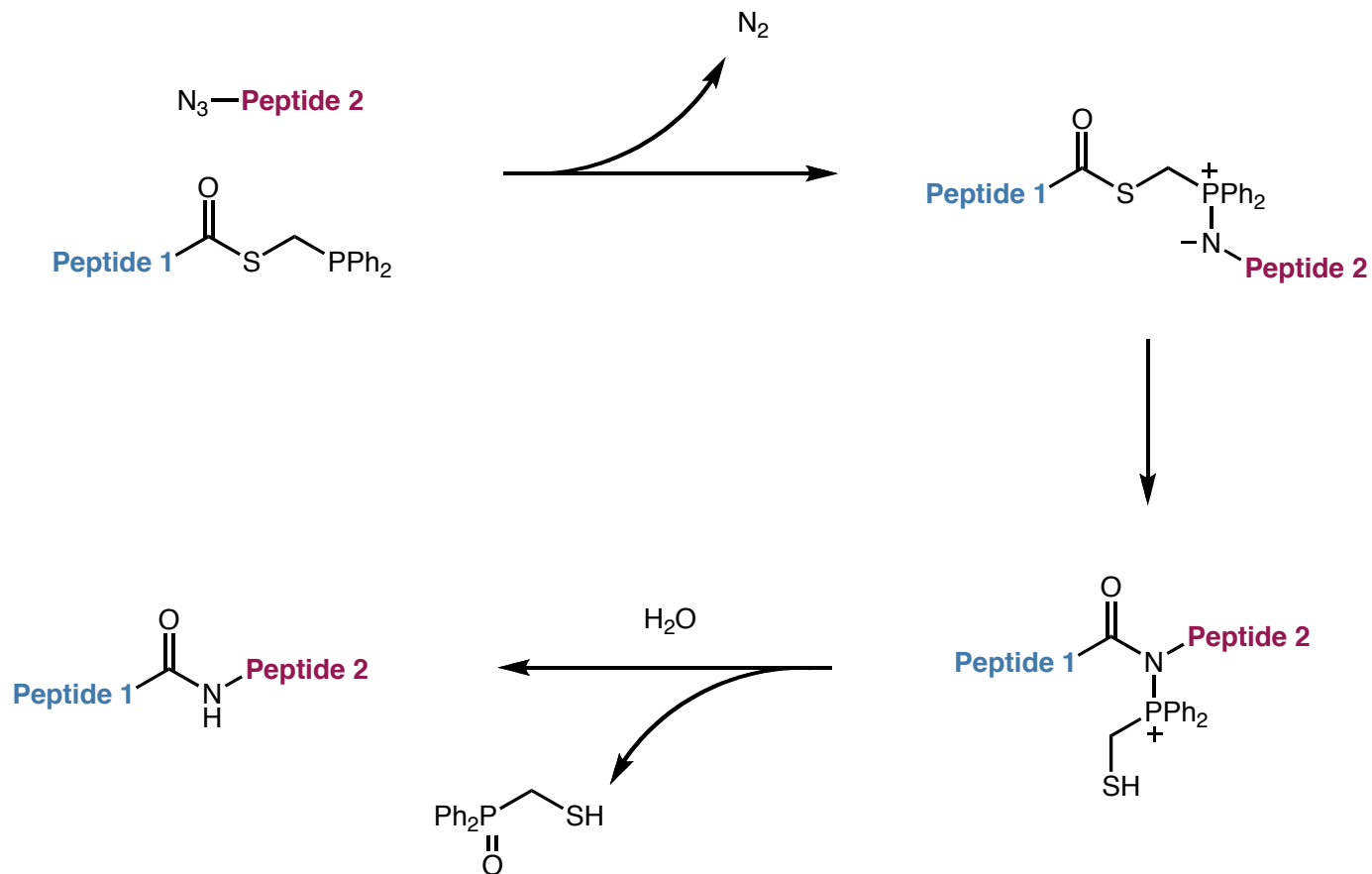
Activation of a Fluorogenic Dye via the Staudinger Ligation

Fluorescence resonance energy transfer (FRET) based probe



"Traceless" Staudinger Ligation

Peptide coupling by the traceless Staudinger ligation

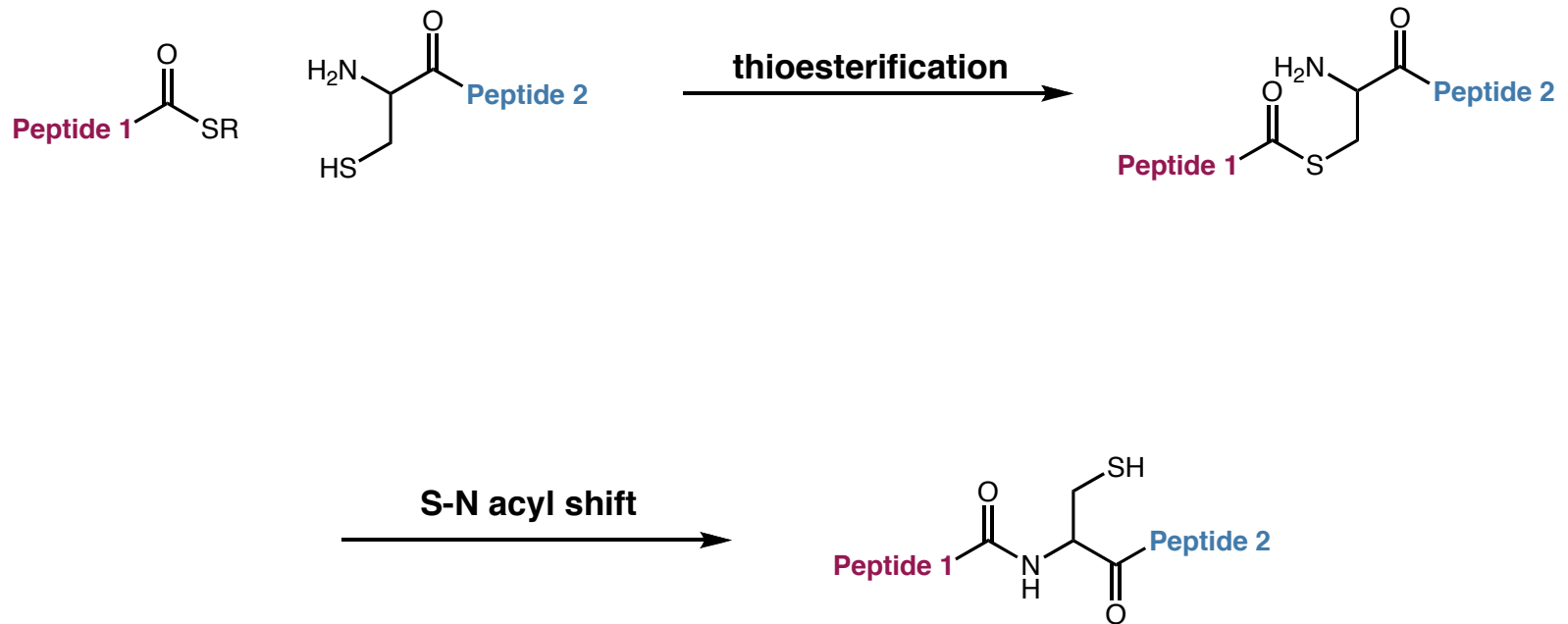


Nilsson, B. L.; Kiessling, L. L.; Raines, R. T. *Org. Lett.* **2000**, *2*, 1939.

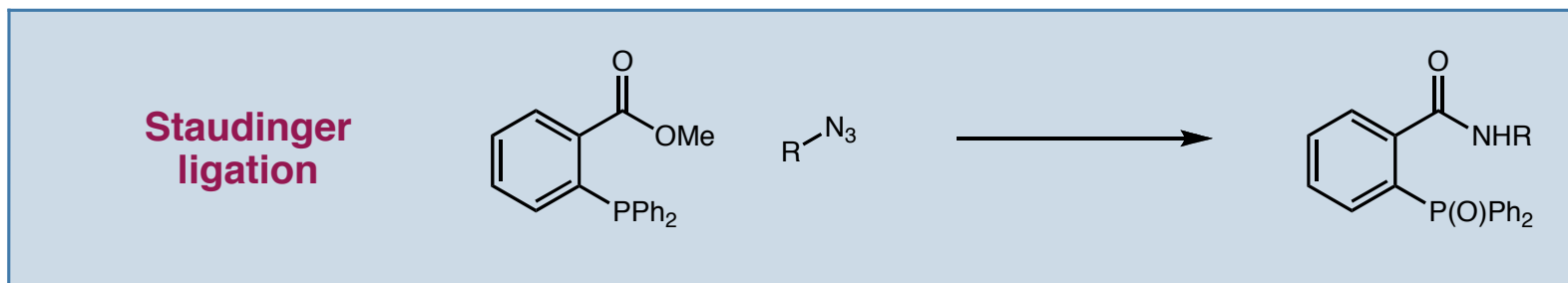
Soellner, M. B.; Nilsson, B. L.; Raines, R. T. *J. Am. Chem. Soc.* **2006**, *128*, 8820.

"Traceless" Staudinger Ligation

Traceless Staudinger ligation is reminiscent of native chemical ligation



Staudinger Ligation



The Staudinger ligation has been used to probe biomolecules within living animals.

Second-order kinetics where the rate-determining step is purported to be attack of the phosphine on the azide.

$$k = 10^{-3} \text{ M}^{-1}\text{s}^{-1} \text{ (very slow)}$$

High concentrations of the phosphine are often necessitated ($> 250 \mu\text{M}$)

Attempts to increase phosphine nucleophilicity has increased the susceptibility of phosphine oxidation.

Lin, F. L.; Hoyt, H. M.; Van Halbeek, H.; Bergman, R. G.; Bertozzi, C. R. *J. Am. Chem. Soc.* **2005**, *127*, 2686.

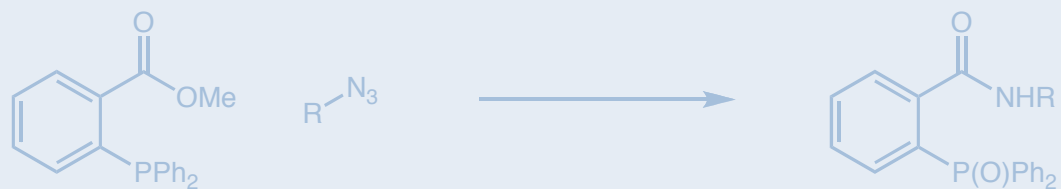
Prescher, J. A.; Dube, D. H.; Bertozzi, C. R. *Nature* **2004**, *430*, 873.

