

Tomer. M. Faraggi MacMillan Lab Group Meeting November 29th 2016

Enzymes as an inspiration for reaction development

Biochemical reactions are catalysed by enzymes with a high degree of precision, under mild conditions



Regioselective hydroxylation in cholesterol biosynthesis



Asymmetric transamination in amino acid biosynthesis

There has been significant interest in mimicking the effect of enzymes in synthetic processes

Biocatalysis and Biomimetic catalysis





Biocatalysis:

The use of enzymes or whole cells as catalysts for synthetic chemistry Biomimetic catalysis:

Chemical catalysis that mimics key features of enzymatic systems

Enzyme structure consists of an active site surrounded by a protein superstructure



Binding site - Binds and orients substrate Provides control for selectivity of reaction Usually specific to a particular substrate

Catalytic site - Groups which catalyze reaction For example groups of amino acid residues Or metal cofactors bound by protein structure

Attempts made to mimic the effects of both sites









C–H functionalization inspired by oxidase enzymes

Oxidase enzymes are integral in the biosynthesis of many natural products and metabolic processes



Taxadiene oxidation in the biosynthesis of taxol

A number of oxidase enzymes have been utilized in industrial scale processes



Selective enzyme mediated oxygenation in the fermentation of artemisinic acid

Li, X-W.; Nay, B. *Nat. Prod. Rep.* **2014**, *31*, 533 Holtmann, D.; Fraajie, M. W.; Arends, I. W. C. E.; Opperman, D. J.; Hollmann, F. *Chem. Commun.* **2014**, *50*, 13180

C–H functionalization inspired by oxidase enzymes



Reactive oxygenation agents at the active sites of some common oxidase enzymes

Gamez, P.; Aubel, P. G.; Driessen, W. L.; Reedijik, J. *Chem. Soc. Rev.* 2001, *30*, 376 Holtmann, D.; Fraajie, M. W.; Arends, I. W. C. E.; Opperman, D. J.; Hollmann, F. *Chem. Commun.* 2014, *50*, 13180

C–H functionalization inspired by oxidase enzymes



Reactive oxygenation agents at the active sites of some common oxidase enzymes

Gamez, P.; Aubel, P. G.; Driessen, W. L.; Reedijik, J. *Chem. Soc. Rev.* 2001, *30*, 376 Holtmann, D.; Fraajie, M. W.; Arends, I. W. C. E.; Opperman, D. J.; Hollmann, F. *Chem. Commun.* 2014, *50*, 13180

Cytochrome P450 class of enzymes



Class of enzymes containing an iron heme cofactor, responsible for biological C–H oxidations

Mechanism proceeds through dioxygen activation and atom transfer via a high valent Fe^{IV} intermediate

Structure has inspired the development of a number of catalytic strategies based on high valent metals

Cytochrome P450 - Mechanism of action



Nam, W. Acc. Chem. Res. 2007, 40, 522

First synthetic reactions by Fe porphyrin complexes



Oxidation reactions utilizing metalloporphyrin catalysts

| x Ar X | Me | tals Axial liga | ands Terminal oxic | dants |
|---------|----|----------------------|------------------------|-------|
| x | N | n Cl ⁻ | O ₂ /air | |
| | F | e OH [.] | H_2O_2 | |
| Ar Ar | C | o AcO | - PhIO | |
| x h h x | R | u CF ₃ SC | D ₃ - NaClO | |
| X Ar X | C | s MeO | H KHSO ₅ | j |

Catalyst design by variation of porphyrin substituents, metal and axial ligand

In general more electron deficient porphyrin ligands lead to a more oxidising catalyst

Effects of axial ligand arise from *trans* effect - more electron donating ligand weakens M=O bond

Reaction mechanism has been adapted for unnatural reactions such as amination

Oxidation reactions utilizing metalloporphyrin catalysts



Effect of axial donation on the energies of the frontier orbitals of iron oxo complexes

Oxidation reactions utilizing metalloporphyrin catalysts



C-H hydrocarbon oxidation has been a major focus of metalloporphyrin chemistry

Oxidation reactions utilizing metalloporphyrin catalysts



Oxidation of other functionalities such as arenes and olefins has also been explored

Hydroxylation of unactivated C–H bonds by Fe porphyrins



Oxidation of other functionalities such as arenes and olefins has also been explored



