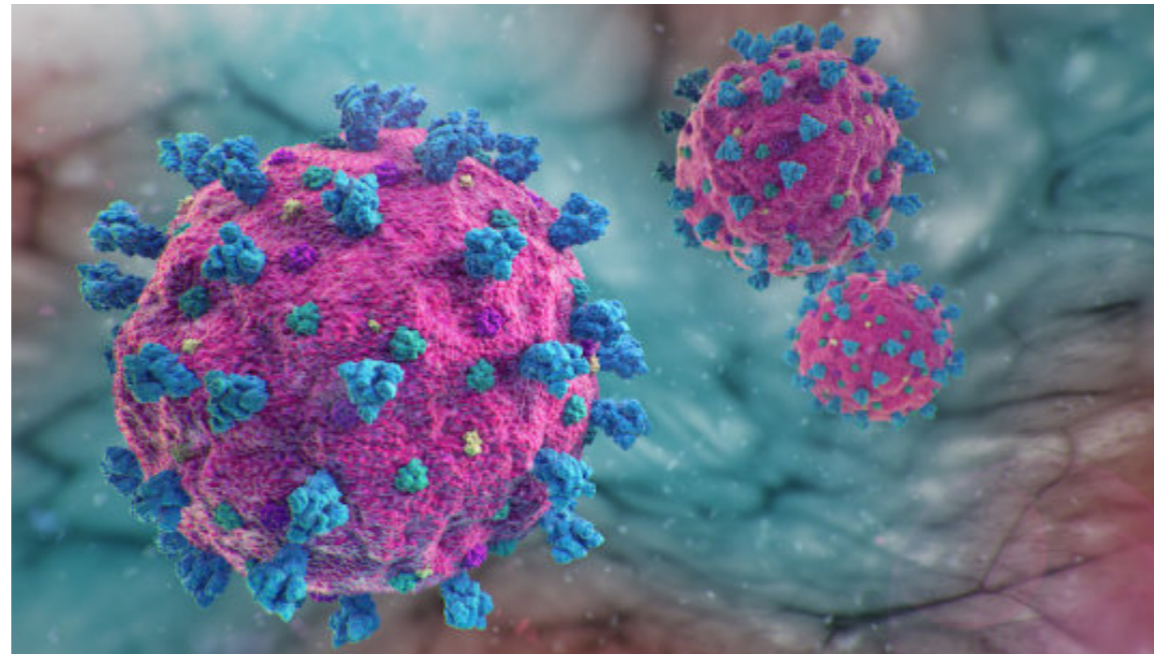


Viral Entry

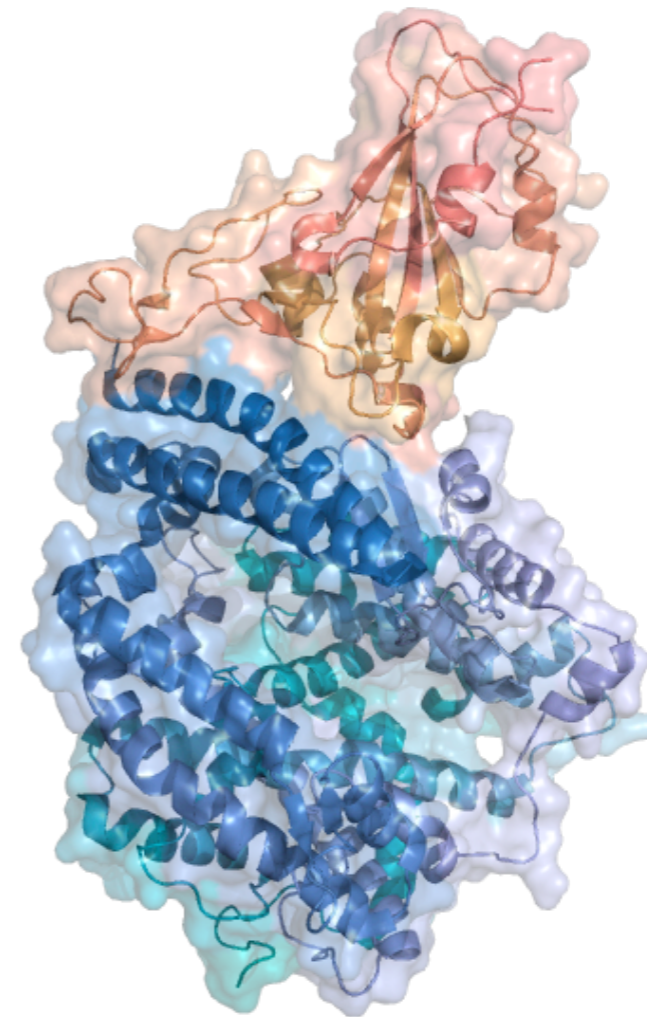


*MacMillan Group Meeting
May 04, 2020*

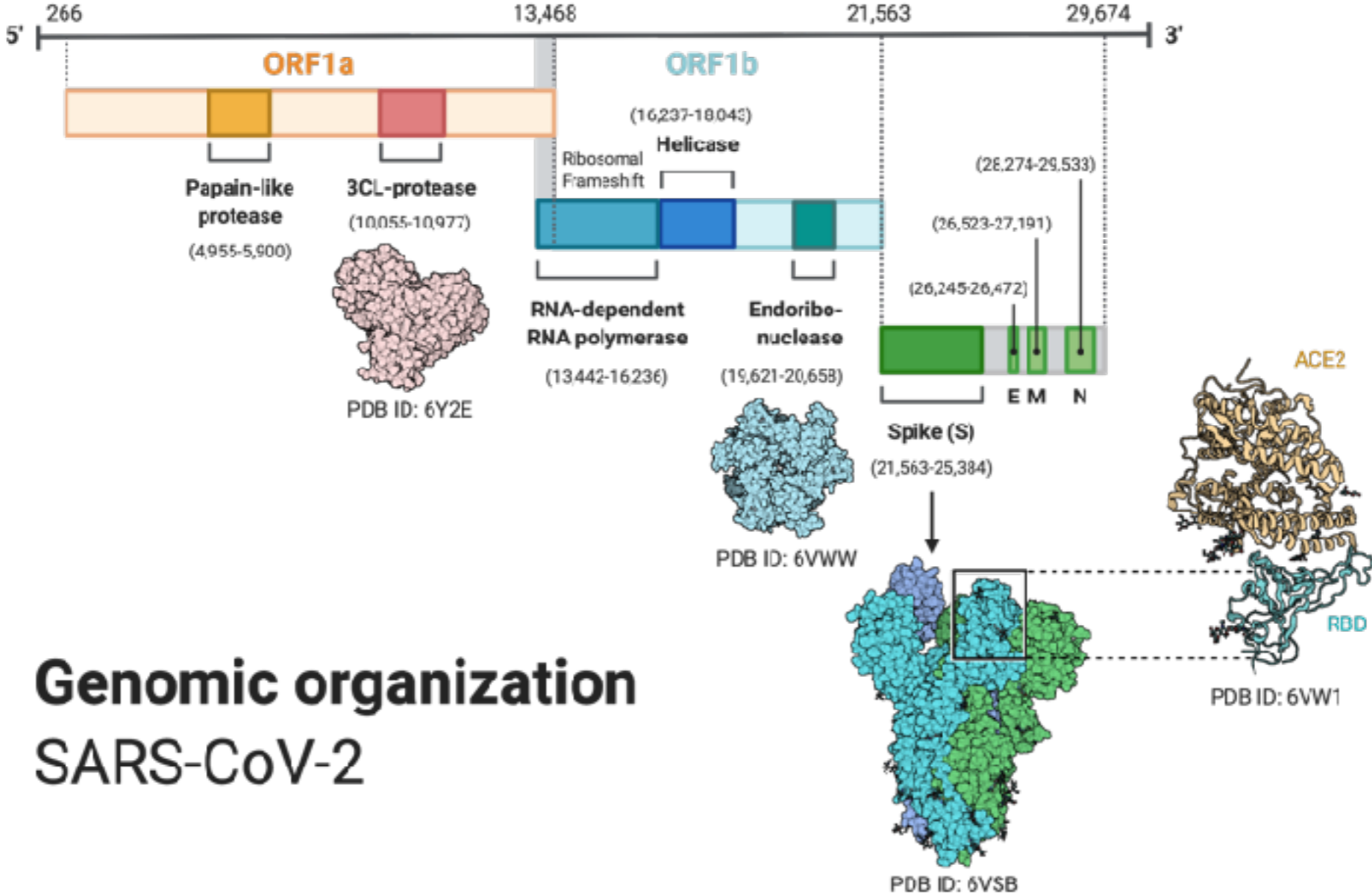
Daniel Kim

Outline

- Brief Overview of Viruses & Nomenclature
- General Mechanisms of Viral Entry
- Important Factors for Successful Viral Entry
 - ▶ Attachement
 - ▶ Signaling
 - ▶ Endocytosis
 - ▶ Penetration
 - ▶ Uncoating
- Details Associated Towards SARS-CoV-2 & Other Examples