Bioorthogonal Chemistry

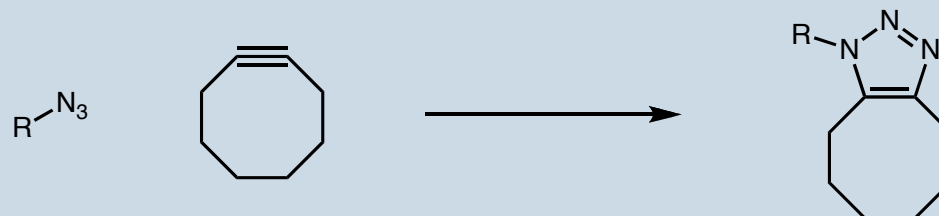
condensation
chemistry



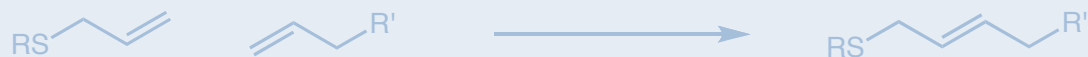
Staudinger
ligation



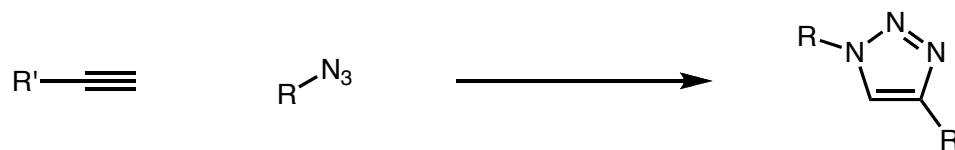
[3+2] cycloadditions



olefin chemistry



[3 + 2] Cycloadditions

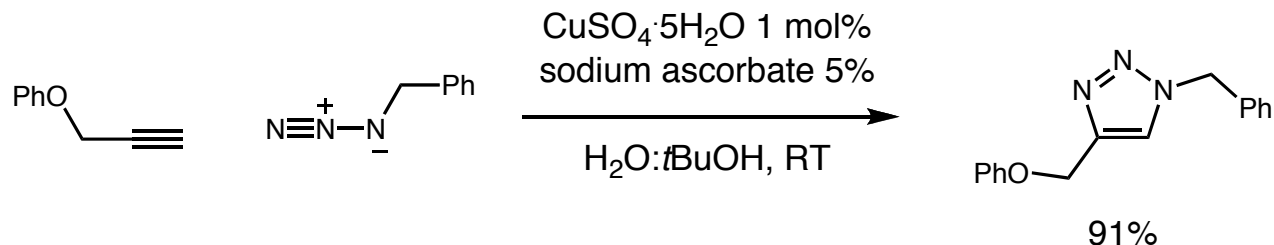


First discovered by Arthur Michael in 1893.

In the 1950s, Rolf Huisgen proposed that the reaction proceeds through a 1,3-dipolar cycloaddition.

High temperatures and pressure required made this reaction largely impractical.

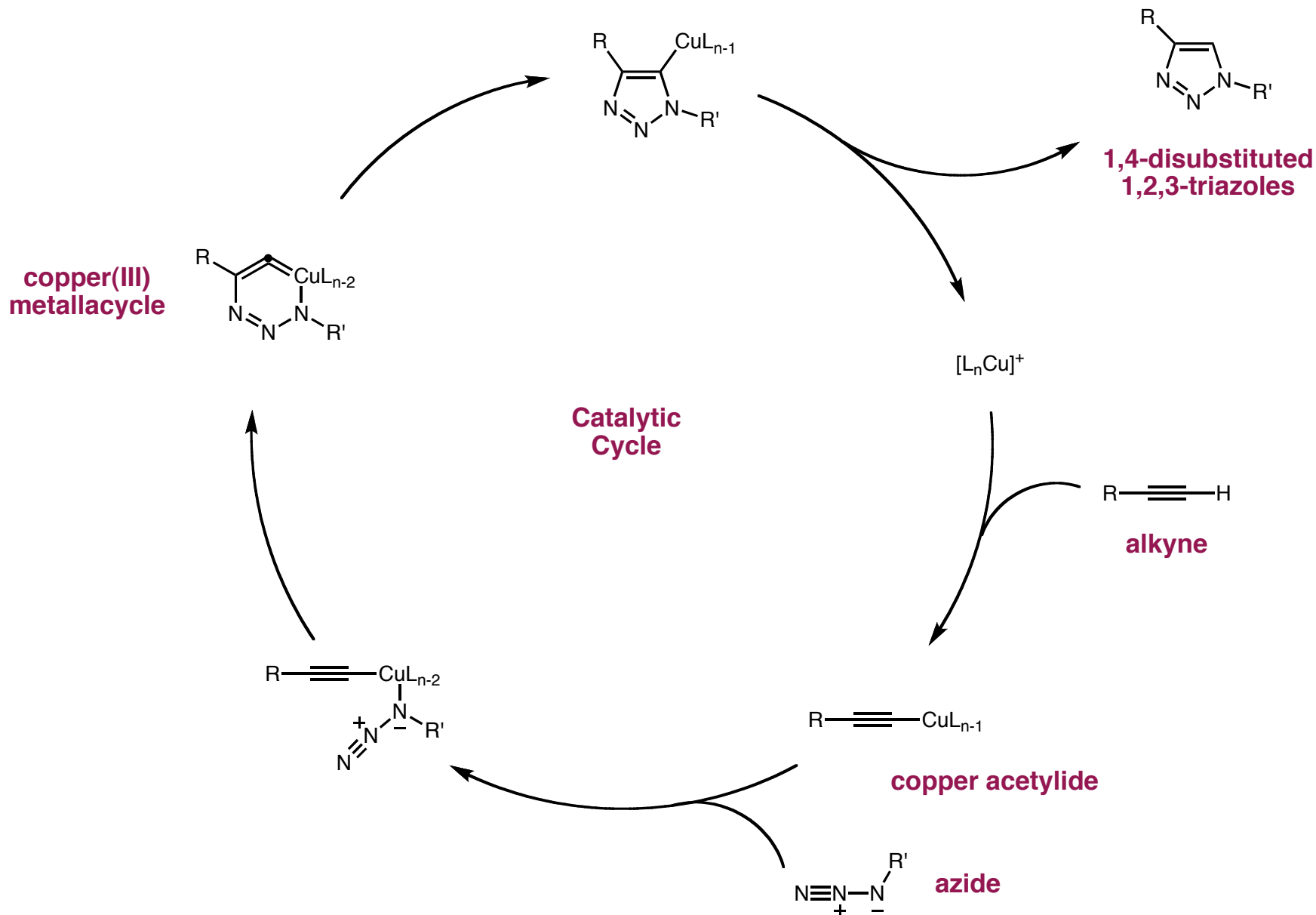
In separate efforts, Meldal and Sharpless reported the copper-catalyzed azide-alkyne 1,3-dipolar cycloaddition (CuAAC).



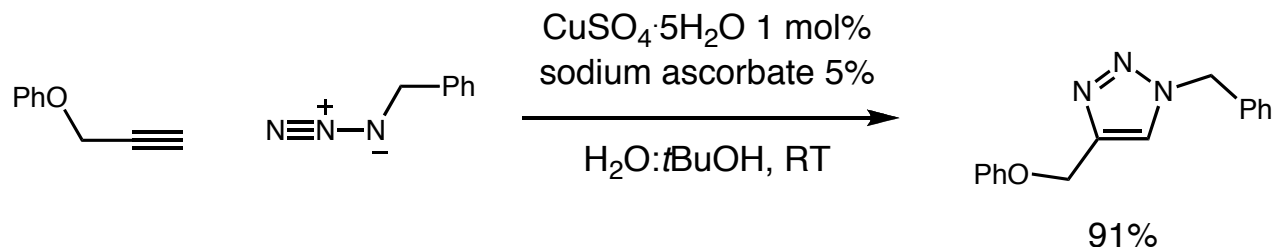
Rostovtsev, V. V.; Green, L. G.; Fokin, V. V.; Sharpless, K. B. *Angew. Chem. Int. Ed.* **2002**, *41*, 2596.

Tornøe, C. W.; Christensen, C.; Meldal, M. *J. Org. Chem.* **2002**, *67*, 3057.

[3 + 2] Cycloadditions



Copper(I) Toxicity



Finn, Sharpless, and coworkers reported the first biomolecule coupling application of [3+2] cycloaddition through the attachment of dyes to the cowpea mosaic virus.

CuAAC is not widely employed, due to copper(I)'s toxicity.

E. Coli stops dividing after exposure to 100 μM CuBr for 16 hours.

Mammalian cells and zebrafish embryos can survive low concentrations of copper (I) (< 500 μM) but considerable cell death is observed above 1 mM.

Wang, Q.; Chan, T. R.; Hilgraf, R.; Fokin, V. V.; Sharpless, K. B.; Finn, M. G. *J. Am. Chem. Soc.* **2003**, *125*, 3192-3193.

Chan, T. R.; Hilgraf, R.; Sharpless, K. B.; Fokin, V. V. *Org. Lett.* **2004**, *6*, 2853.

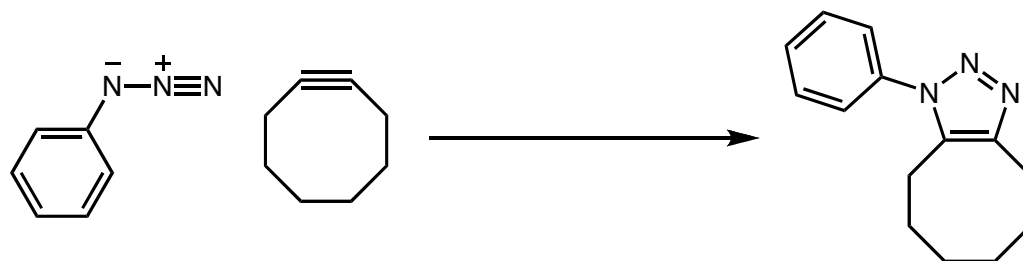
Link, A. J.; Tirrell, D. A. *J. Am. Chem. Soc.* **2003**, *125*, 11164.

Speers, A. E.; Adam, B. F.; Cravatt, B. F. *J. Am. Chem. Soc.* **2003**, *125*, 4686.

[3+2] Cycloaddition - Ring Strain

Strain-promoted azide cycloadditions were first investigated by Alder and Stein in the 1930s.

In the 1960s, Krebs and Wittig commented that phenylazide and cyclooctyne



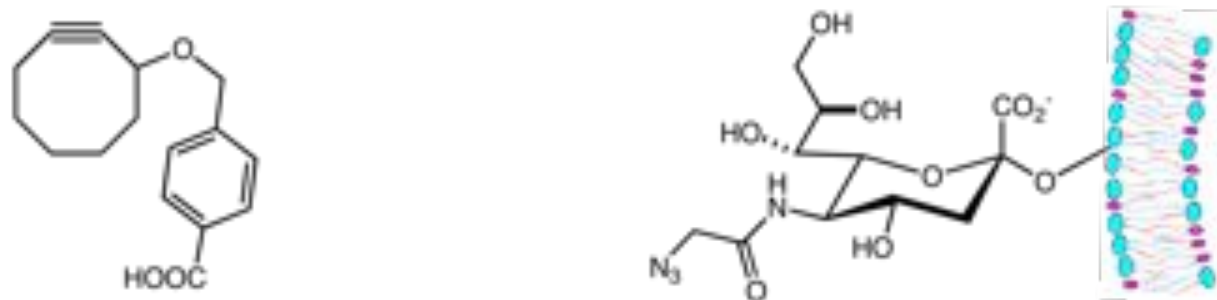
"proceeded
like an explosion"

massive bond angle deformation of the acetylene to 163°
 ~ 18 kcal/mol of ring strain

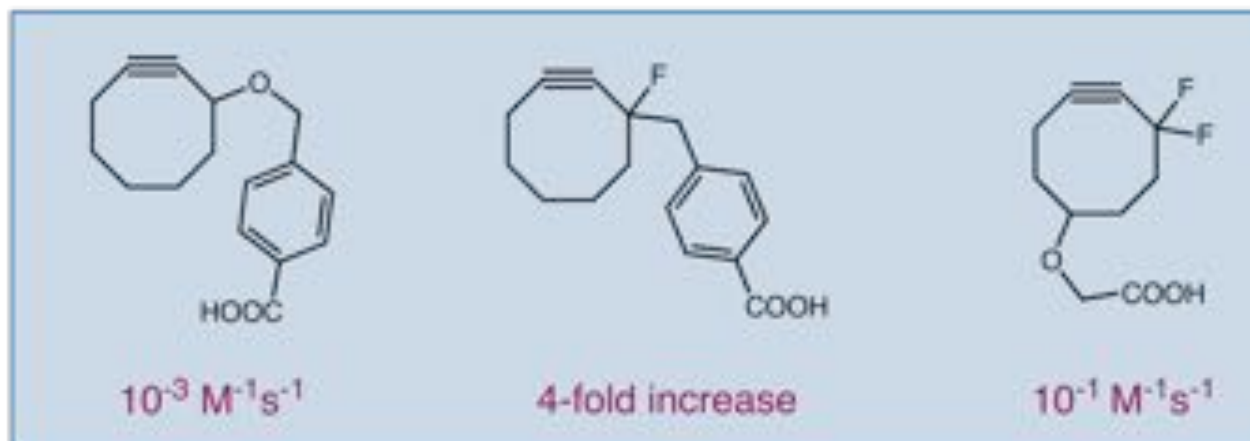
Wittig, G.; Krebs, A. *Chem. Ber.* **1961**, *94*, 3260.
Alder, K.; Stein, G. *Justus Liebigs Ann. Chem.* **1931**, *485*, 211.