Fluorinated aromatic on porphyrin core protects against oxidative degradation

Hydroxylation of unactivated C–H bonds by Fe porphyrins



Metalloporphyrin catalyzed C–H amination

First example of C-H amination on aromatic stereoid sybstrate



47% yield approx 40 turnovers

Unlike corresponding oxidation process, no side reaction on aromatic ring



Metalloporphyrin catalyzed C–H amination

Conditions expanded to allow for the use of amines as aminating agent



Amination of unactivated C–H bonds

Metal nitrene catalysts cannot functionalize unactivated hydrocarbons as readily as their oxo analogues



Use of an axial NHC ligand increases the reactivity of the Ru nitrene and carbene intermediates





Chan, K-H.; Guan, X.; Lo, V. K-Y.; Che, C-M. Angew. Chem. Int. Ed. 2014, 53, 2982

Metalloporphyrin catalyzed C–H amination

Amination using aryl azides as nitrogen source



Metalloporphyrin catalyzed C–H amination

Intramolecular C–H amination with arylsulfonyl azide



C–H amination using an engineered P450

■ Intramolecular C–H amination with aryIsulfonyl azide via "chemomimetic" enzyme modification



| Free Enzyme (0.1 mol%) | Turnovers | | %ее | |
|---------------------------|-----------|-----------|-----|--|
| Wild type P450 | 2.1 | | nd | |
| Modified P450 | 383 | | 73 | |
| | | | | |
| Enzyme (<i>E. Coli</i>) | % Yield | Turnovers | %ee | |
| Wild type P450 | 0.5 | 5.1 | nd | |
| Modified P450 | 58 | 430 | 87 | |

Fisrt example of a highly active enzyme catalyst for C–H amination

McIntosh, J. A.; Coelho, P. S.; Farwell, C. C.; Wang, Z. J.; Lewis, J. C.; Brown, T. R.; Arnold, F. H. Angew. Chem. Int. Ed. 2013, 53, 9309

C–H halogenation via metalloporphyrin catalysis



Use of strongly donating axial ligands stabilizes M-OH species, and slows down radical recombination

Allows for substitution of fluoride onto metal center

C–H halogenation via metalloporphyrin catalysis



C–H halogenation via metalloporphyrin catalysis



Liu, W.; Huang, X. Y.; Cheng, M. J.; Nielsen, R. J.; Goddard, W. A.; Groves, J. T. Science 2012, 337, 1322

C-H halogenation via metalloporphyrin catalysis







Supramolecular complexes as enzyme mimics

"Artificial Enzymes" designed around an active site linked to a supramolecular scaffold



- Structures contain well defined binding pockets capable of stabilizing reactive intermediates
- Artificial enzymes display a range of binding modes, including H bonding, $\pi \pi$ Interactions, etc
- Reactions isolated from the surrounding environment, allowing for enzyme-like selectivity

Raynal, M.; Ballester, P. Vidal-Ferran, A.; van Leeuwen, P. W. N. M. Chem. Soc. Rev. 2014, 43, 1734

Supramolecular complexes as enzyme mimics

A range of scaffolds have been utilized as host structures for supramolecular enzyme models



Cyclodextrins as a scaffold for substrate binding

Cyclodextrin family of compounds consists of a range of cyclic oligosaccarides



 β -Cyclodextrin

Conical structure

Hydrophobic cavity can accomodate a number of hydrophobic guest molecules

A numbe of isomers readily available, subject of many early reports on artificial enzymes

Catalytic systems using the cyclodextrin moiety



Initial report of an enzyme mimic

First description of an "artificial enzyme" for the hydrolysis of p-nitrophenyl acetate



Catalyst consists of metal complex covalently linked to β -cyclodextrin scaffold

Hydrolysis shows a 4x rate enhancement compared to the free metal complex alone

Rate acceleration attributed to substrate binding within hydrophobic cyclodextrin pocket

Artificial P450 enzyme for Stereoid Hydroxylation

Artificial Cytochrome P450 enzyme to mimic selectivity seen in steroid biosynthesis



Catalyst system based on metalloporphyrin core with appended cyclodextrin rings



Ester side chains required to facilitate binding to cyclodextrin groups on catalyst and control regioselectivity

Selective oxidation observed at C-6 position, without overoxidation to ketone

Artificial P450 enzyme for Stereoid Hydroxylation

Artificial Cytochrome P450 enzyme to mimic selectivity seen in steroid biosynthesis



Second generation catalyst contains fluorinated aromatic to prevent oxidative degradation



quantitative yield

Selective oxidation observed with increased efficiency at C-6 position, without overoxidation to ketone