Virus Overview



**Genomic organization
SARS-CoV-2**

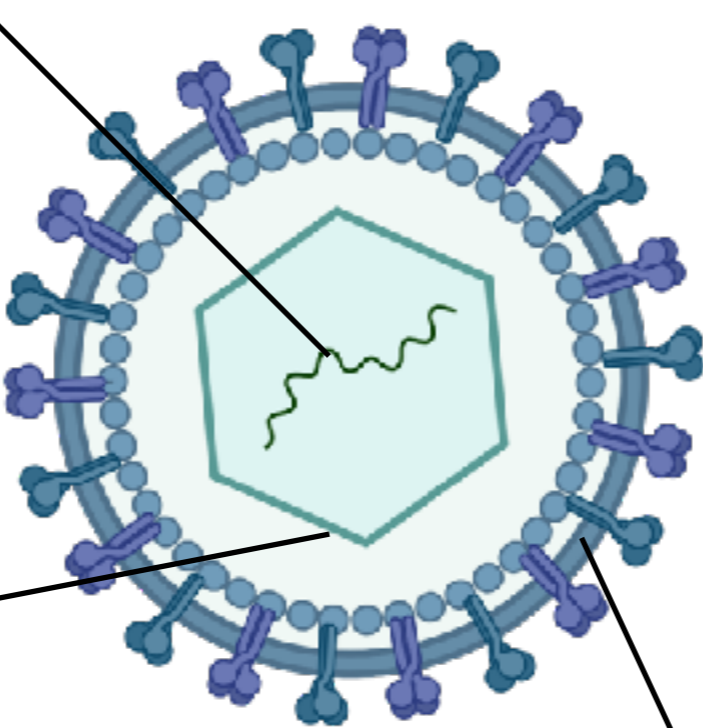
Virus Overview

DNA or RNA

the genetic information of the virus responsible for viral replication

Baltimore Classification System

simplified virus classification based on the viral use of nucleic acid



Capsid Shell

tightly packs nucleic information

Lipid Bilayer Membrane

found only in envelope viruses & contains additional virally encoded proteins