[3+2] Cycloaddition - Ring Strain

Cyclooctynes react smoothly on cell-surface glycans with no apparent cytotoxic effects.



Reactions kinetics is still an issue.

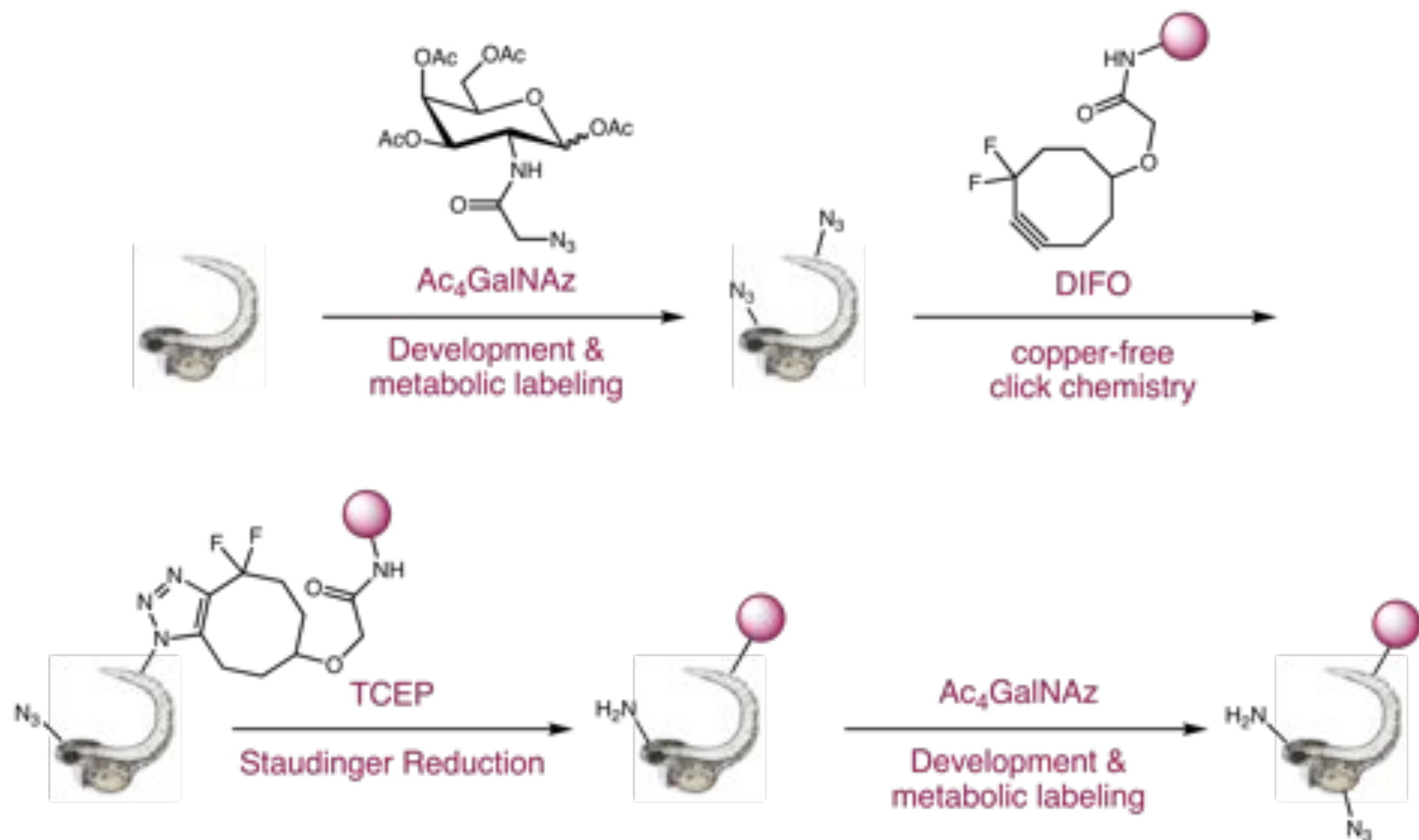


Agard, N. J.; Prescher, J. A.; Bertozzi, C. R. *J. Am. Chem. Soc.* **2004**, *126*, 15046.

Agard, N. J.; Baskin, J. M.; Prescher, J. A.; Lo, A.; Bertozzi, C. R. *ACS Chem. Biol.* **2006**, *1*, 644.

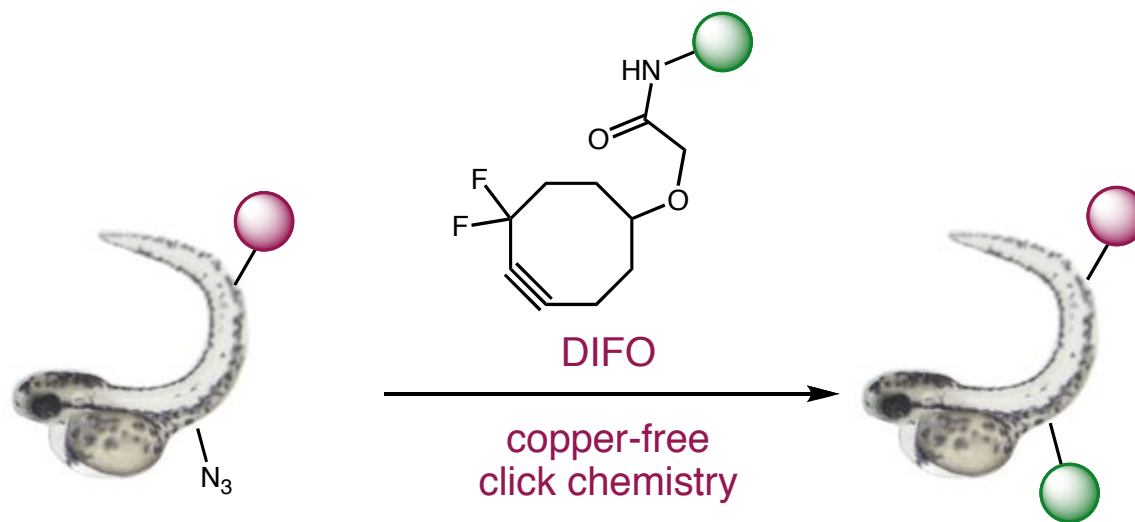
Baskin, J. M.; Prescher, J. A.; Laughlin, S. T.; Agard, N. J.; Chang, P. V.; Bertozzi, C. R. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 16793.

In Vivo Imaging of Zebrafish Embryos



Laughlin, S. T.; Baskin, J. M.; Amacher, S. L.; Bertozzi, C. R. *Science* **2008**, *320*, 664.

In Vivo Imaging of Zebrafish Embryos



In Vivo Imaging of Zebrafish Embryos

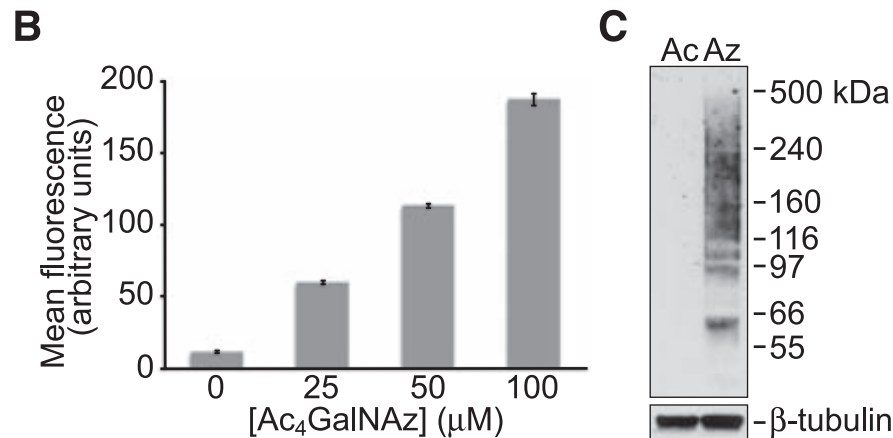
First confirmed that zebrafish glycan biosynthetic enzymes are tolerant of the unnatural sugar.

Zebrafish cell line ZF4 was incubated with various doses of Ac₄GalNAz_m reacted with DIFO-488 and analyzed by flow cytometry.

Azide-labeled cell lysates were further characterized by treatment with a DIFO-Flag peptide conjugate.

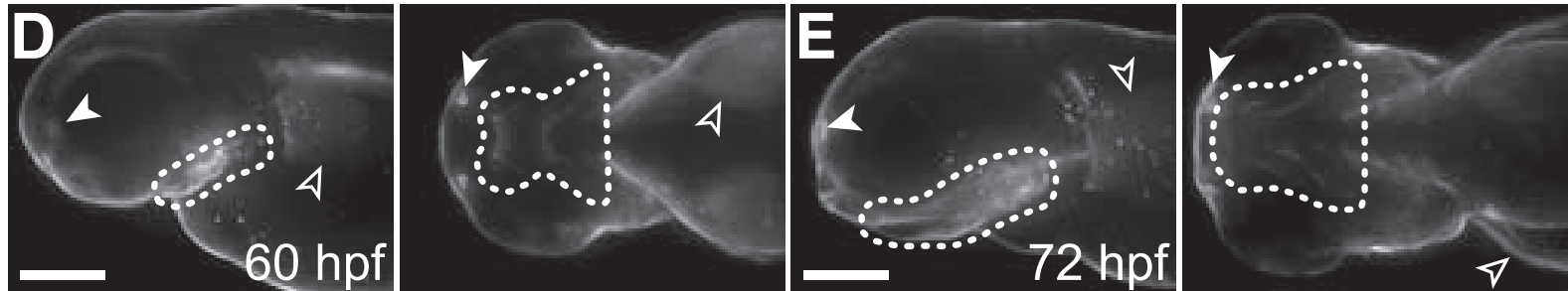
Observed high-molecular weight species were consistent with labeled glycoproteins.

Flag-containing species (glycoproteins like b-hexosaminidase, b-integrin, nicastrin) were known or predicted sites of mucin-type O-linked glycosylation.

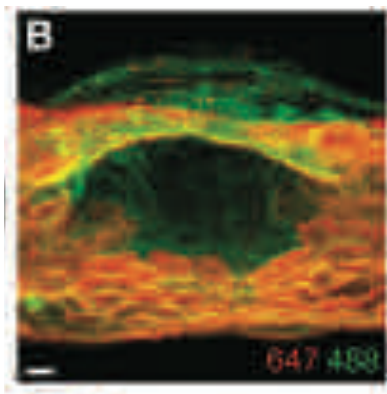


In Vivo Imaging of Zebrafish Embryos

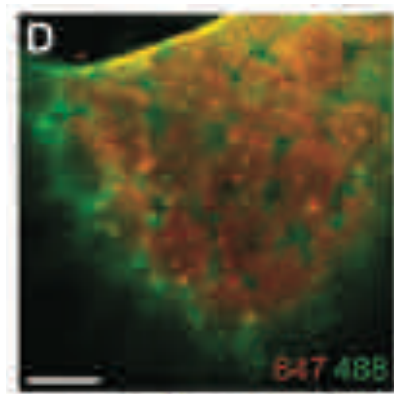
From 60 hours post-fertilization (hpf) to 72 hpf, a burst in fluorescence intensity in the jaw region, pectoral fins, and olfactory organs.