Substrates without both ester side chains give a mix of products

Artificial P450 enzyme for Stereoid Hydroxylation

Artificial Cytochrome P450 enzyme to mimic selectivity seen in steroid biosynthesis



Second generation catalyst contains fluorinated aromatic to prevent oxidative degradation



Supramolecular enzyme mimic as an artificial peptidase

Peptide bond cleavage is a major biological process, vatalyzed by a number of enzymes



Internal X-Pro residues are difficult to cleave, with few enzymes that are able to do so



Supramolecular enzyme mimic as an artificial peptidase

Pd aqua complexes found to selectively cleave all X-Pro linkages

Arg-Pro-Pro-Gyl-Phe-Ser-Pro-Phe-Arg
$$(Pd(H_2O)_4)^{2+}$$
 Arg-Pro Pro Gyl-Phe-Ser Pro-Phe-Arg
Arg-Pro Pro Gyl-Phe-Ser Pro-Phe-Arg
Arg-Pro Pro Gyl-Phe-Ser Pro-Phe-Arg
However, sequence specific peptide avage often required for biochemical applications



Cyclodextrin moiety intended to bind aromatic side chains Thus specific to X-Pro-Ar sequences (Phe, Tyr and Trp)

Milovic, N. M.; Badjic, J. D.; Kostic, N. M. J. Am. Chem. Soc. 2004, 126, 696

Supramolecular enzyme mimic as an artificial peptidase

Pd aqua complexes found to selectively cleave all X-Pro linkages





Non-covalent self assembled molecules have been used as "enzyme mimics" to encapsulate reactions



Container molecules made up of non-covalent linkages, eg. H-bonding, metal coordination, etc.

Well defined binding pocket, but present some flexibility when incorporating guest complexes

Easily prepared by combining components which assemble through complementary interactions

Directing regioselectivity of Diels-Alder reaction by M₆L₄ molecular cage



Diels-Alder of anthracene and phthalimide proceeds to yield adduct bridging at the center ring



Proposed to use host complexes to override the selectivity of uncatalyzed D-A reaction

Directing regioselectivity of Diels-Alder reaction in molecular hosts



Use of supramolacular catalyst found to override selectivity in favor of terminal D-A adduct



Substrates without bulky group on phthalamide found to give bridging product

Directing regioselectivity of Diels-Alder reaction in molecular hosts



Regioselectivity explained via fixed orientation of guest molecules prior to reaction



Bent structure of product leads to a lower affinity for host than substrates, allowing catalytic turnover



M₄L₆ have a smaller aperture and an interior more segregated from bulk solution than M₆L₄

- Encapsulation occurs with entropically favorable liberation of solvent molecules
- Anionic host framework has a stabilizing effect on cationic guests and intermediates



Control of the rate of a step of a catalytic cycle via encapsulation rather than via ligand

Merger of biomimetic envioronment control with unnatural mode of reactivity

Kaphan, D. M.; Levin, M. D.; Bergman, R. G.; Raymond, K. M. Toste, F. D. *Science* **2014**, *350*, 1235 Levin, M. D.; Kaphan, D. M.; Hong, C. M.; Bergman, R. G.; Raymond, K. M.; Toste, F. D. *J. Am. Chem. Soc.* **2016**, *138*, 9682

Stoichiometric reductive elimination from dimethyl Au complex



Background reactivity measured using strongly binding Et₄P⁺ to clock binding pocket

Proposed mechanismbased on pre- equilibrium between neutral and cationic substrates:



Kinetic invesigation indicates enzyme like mechanism following Michaelis-Menten like kinetics

Kaphan, D. M.; Levin, M. D.; Bergman, R. G.; Raymond, K. M. Toste, F. D. *Science* **2014**, *350*, 1235 Levin, M. D.; Kaphan, D. M.; Hong, C. M.; Bergman, R. G.; Raymond, K. M.; Toste, F. D. *J. Am. Chem. Soc.* **2016**, *138*, 9682

Applied to stoichiometric sp³-sp³ coupling using a Pt catalyst



Efficient coupling observed only with both catalysts, deuteration indicates both starting materials incorporated



Kaphan, D. M.; Levin, M. D.; Bergman, R. G.; Raymond, K. M. Toste, F. D. *Science* **2014**, *350*, 1235 Levin, M. D.; Kaphan, D. M.; Hong, C. M.; Bergman, R. G.; Raymond, K. M.; Toste, F. D. *J. Am. Chem. Soc.* **2016**, *138*, 9682