Virus Overview

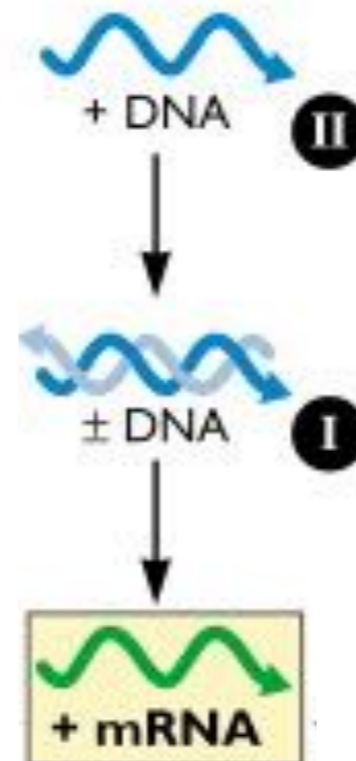
Baltimore Classification System



mRNA encodes for proteins

Virus Overview

Baltimore Classification System

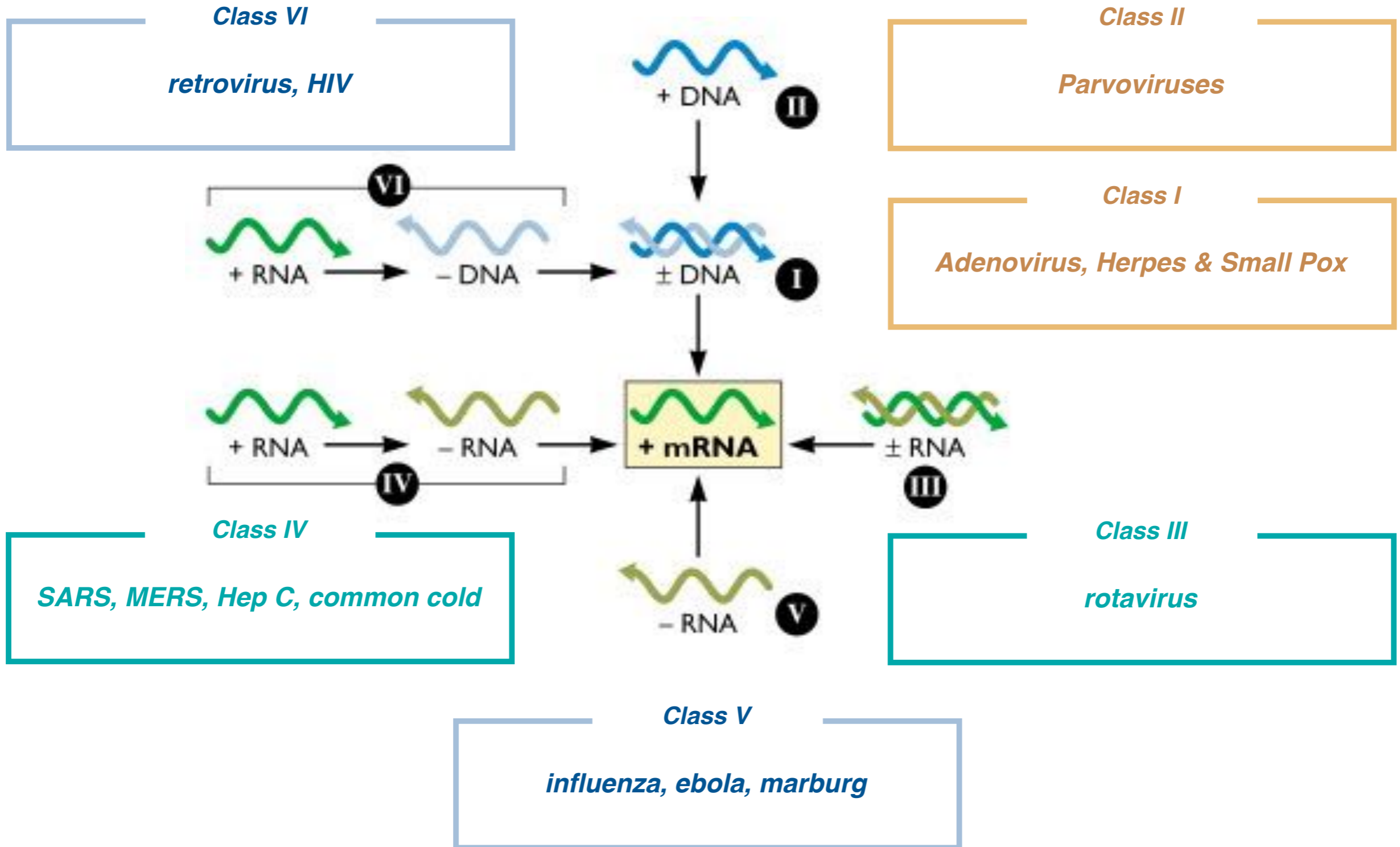


Transcription

RNA polymerase copies DNA sequence into RNA

Virus Overview

Baltimore Classification System



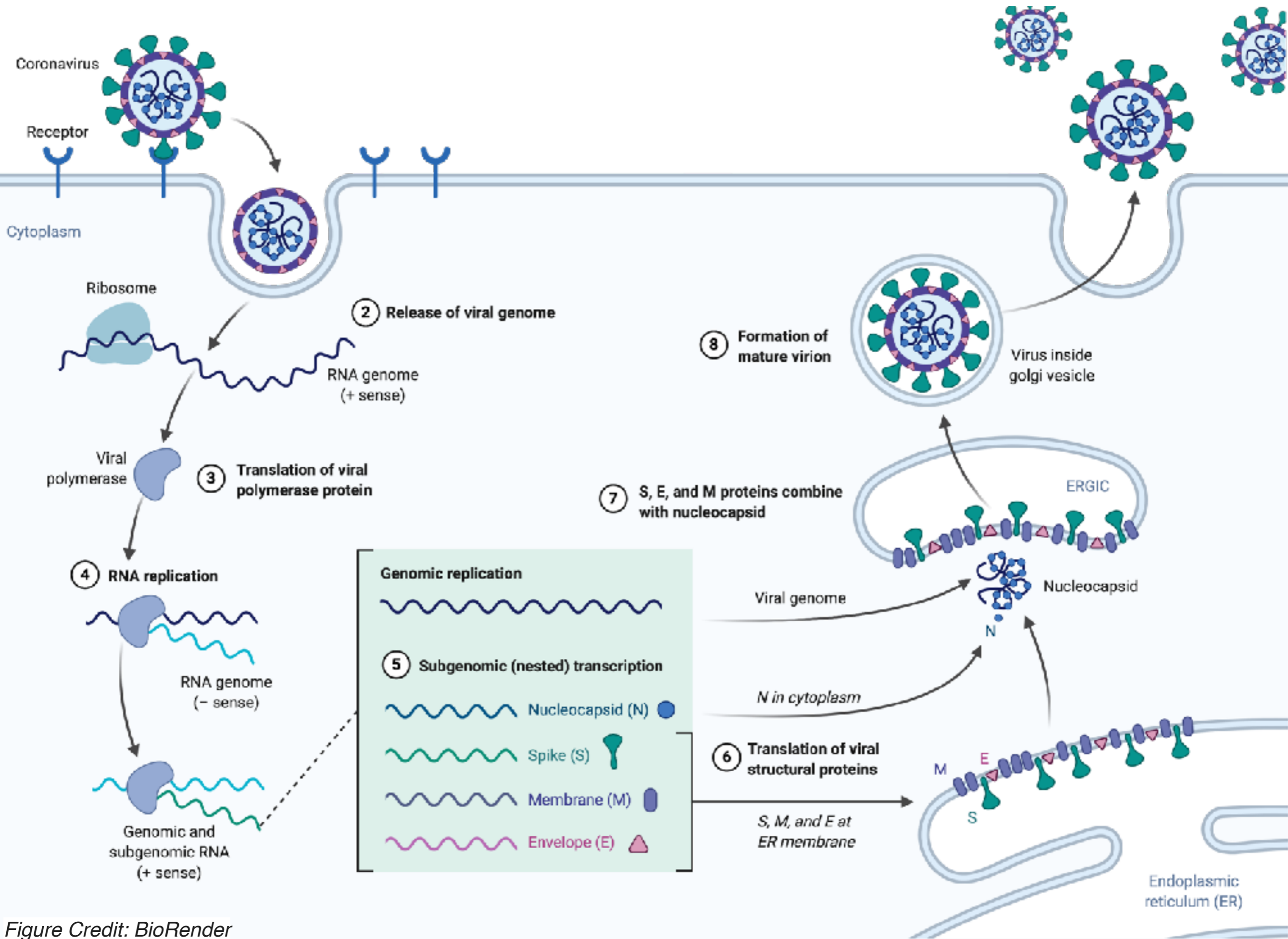
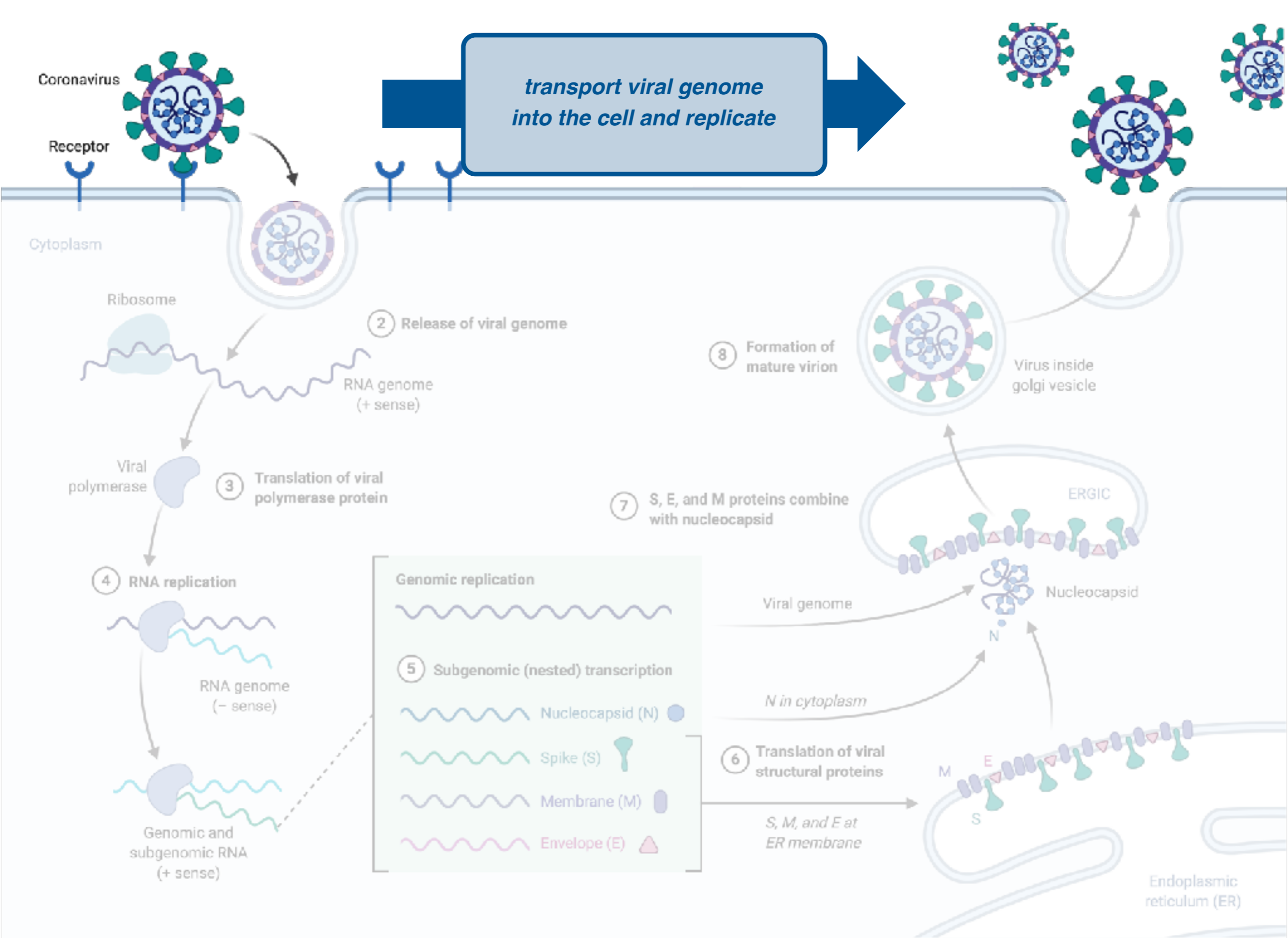
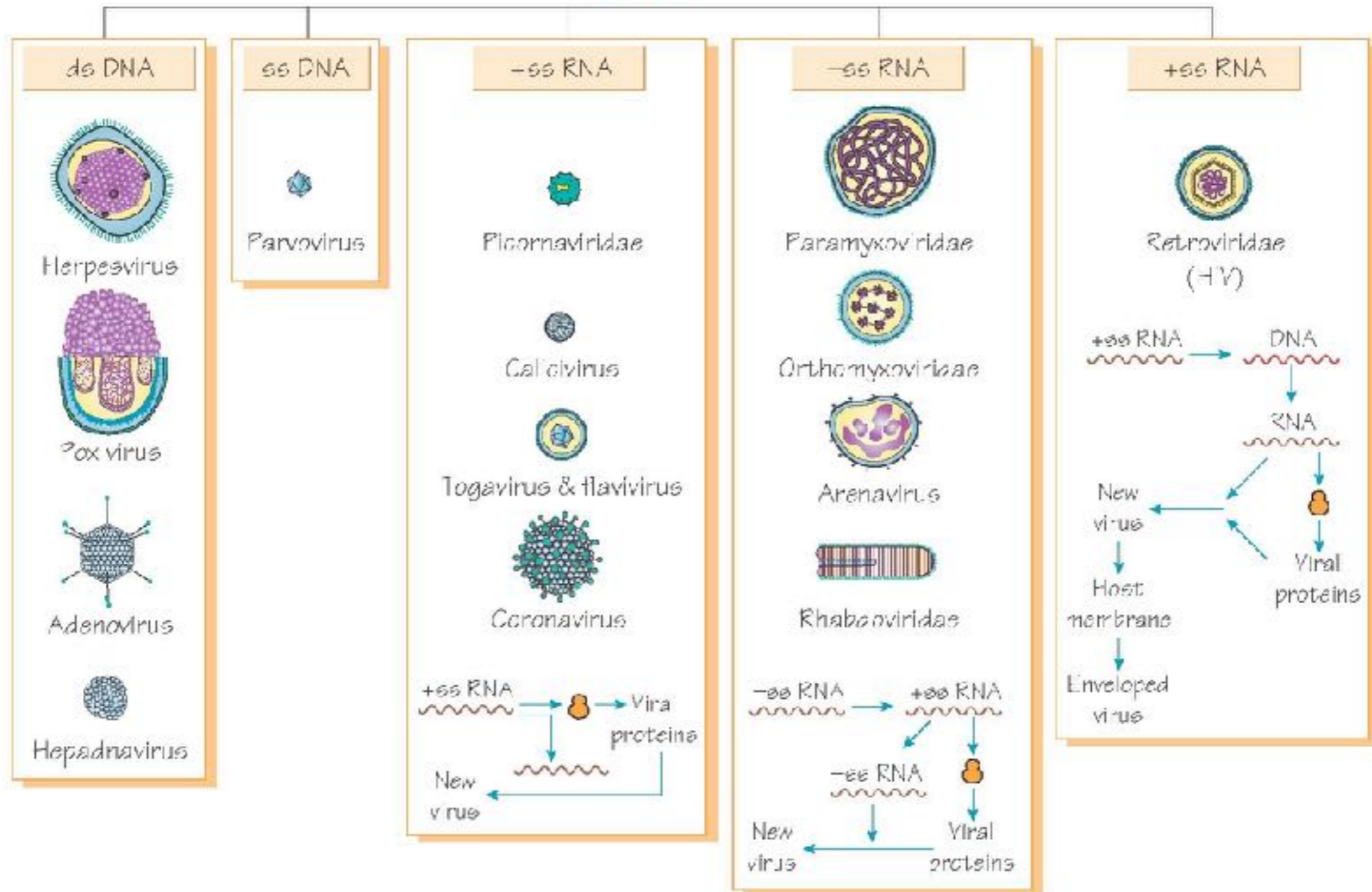