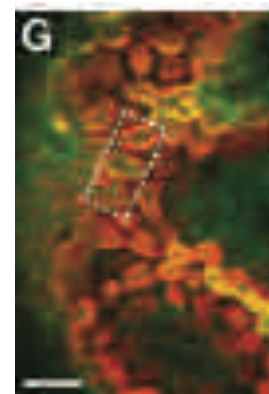
These three areas are reexamined and reacted with **DIFO-647** between 60 and 61 hpf and **DIFO-488** between 61 and 62 hpf.



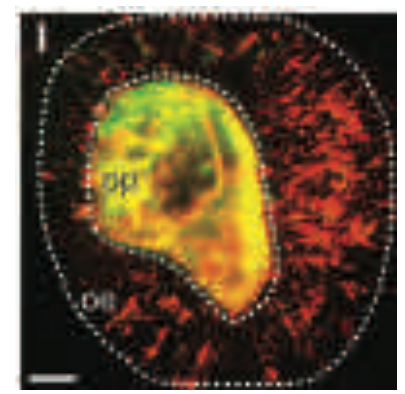
mouth region



pectoral fin



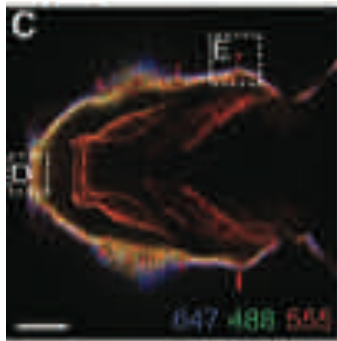
jaw region



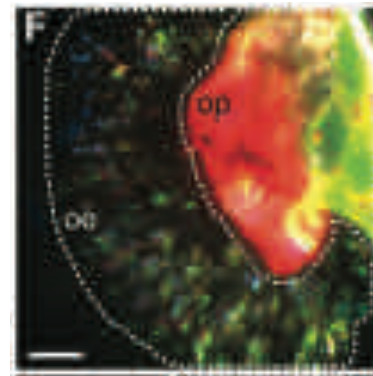
olfactory region

In Vivo Imaging of Zebrafish Embryos

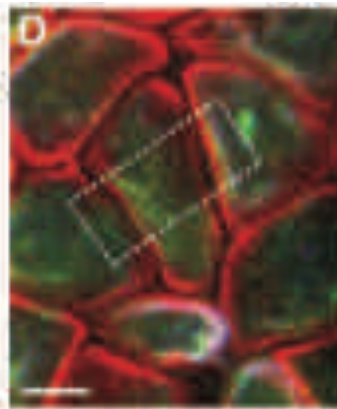
Three-dye bioimaging - **DIFO-647** between 60 and 61 hpf, **DIFO-488** between 62 and 62 hpf, and then **DIFO-555** between 72 and 73 hpf.



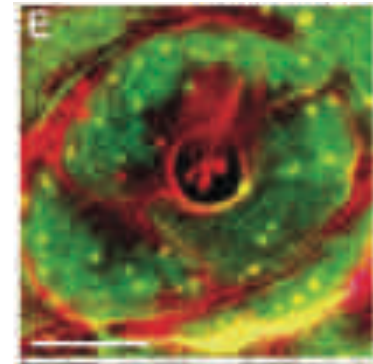
jaw region



olfactory region

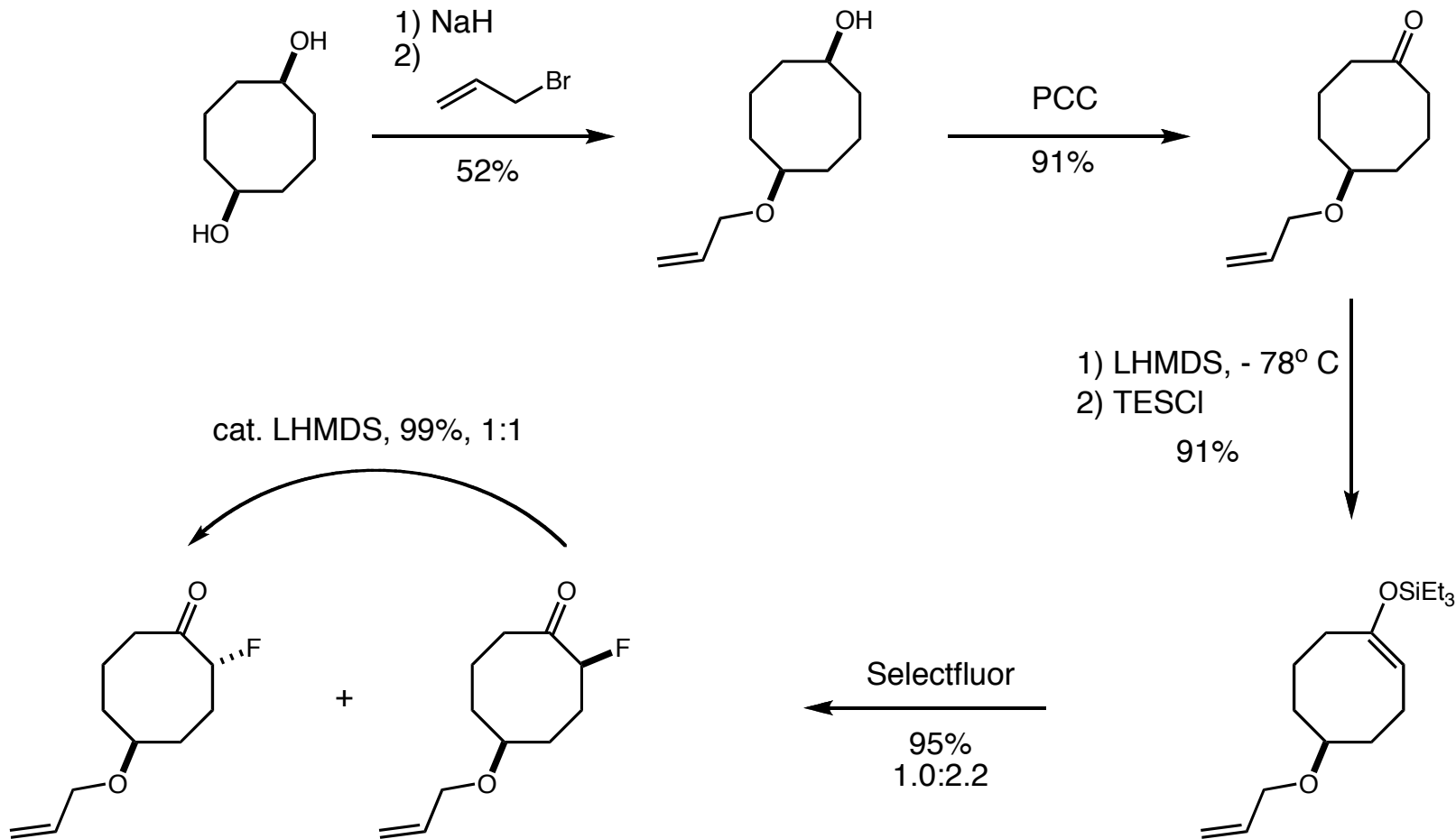


cells

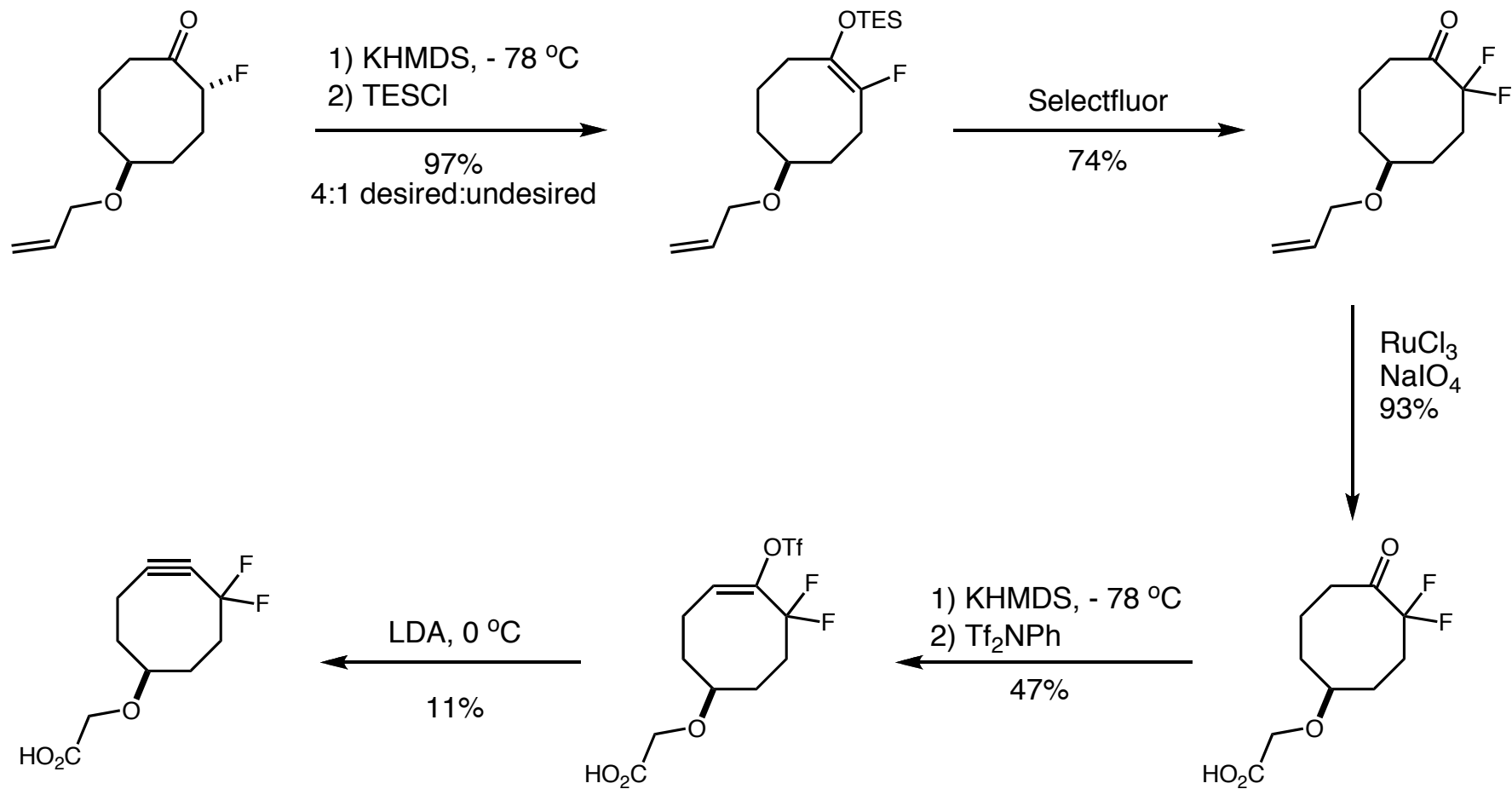


kinocilia

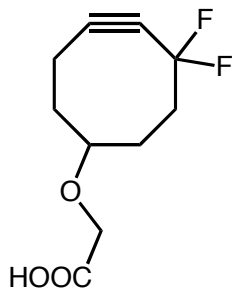
DIFO Synthesis



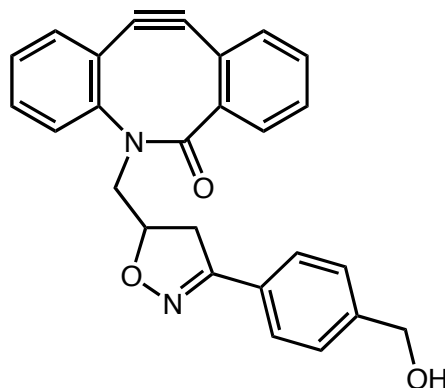
DIFO Synthesis



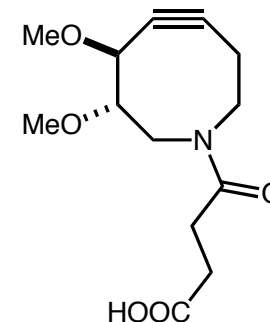
Cyclooctyne Analogues



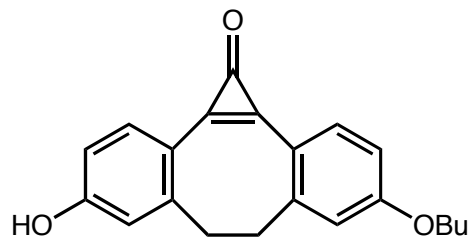
DIFO, Bertozzi Lab
 $k = 0.076 \text{ M}^{-1}\text{s}^{-1}$