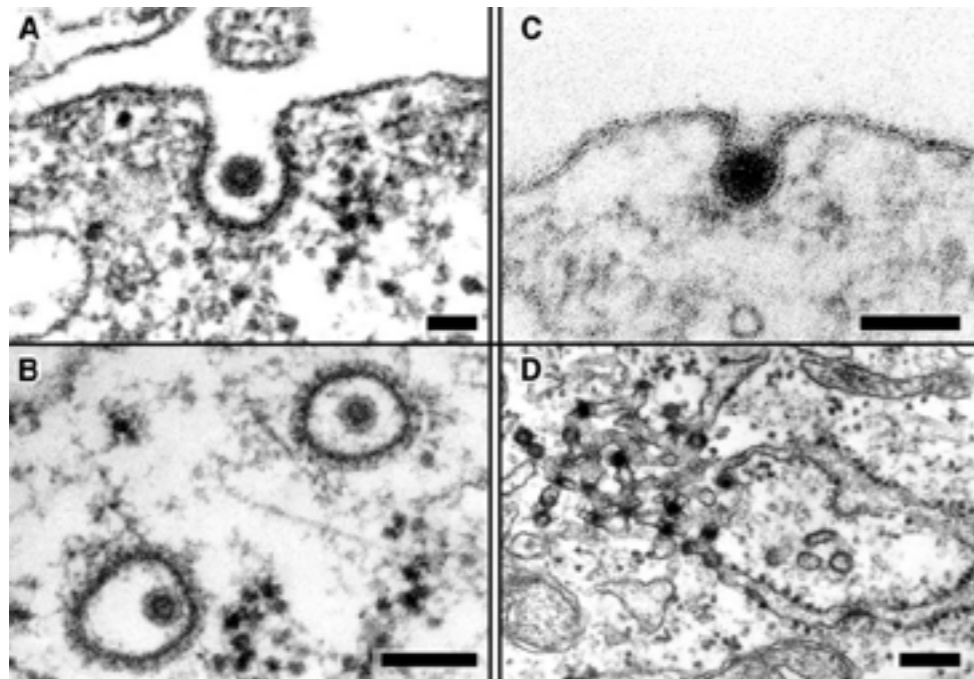
Figure Credit: BioRender



Examples of Different Classes of Viruses



Challenges of Understanding Viral Entry



Understanding Size

× 1,000,000 size magnification

virus
orange

animal cell
circus tent

Techniques

light & electron microscopy

in vitro modeling

perturbations via

chemical inhibitors
mutant cells or viruses
siRNA silencing

Expertise

virology

cell & molecular biology

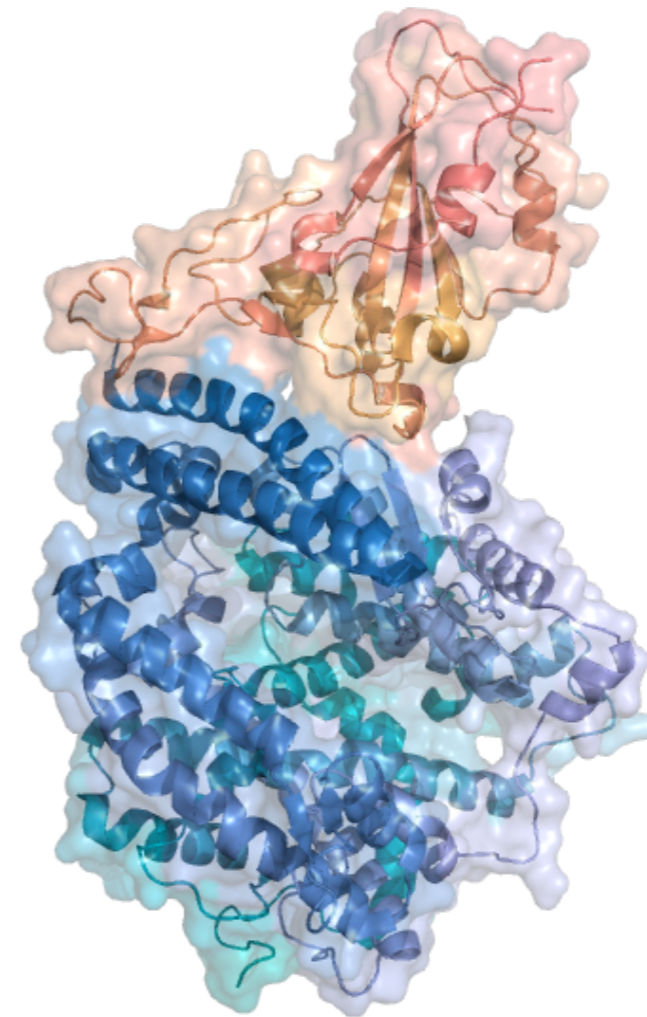
biochemistry & biophysics

systems biology

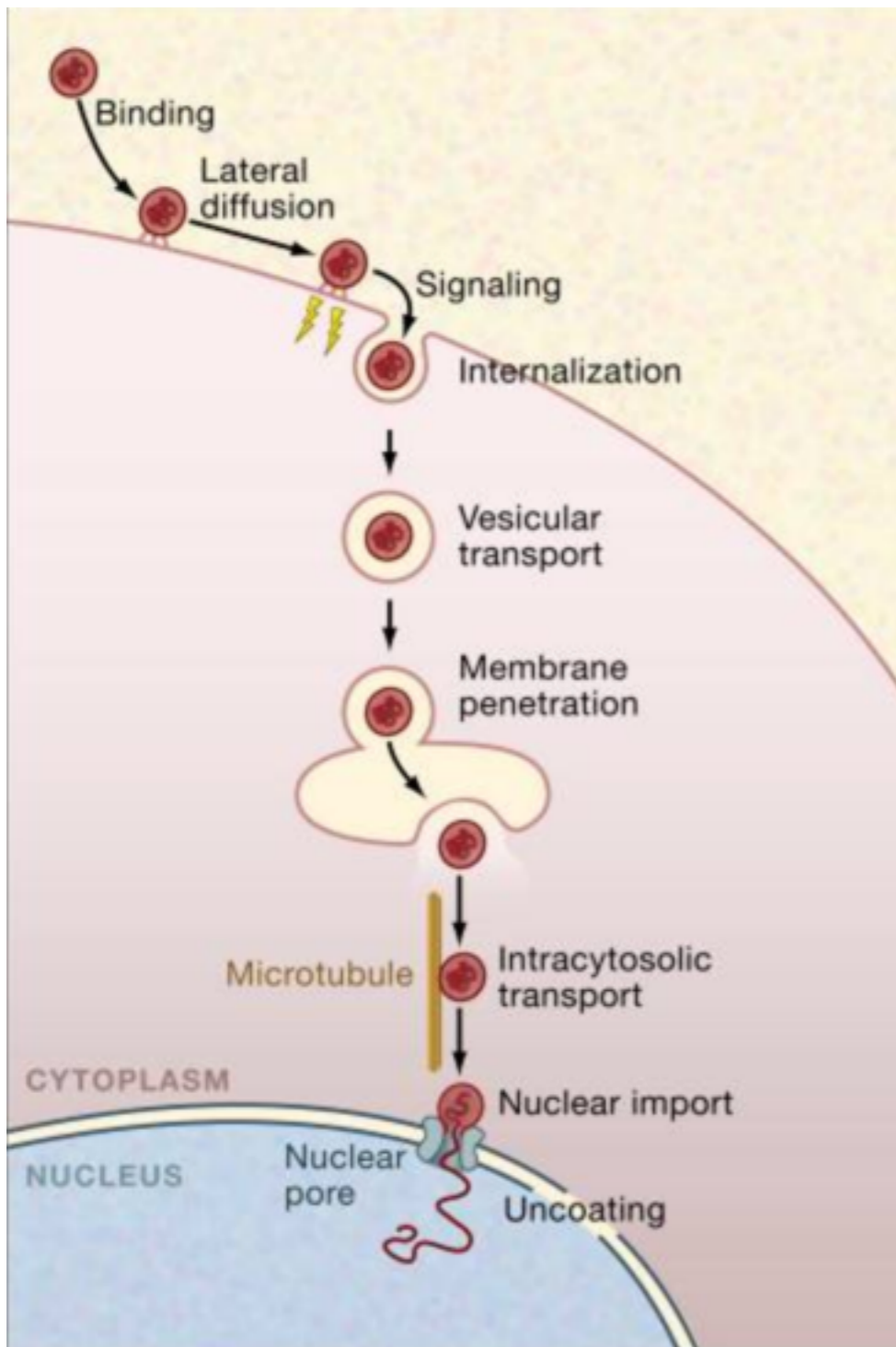
computer science

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Viral Entry Overview



General Lessons Learned

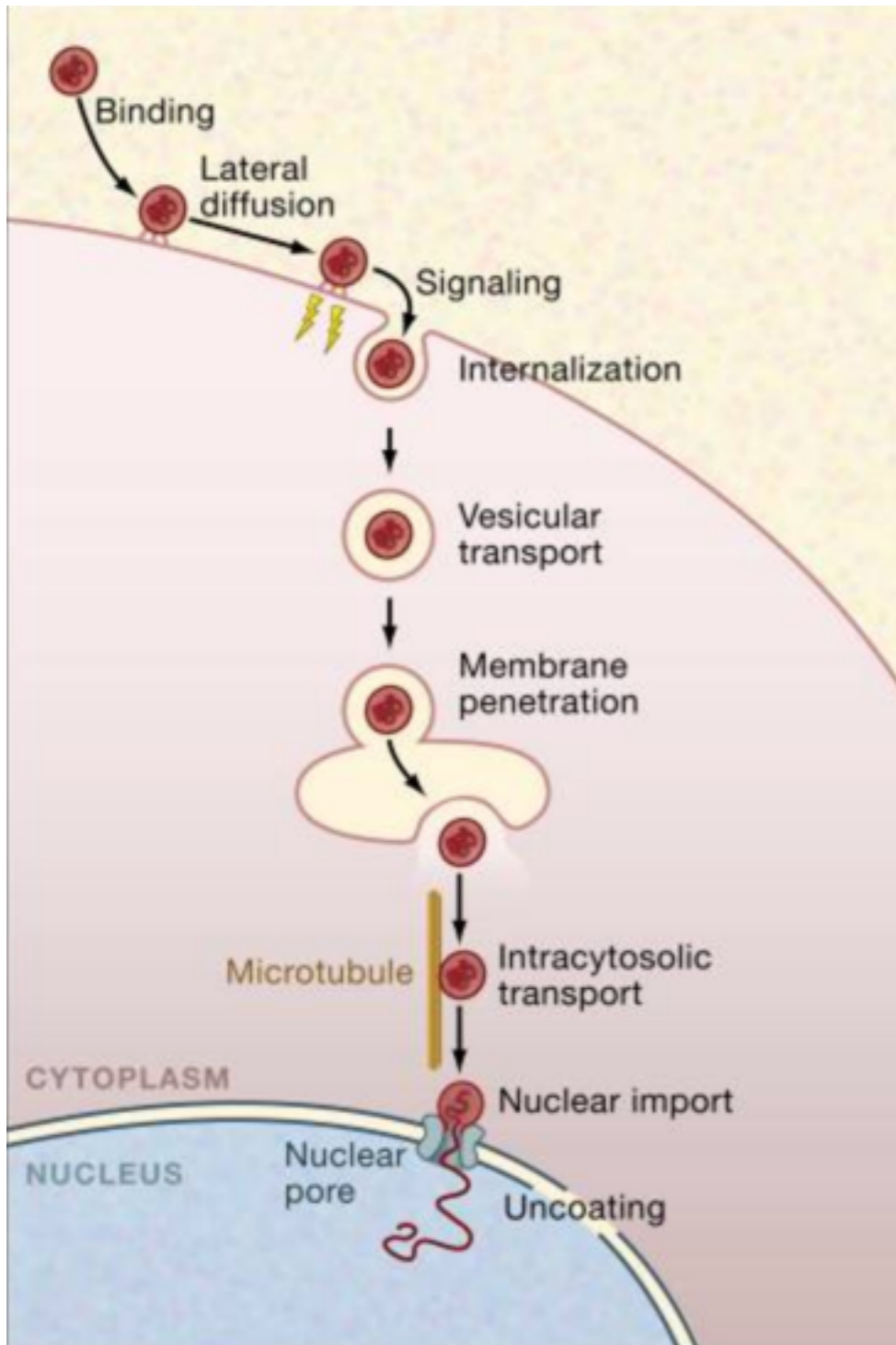
*entry into the cell is a process of multiple steps
not simple, very complex system*

uncoating: built-in program (passive)

virus proteins are meta-stable

proteins respond to cellular cues

Viral Entry Overview



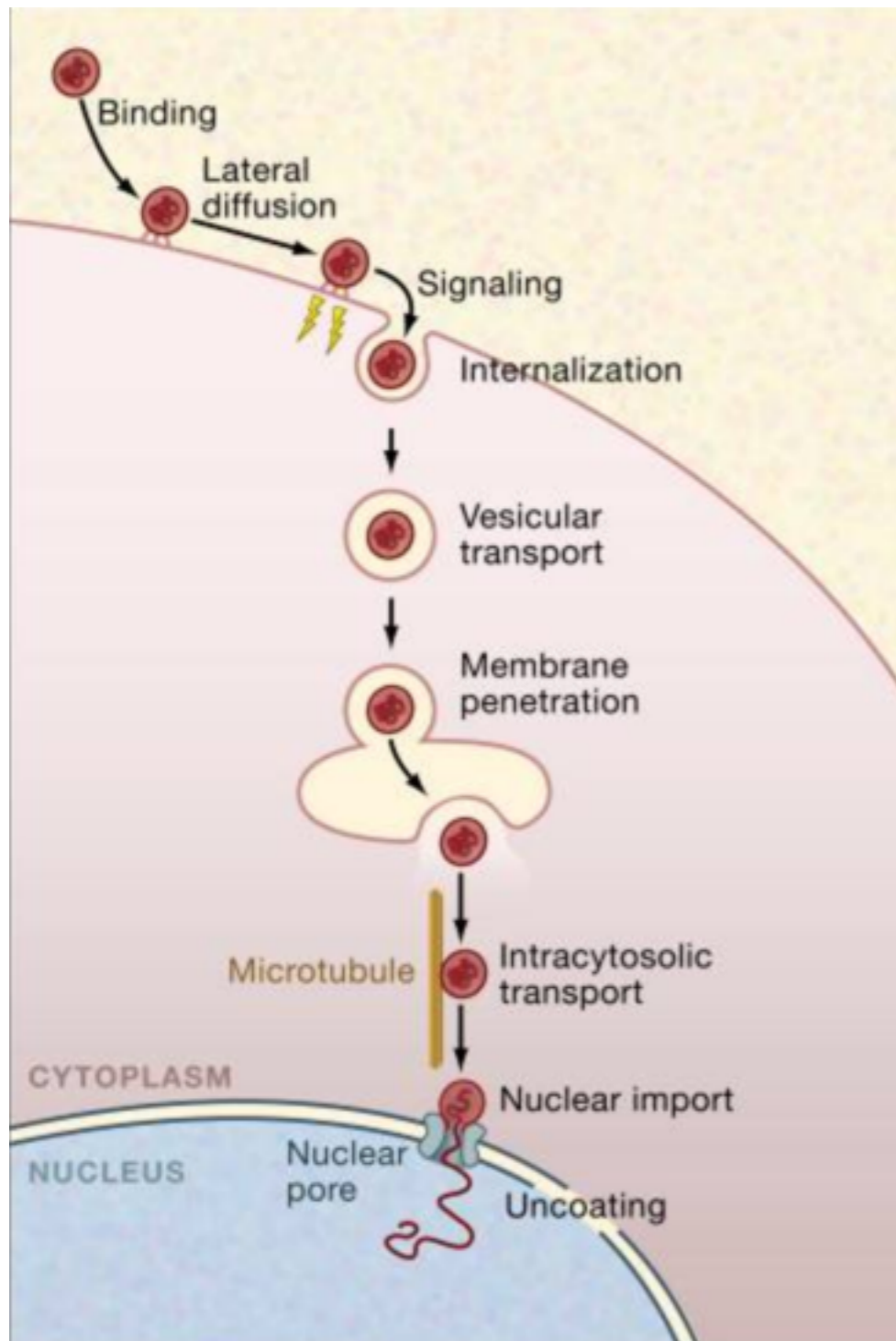
General Lessons Learned

*cellular processes & cellular factors
are critical components*

viruses activate cellular signaling pathways

viruses 'speak' the language of the cell

Cellular Cues for Viral Entry



what leads to stepwise mechanisms?

binding to cell surface receptors

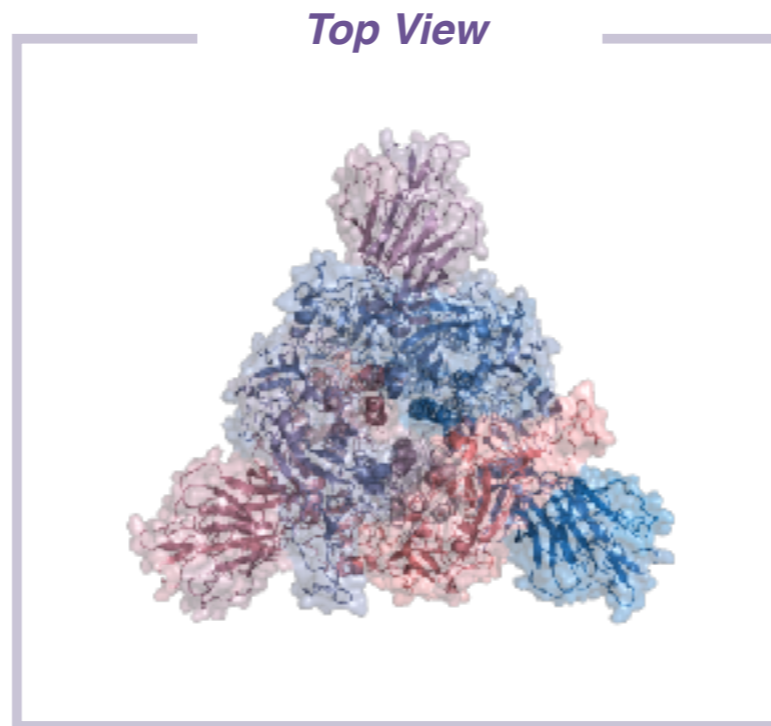
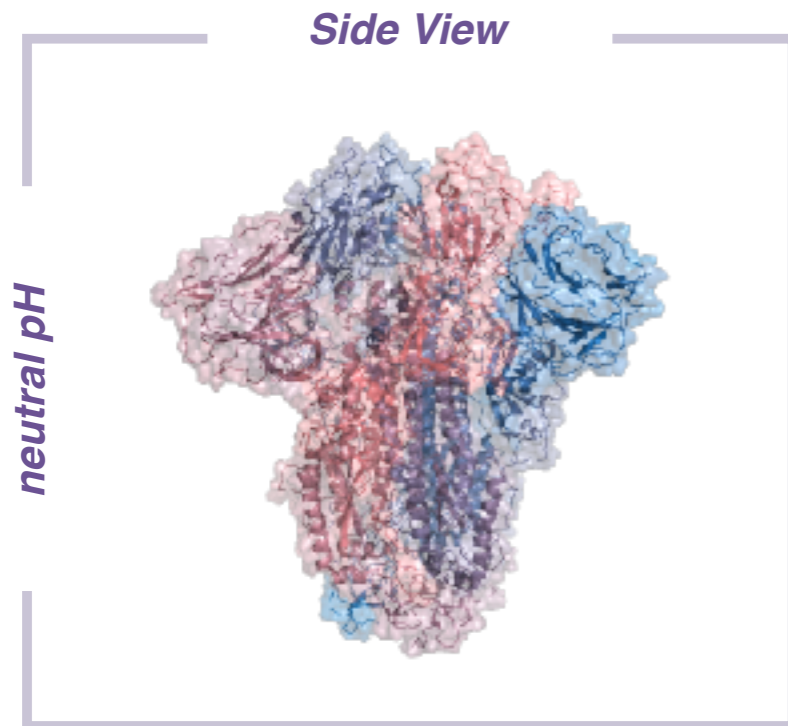
exposure to lower pH

cleavage by cellular proteases

re-entry into a reducing environment

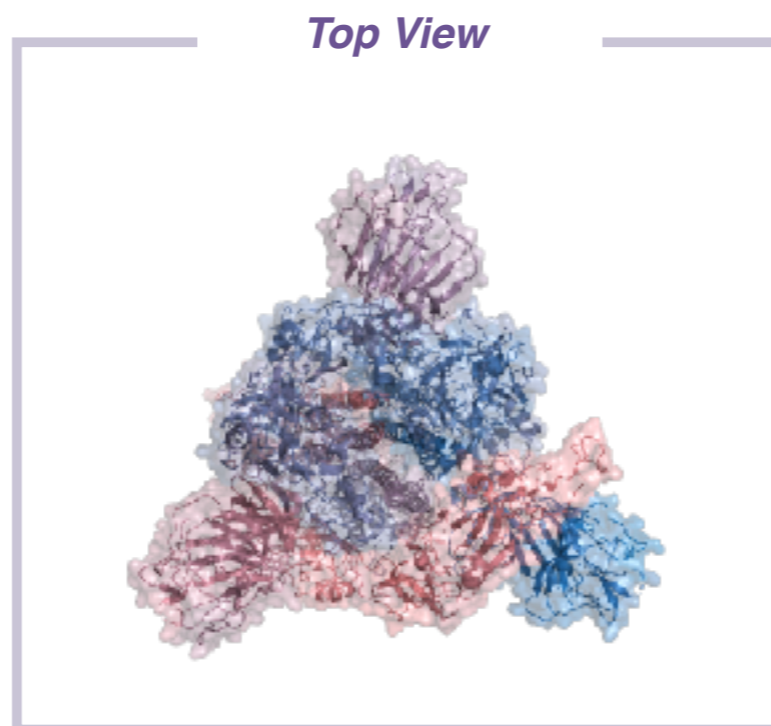
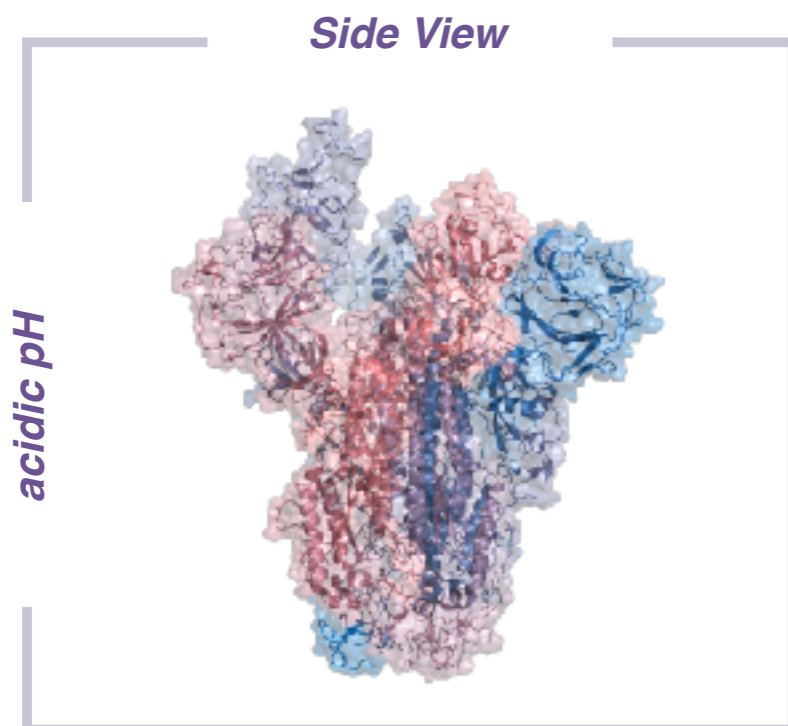
exposure to other enzymes (thiol-oxidoreductases)

Example of pH Response in SARS-CoV-2 Spike Protein



*spike protein in novel 2019
SARS-CoV-2 virus
PDB: 6ACC & 6ACD*

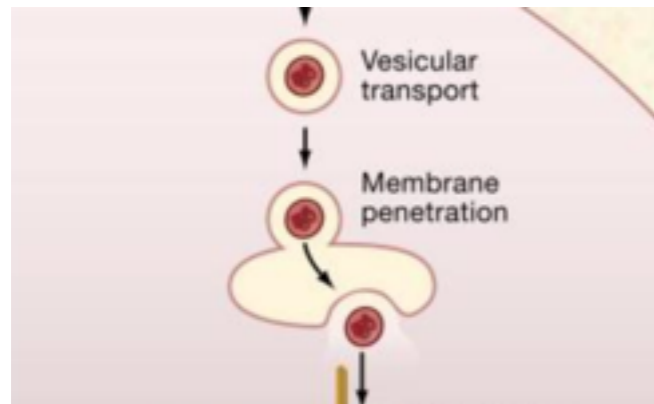
*endocytosis changes viral surroundings
to a more acidic (lower pH) environment*



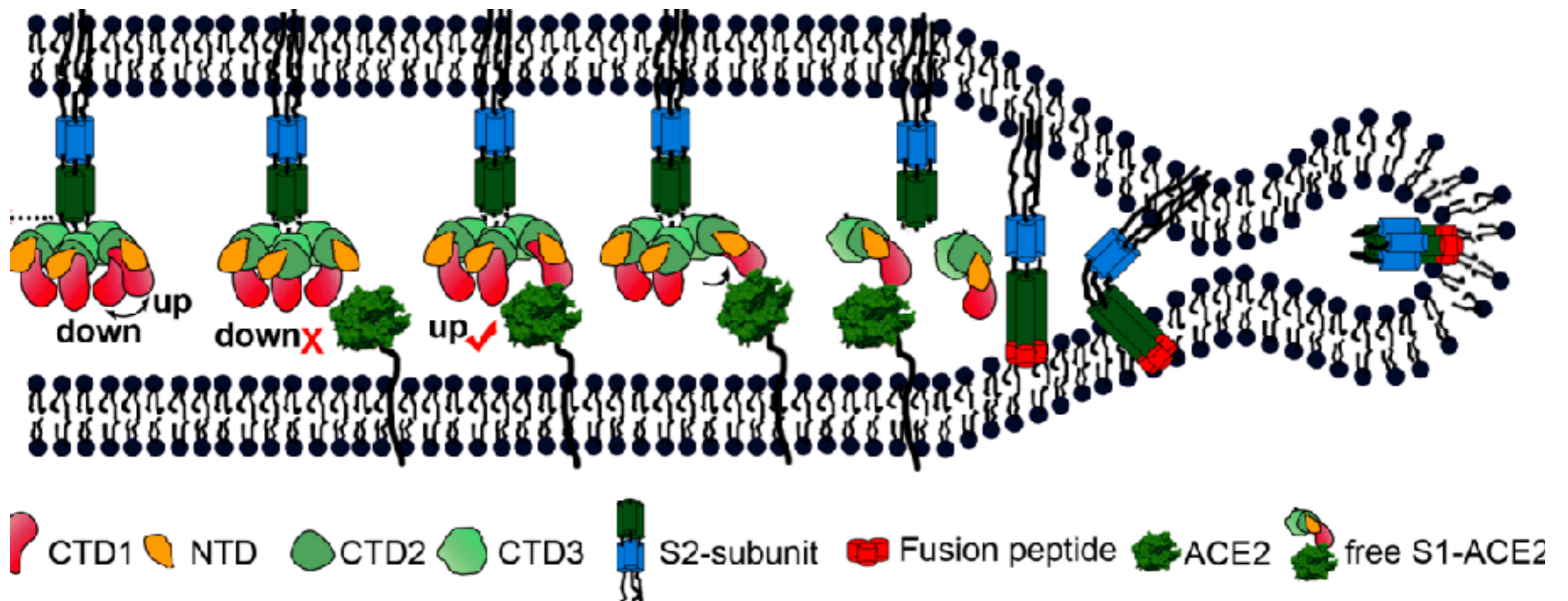
viral proteins exhibit meta-stable state