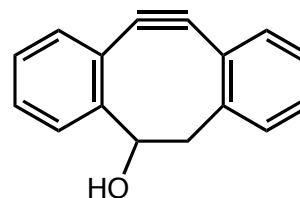
BARAC, Bertozzi Lab
 $k = 0.96 \text{ M}^{-1}\text{s}^{-1}$



DIMAC, Bertozzi Lab
more water soluble
 $k = 0.003 \text{ M}^{-1}\text{s}^{-1}$



Boons and Popik Labs
photocaged
 $k = 0.076 \text{ M}^{-1}\text{s}^{-1}$



DIBO, Boons Lab
5 steps
nontoxic, $k = 0.057 \text{ M}^{-1}\text{s}^{-1}$

Sletten, E. M.; Bertozzi, C. R. *Org. Lett.* **2008**, *10*, 3097.

Ning, X. H.; Guo, J.; Wolfert, M. A.; Boons, G. J. *Angew. Chem. Int. Ed.* **2008**, *47*, 2253.

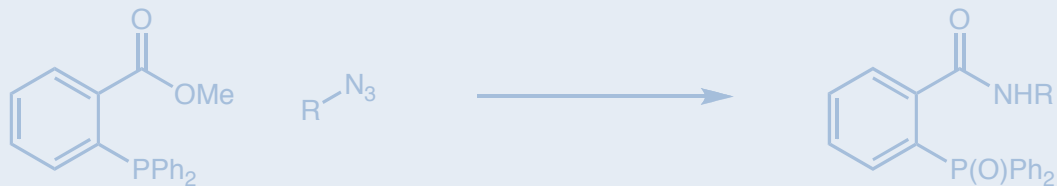
Sletten, E. M.; Bertozzi, C. R. *Acc. Chem. Res.* **2009**, *44*, 666-676.

Bioorthogonal Chemistry

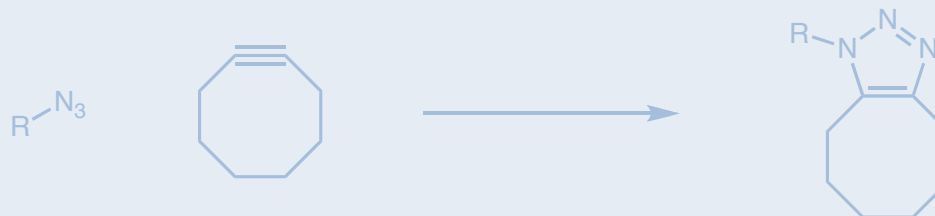
condensation
chemistry



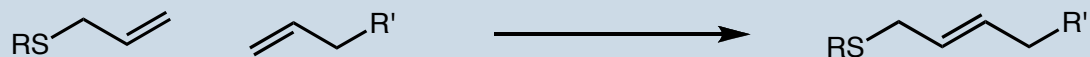
Staudinger
ligation



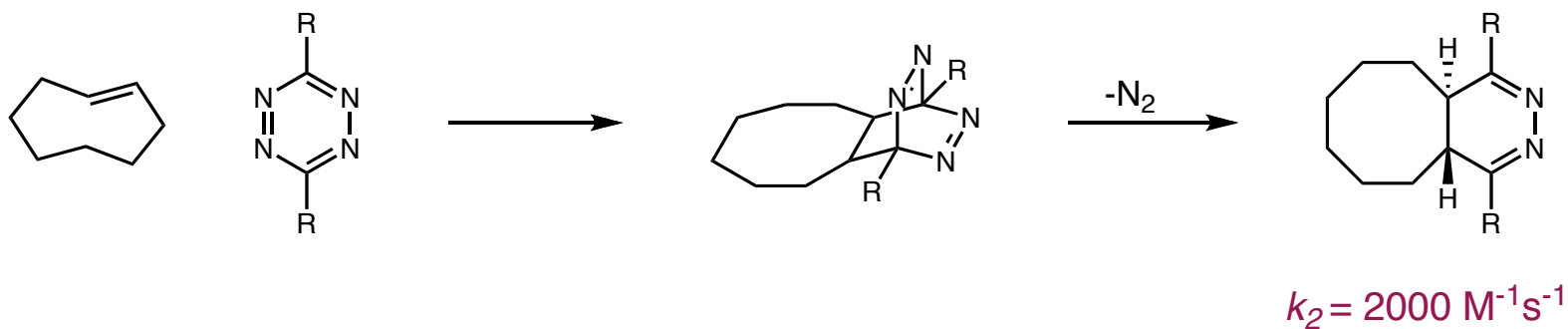
[3+2] cycloadditions



olefin chemistry

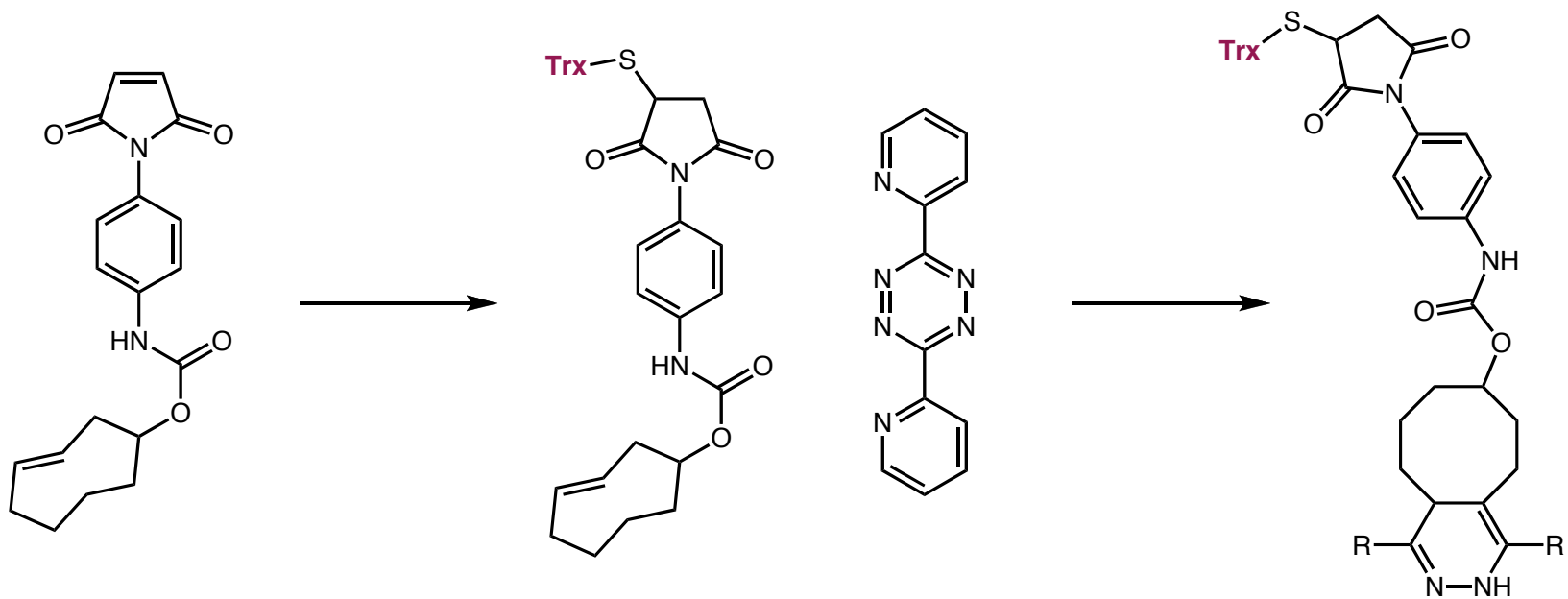


Inverse-Electron Demand Diels-Alder

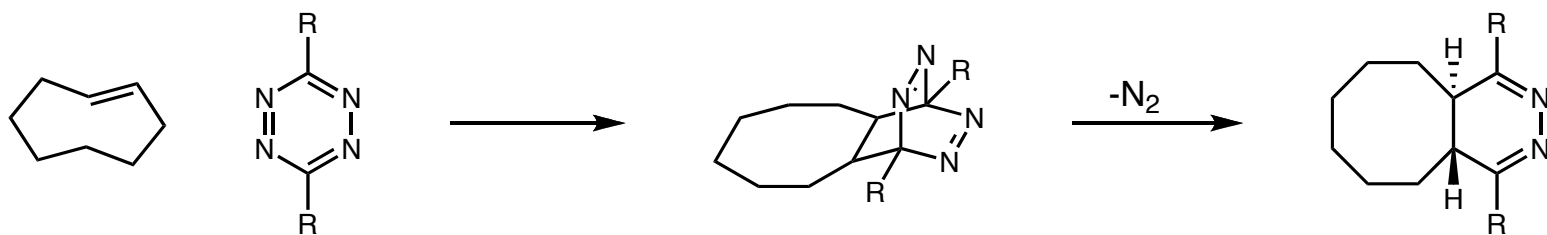


thioredoxin

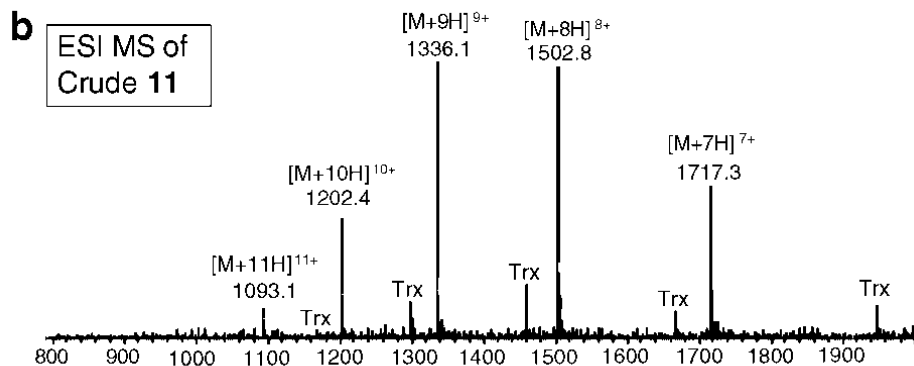
Trx-SH



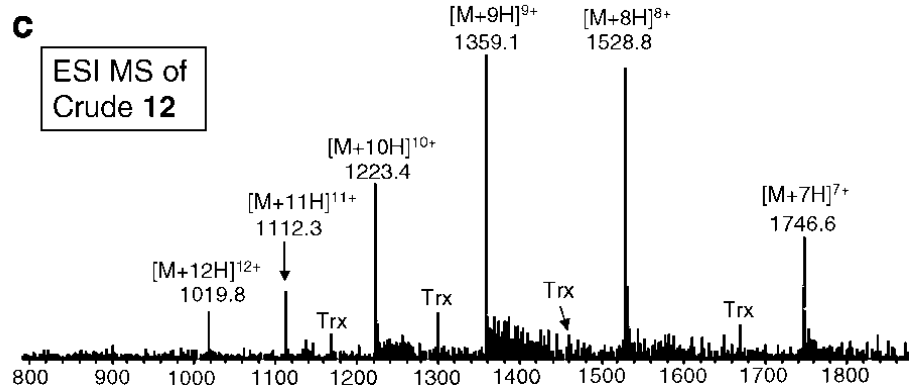
Inverse-Electron Demand Diels-Alder



$$k_2 = 2000 \text{ M}^{-1}\text{s}^{-1}$$



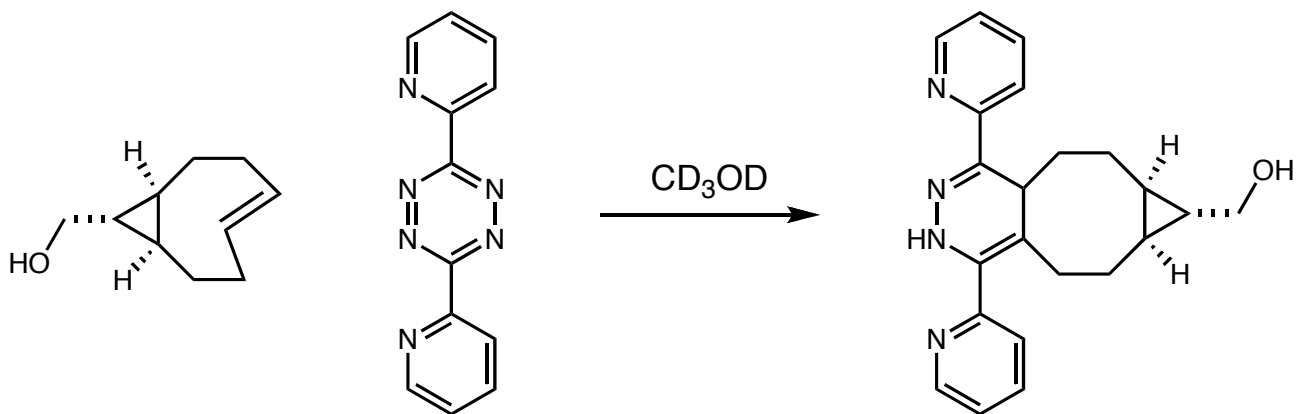
Trx = 11700 Da



Michael product = 12014 Da

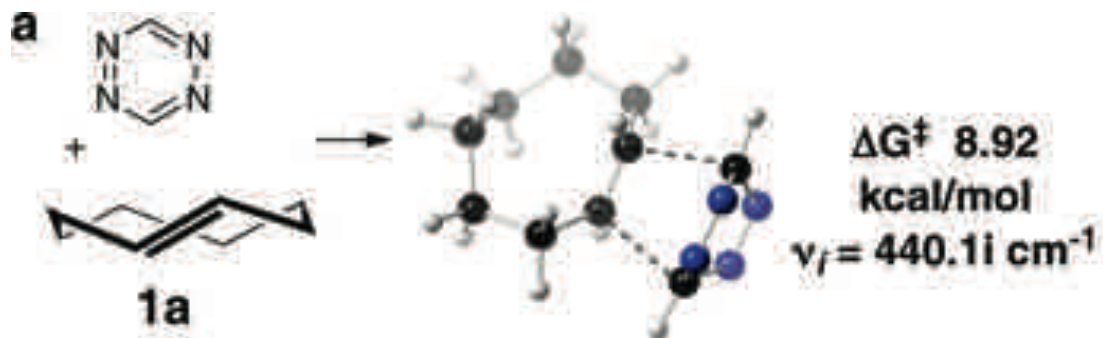
Diels-Alder product = 12222 Da

Inverse-Electron Demand Diels-Alder

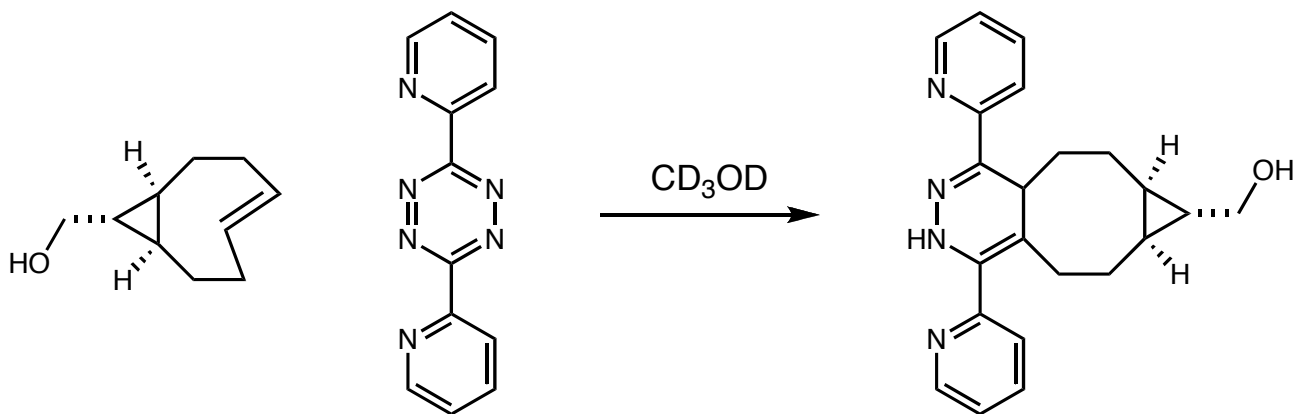


$$k_2 = 22,000 \text{ M}^{-1} \text{ s}^{-1}$$

trans-cyclooctene ring is designed via computation (M06L/6-311+G(d,p)-optimized transition structure)

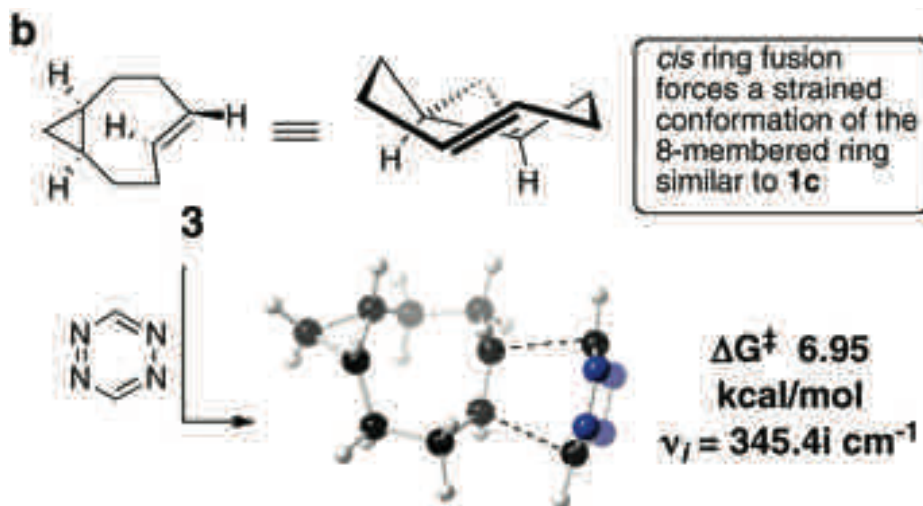


Inverse-Electron Demand Diels-Alder

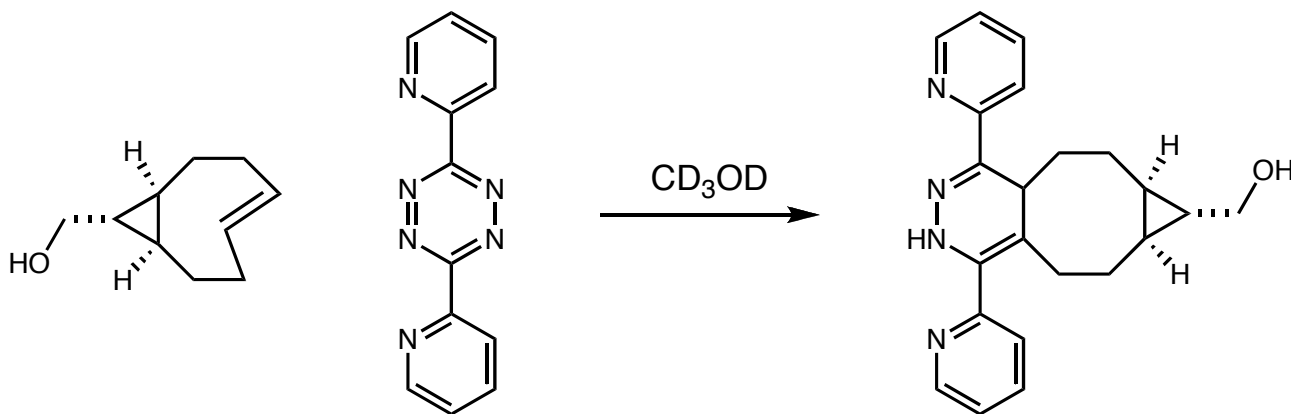


$$k_2 = 22,000 \text{ M}^{-1} \text{ s}^{-1}$$

trans-cyclooctene ring is designed via computation (M06L/6-311+G(d,p)-optimized transition structure)

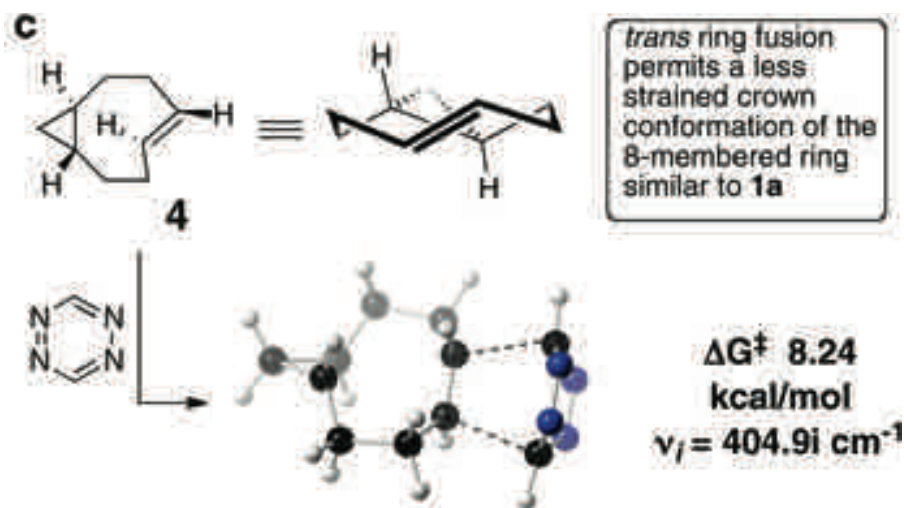


Inverse-Electron Demand Diels-Alder



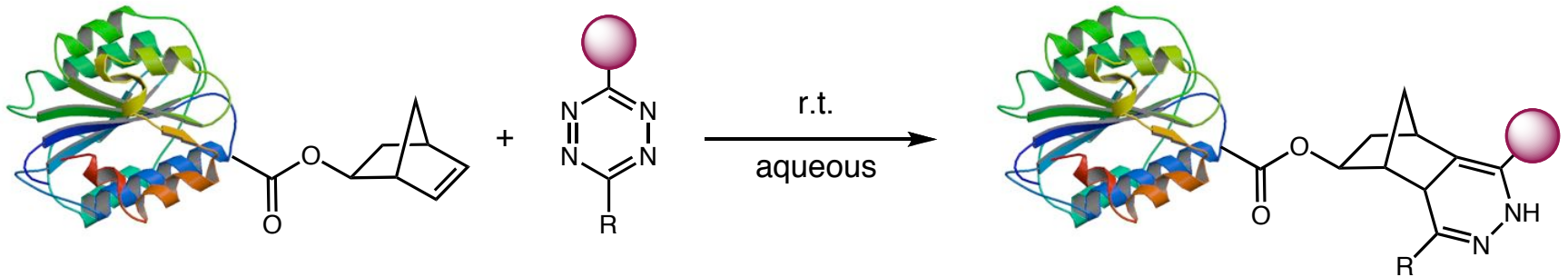
$$k_2 = 22,000 \text{ M}^{-1} \text{ s}^{-1}$$

trans-cyclooctene ring is designed via computation (M06L/6-311+G(d,p)-optimized transition structure)