stepwise process for viral entry

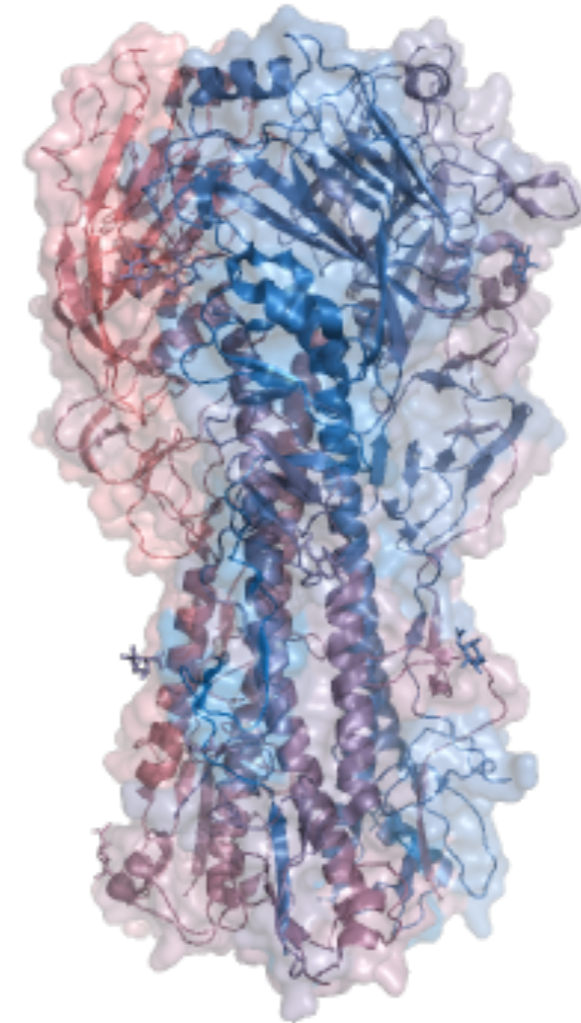
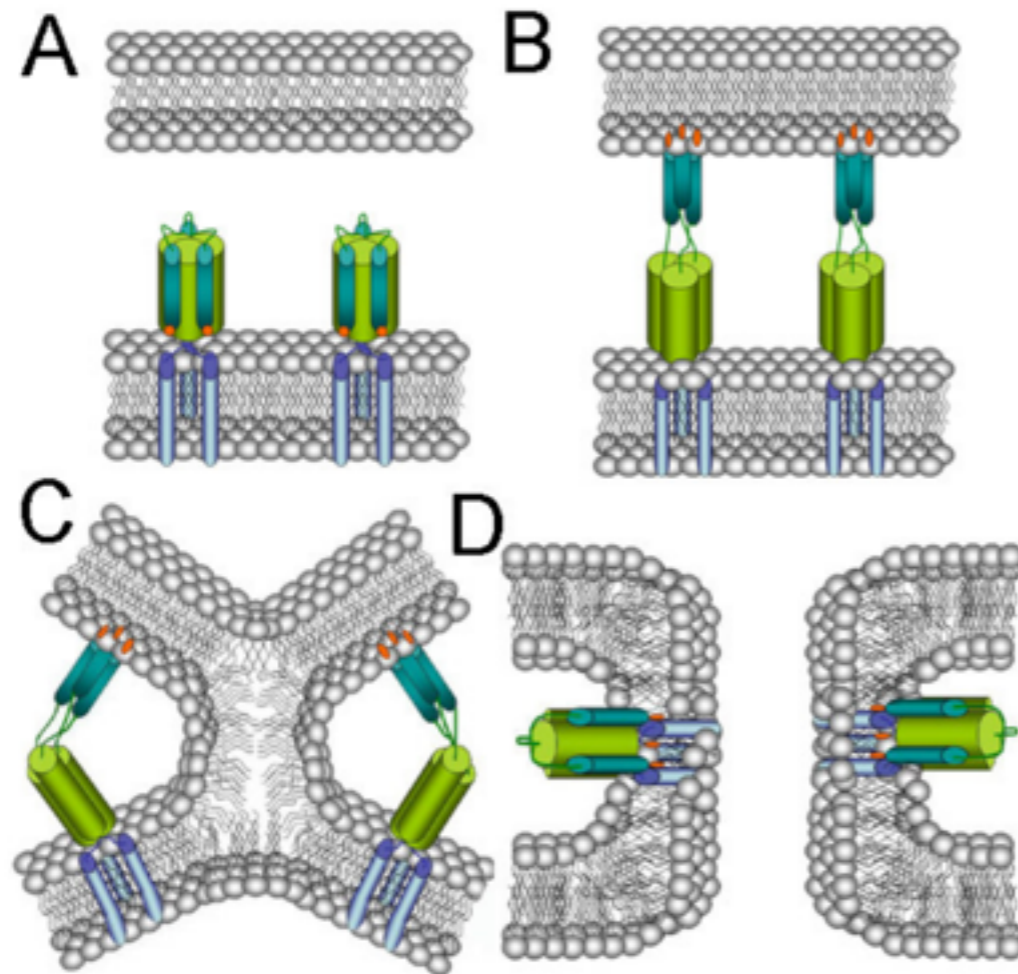
Example of pH Response in SARS-CoV-2 Spike Protein



large conformational changes in response to pH facilitates endocytosis

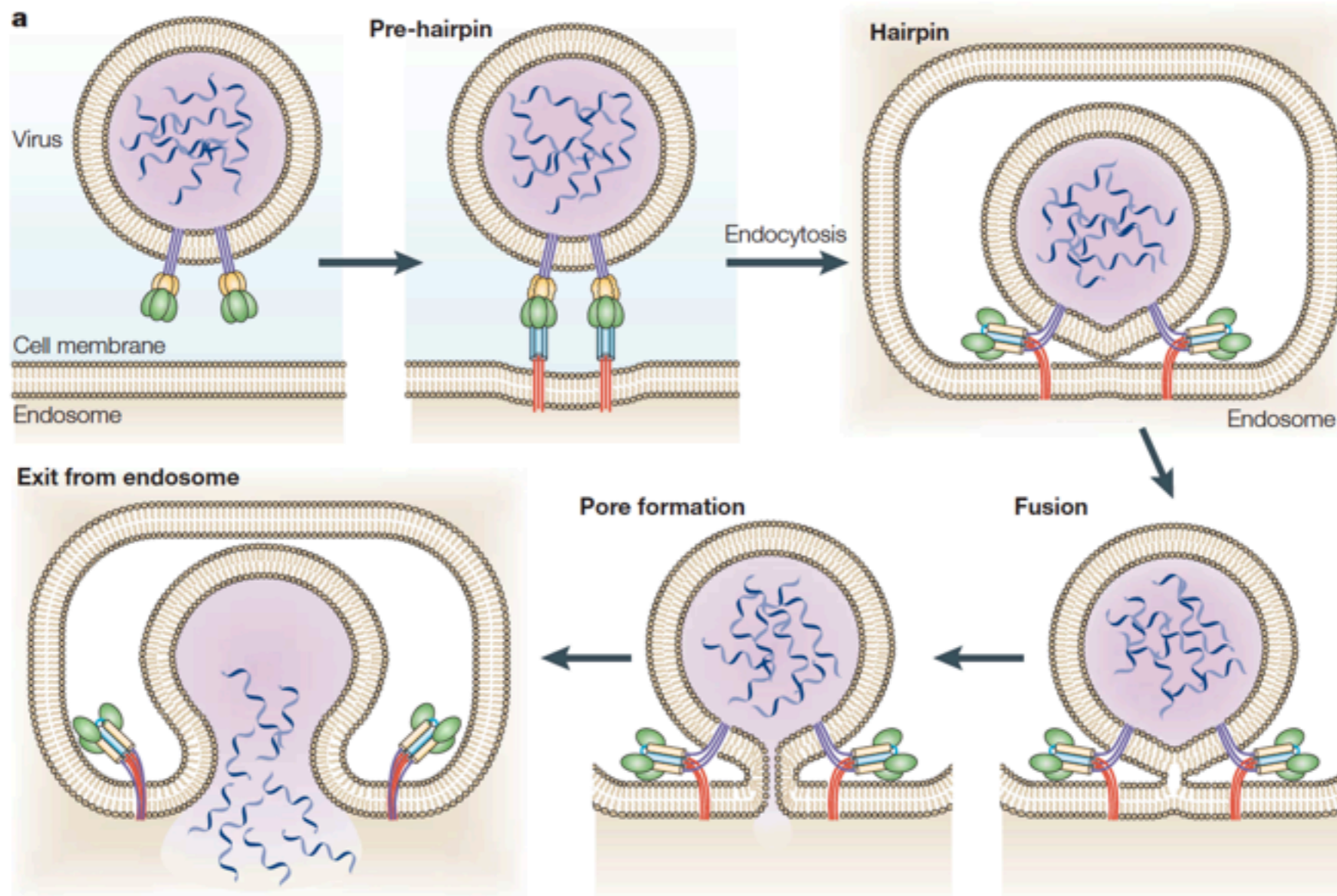


Example of pH Response in Influenza HA Protein



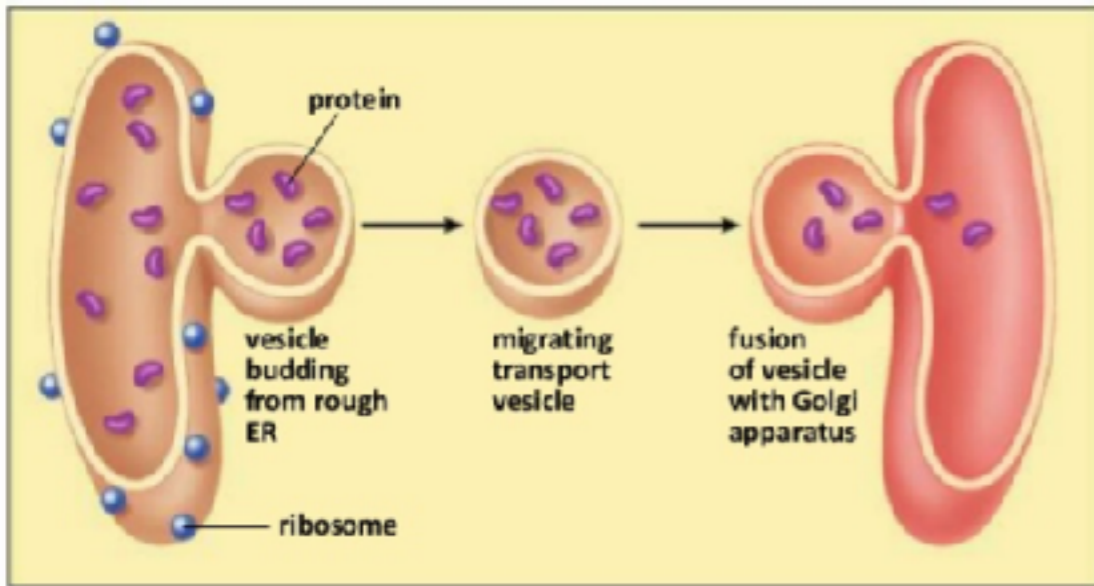
lipid bilayer glycoprotein
hemagglutinin PDB: 1ruz