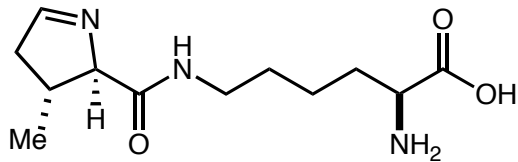


Inverse-Electron Demand Diels-Alder

Genetically encoded norbornene directs site-specific cellular protein labelling



Orthogonal synthetase/tRNA pair used to install a norbornene-containing amino acid



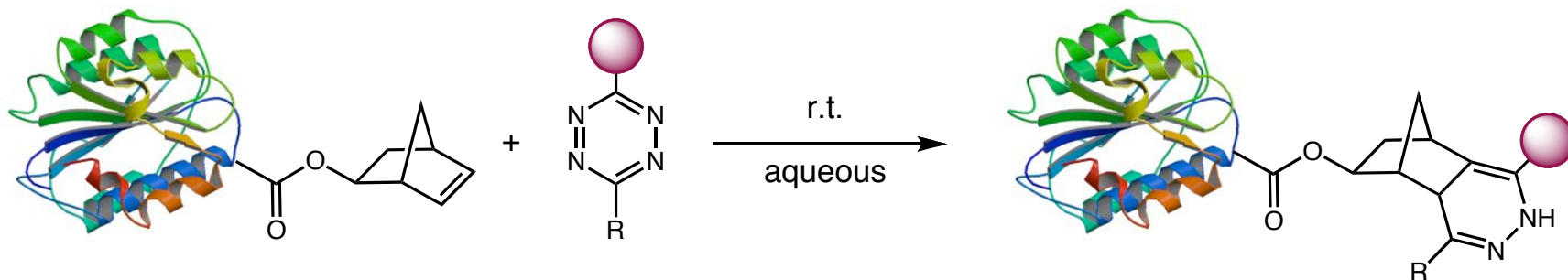
Pyrrolysine
"The 22nd Amino Acid"

Genetically coded amino acid used by some methanogenic archaea.

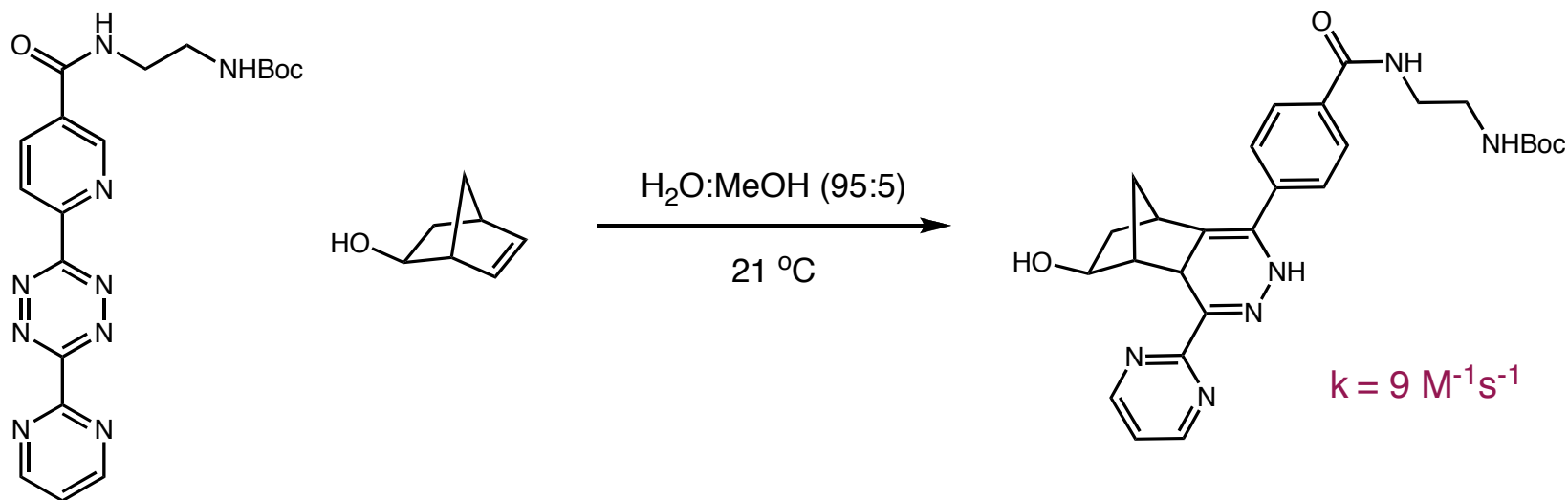
Pyrrolysyl-tRNA synthetase/tRNA_{CUA} is orthogonal to endogenous tRNAs and aminoacyl-tRNA synthetases in *E. coli* and eukaryotic cells.

Inverse-Electron Demand Diels-Alder

Genetically encoded norbornene directs site-specific cellular protein labelling

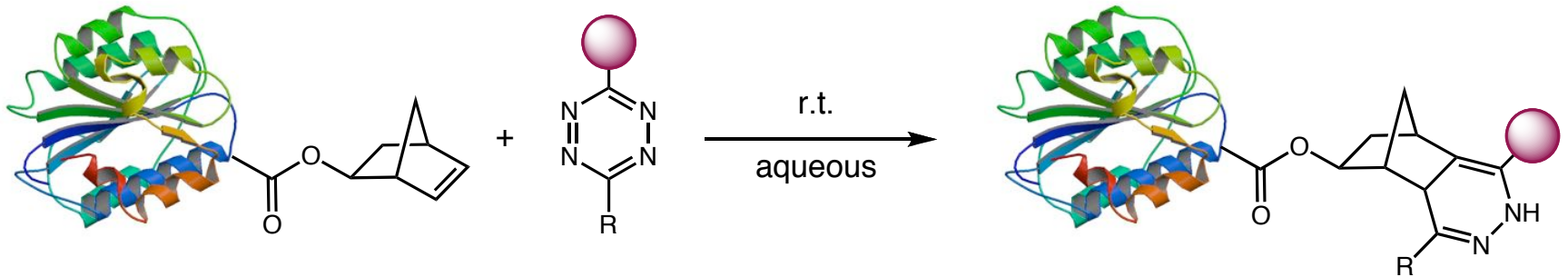


Orthogonal synthetase/tRNA pair used to install a norbornene-containing amino acid



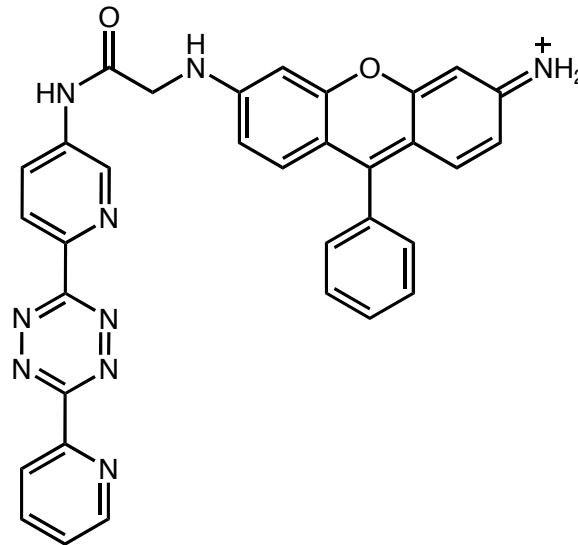
Inverse-Electron Demand Diels-Alder

Genetically encoded norbornene directs site-specific cellular protein labelling



The fluorescence of certain probes increased by a 5-10 fold increase after the cycloaddition

"turn-on" fluorophorogenic probes - tetrazine can quench fluorophore via energy transfer

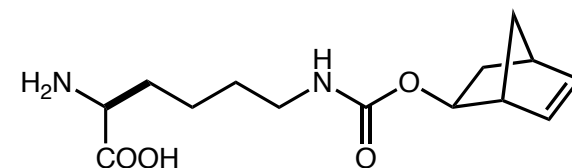
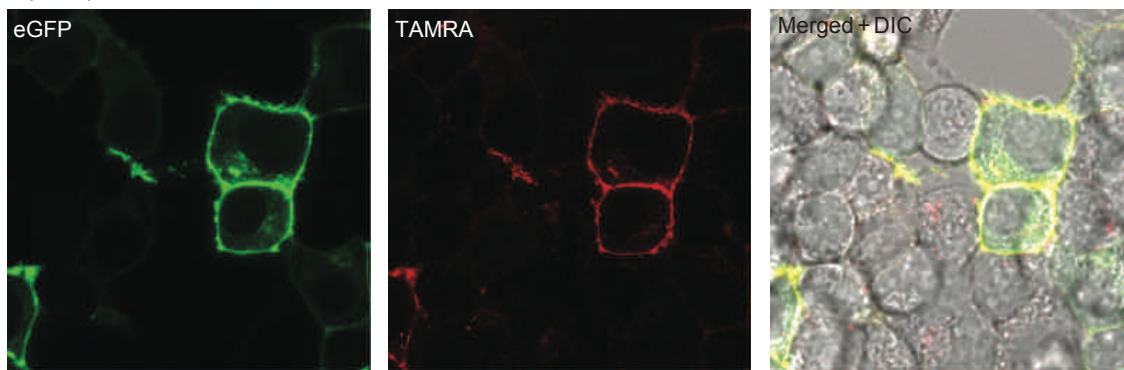


tetramethylrhodamine
(TAMRA)

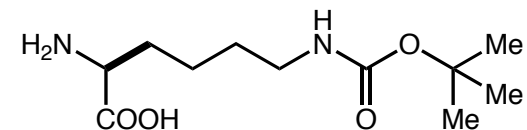
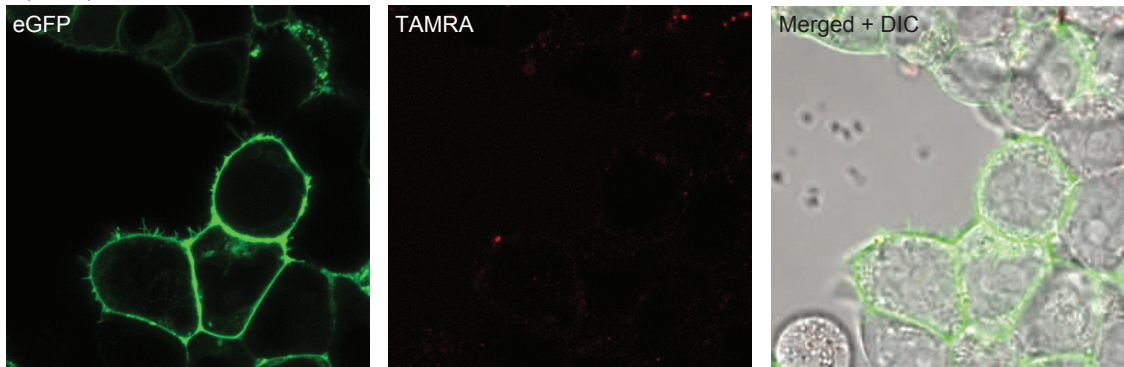
Inverse-Electron Demand Diels-Alder