Example of pH Response in Influenza HA Protein



Enveloped vs Non-Enveloped Viruses

Vesicle transport



enveloped viruses use lipid bilayer

similar mechanism as a vesicle

mechanism to transport from infected cell to uninfected cell

Infected Cell



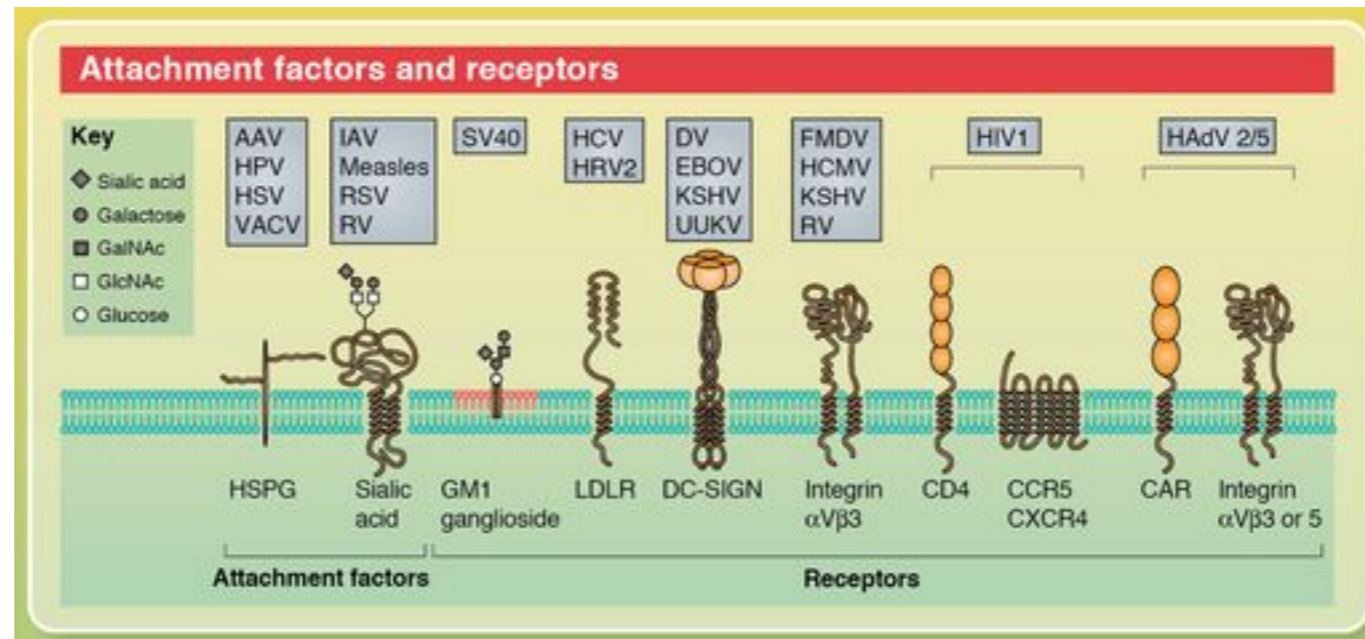
Uninfected Cell

budding

fusion

Binding: Viral Proteins & Cellular Receptors

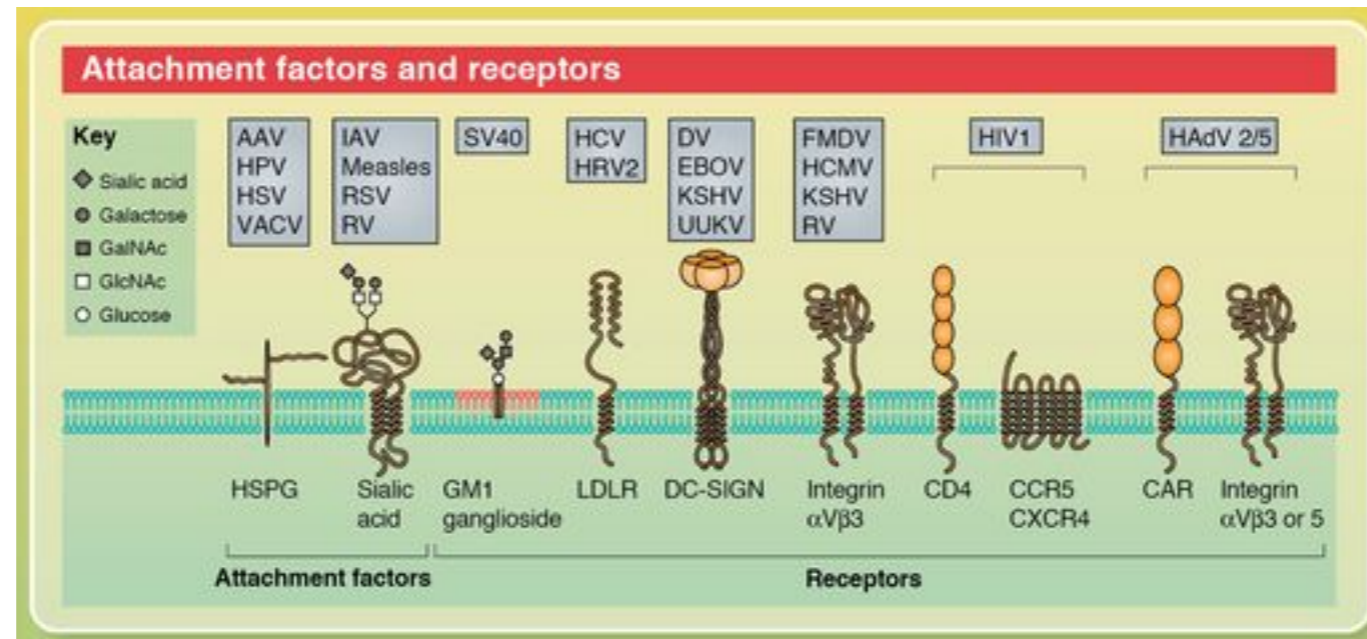
a virus cannot infect a cell it cannot bind to



*attachement factors: simply bind the virus
(to increase concentration of viral particles on cell surface)*

Binding: Viral Proteins & Cellular Receptors

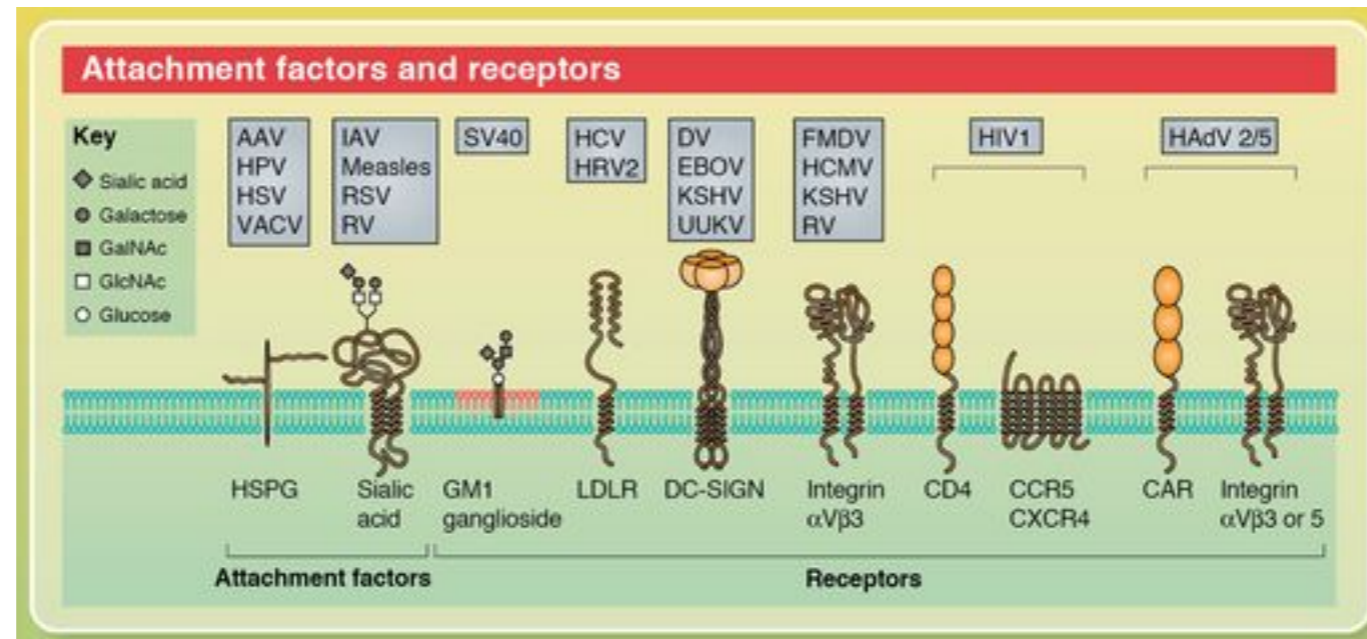
a virus cannot infect a cell it cannot bind to



*receptors: bind virus & provides additional signaling or viral conformational changes
(mechanisms vary, but can also start the endocytosis process)*

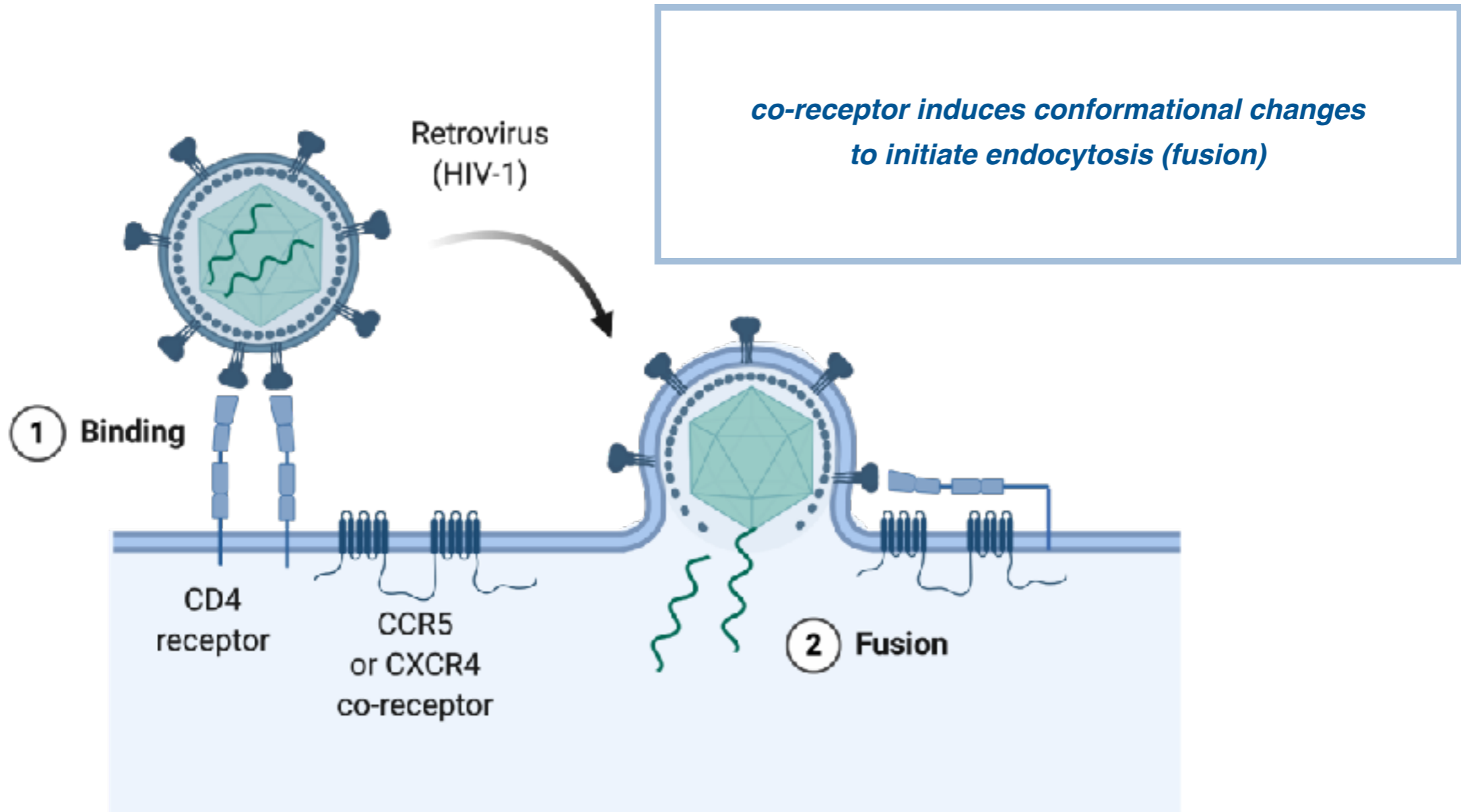
Binding: Viral Proteins & Cellular Receptors

a virus cannot infect a cell it cannot bind to



*important note: most viruses are known to have more than one type of receptor binding they can also be **multivalent** (binding to multiple receptor or attachment factors at the same time)*

HIV-1 is Multivalent & Uses Multiple Receptors



Examples of Viruses & Their Target Receptors

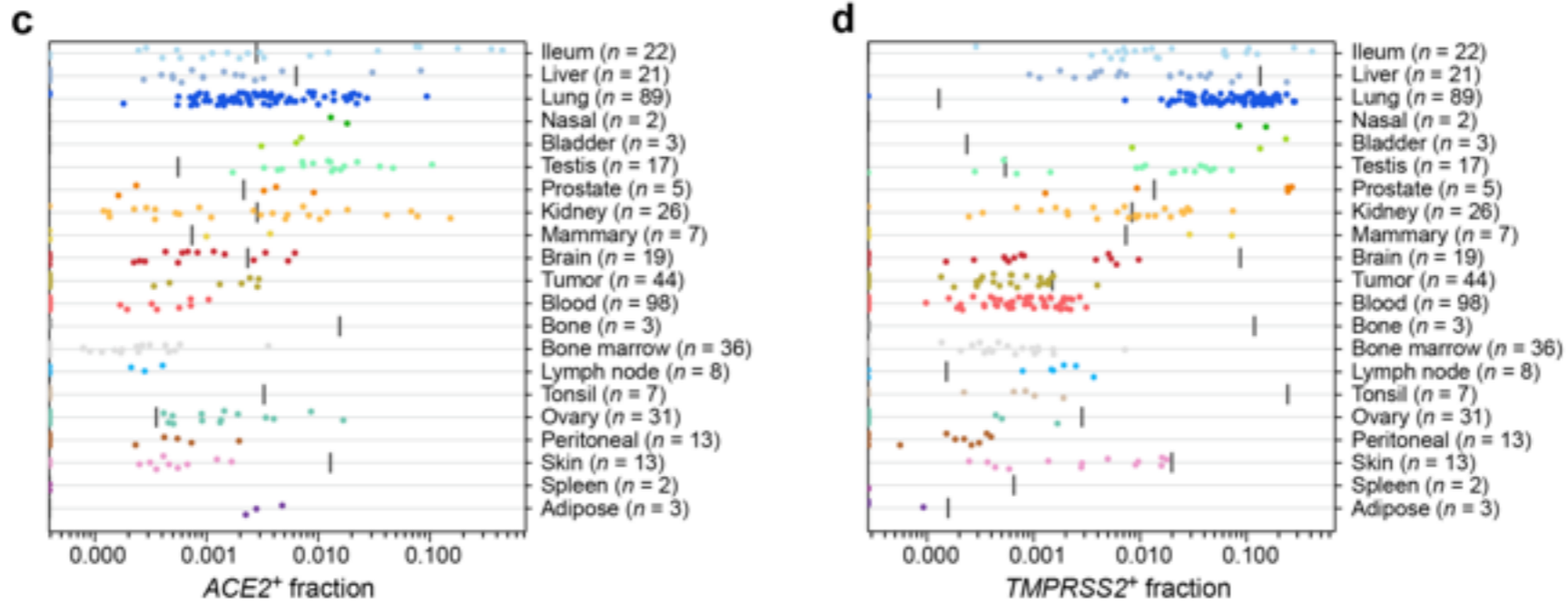
Virus	Entry protein*	Receptor †	Co-receptor	Alternative receptor §	Notes
Influenza A	Haemagglutinin	Sialic acid (mM)	Unknown	unknown	There are no indications that influenza needs co-receptor(s) for entry
HIV-1	gp160 (gp120)	CD4 (nM)	CCR5, CXCR4, other (nM– μ M)	Galactosyl ceramide (μ M)	Some HIV-1 isolates are CD4-independent and can use CCR5 or CXCR4 as receptors; affinities for co-receptors are higher in the presence of CD4; entry in the absence of CD4 is typically much less efficient.
SARS-CoV	S (S1)	ACE2 (nM)	Unknown	Unknown	
Herpes simplex virus 1 (HSV-1)	Glycoprotein D (gD)	HveA (μ M)	Unknown	Unknown	Several other viral (gB, gC, and the heterodimer gH/gL) and cellular (heparin sulphate, nectin-1) receptors are implicated in the complex entry mechanism; a truncated form of gD exhibits 100-fold higher affinity for HveA
Poliovirus 1	Capsid shell (VP1, VP2, VP3)	CD155 (nM– μ M)	Unknown	Unknown	CD155 is the receptor for all three serotypes; affinities for cell surface receptors significantly differ from those for soluble receptors and are also temperature-dependent.
Rhinovirus 3 (HRV3)	Capsid shell (VP1, VP2, VP3)	ICAM-1 (μ M range)	Unknown	Unknown	Minor-group human rhinoviruses use VLDL-R as a receptor; there are structural similarities between ICAM-1 binding to capsid shell and CD4 binding to gp120.
Adenovirus 2	Fibre, penton base	CAR (nM)	α v integrins	Sialic acid and heparin sulphate proteoglycans	Adenovirus fibre attaches to the CAR and integrins interact with the penton base, leading to internalization.
Reovirus 1	σ 1	JAM-1 (nM)	Unknown	Sialic acid	There are structural similarities between adenovirus fibre and σ 1, and between CAR and JAM-1. All serotypes bind JAM-1.

Dimitrov, D. Virus entry: molecular mechanisms and biomedical applications. *Nat Rev Microbiol* **2**, 109–122 (2004).

Hoffmann, M. *et al.* SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* **181**, 271–280 (2020).

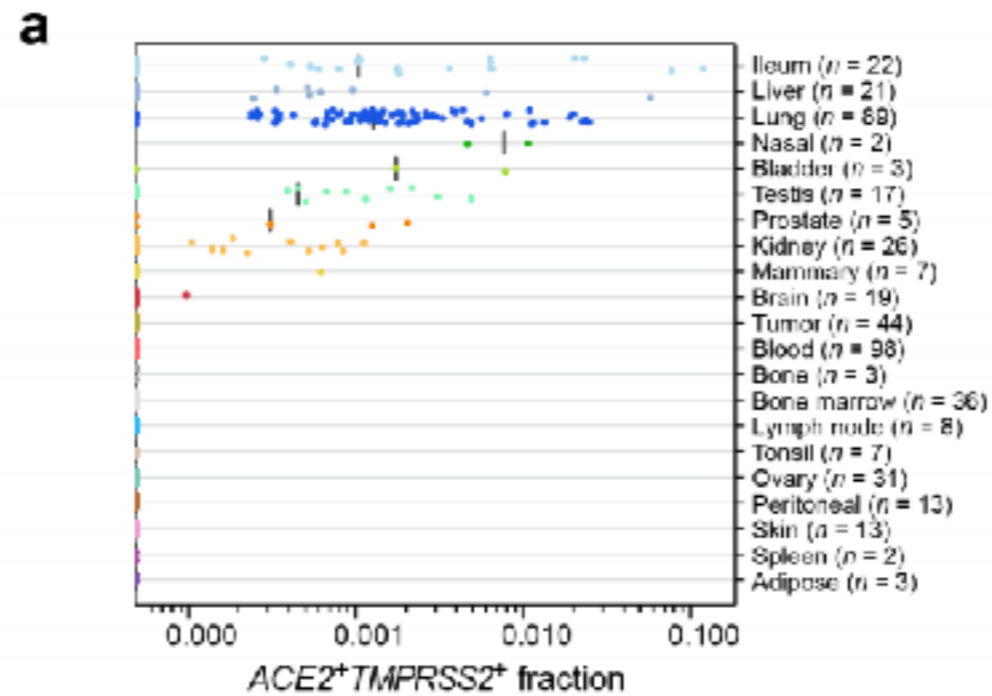
Binding: Viral Proteins & Cellular