Cycloaddition tested with mutated epidermal growth factor receptor (EGFR-GFP)

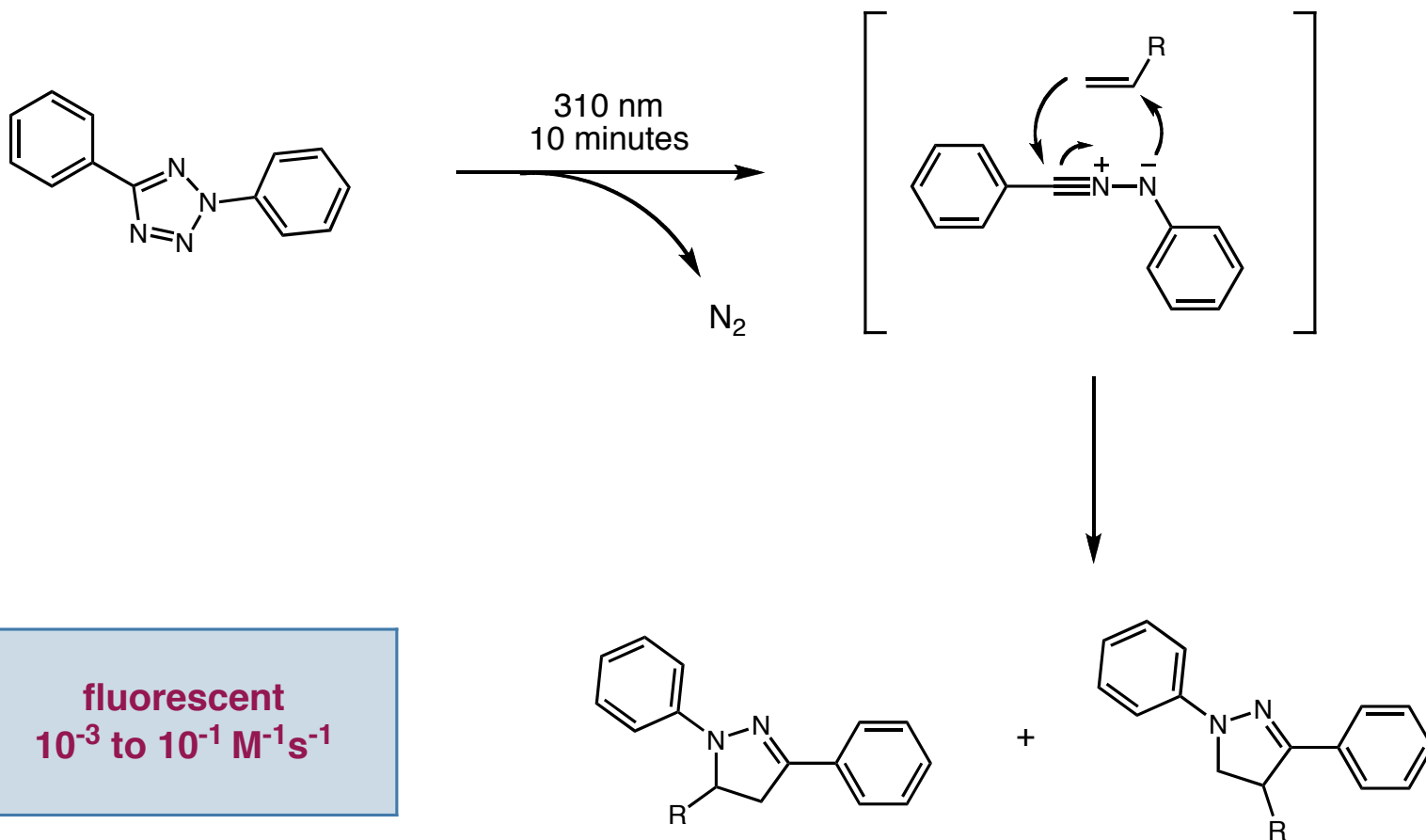
2 (1 mM)



3 (1 mM)



Photochemical 1,3-Dipolar Cycloaddition



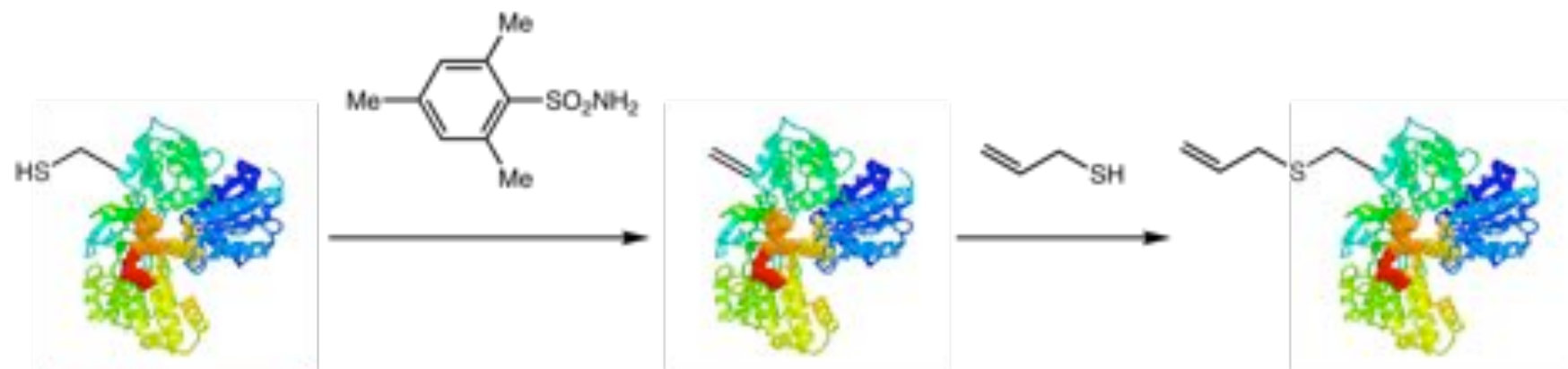
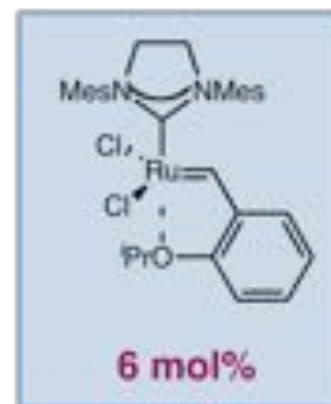
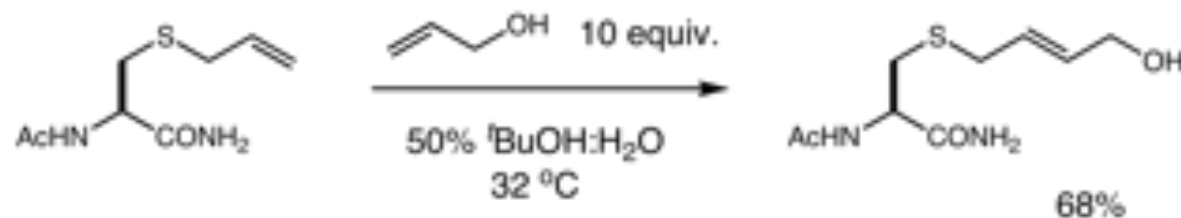
Song, W.; Wang, Y.; Qu, J.; Madden, M. M.; Lin, Q. *Angew. Chem. Int. Ed.* **2008**, *47*, 2832.

Wang, Y.; Hu, W. J.; Song, W.; Lim, R. K. V.; Lin, Q. *Org. Lett.* **2008**, *10*, 3752.

Song, W.; Wang, Y.; Lin, Q. *J. Am. Chem. Soc.* **2008**, *130*, 9654.

Bioorthogonal Cross-Metathesis

Interestingly, allyl sulfides are required due to purported sulfur coordination to the ruthenium center.

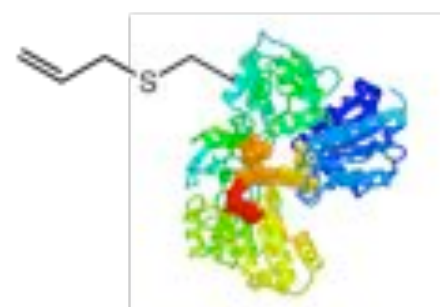


serine protease
subtilisin *Bacillus lentus* (SBL)

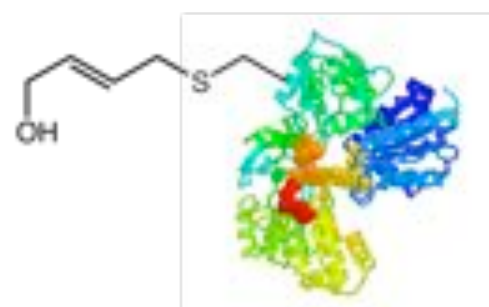
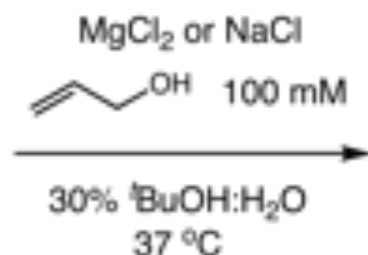
Bernardes, G. J. L.; Chalker, J. M.; Errey, J. C.; Davis, B. G. *J. Am. Chem. Soc.* **2008**, *130*, 5052-5053.

Lin, Y. A.; Chalker, J. M.; Floyd, N.; Bernardes, G. J. L.; Davis, B. G. *J. Am. Chem. Soc.* **2008**, *130*, 9642-9643.

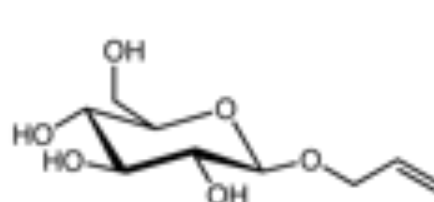
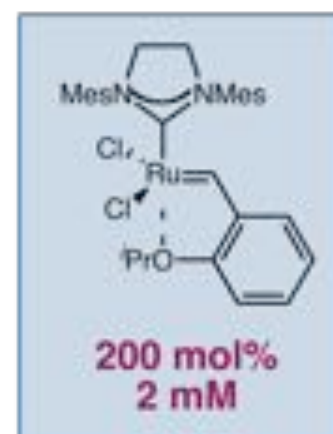
Bioorthogonal Cross-Metathesis



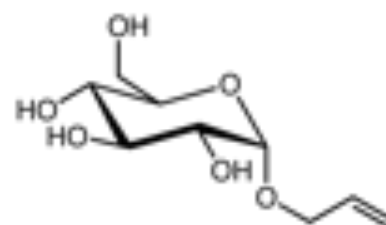
0.01 mM



90% yield



50%



60%

Salts, such as MgCl₂ or NaCl, were added to disrupt any nonproductive chelation to ruthenium

Bernardes, G. J. L.; Chalker, J. M.; Errey, J. C.; Davis, B. G. *J. Am. Chem. Soc.* **2008**, *130*, 5052-5053.

Lin, Y. A.; Chalker, J. M.; Floyd, N.; Bernardes, G. J. L.; Davis, B. G. *J. Am. Chem. Soc.* **2008**, *130*, 9642-9643.

Future Outlook

Groups 15 elements have been particularly lucrative.

Perhaps larger elements in this group, like bismuth or antimony, may be of use.

Pericyclic reactions have been very promising as well, due to concerted mechanisms that leave little room for interruption from other components.

Applications with other sources of energy, such as light or ultrasound, may occur.

We may see the extension of chemical reporters to other small-molecule metabolites.

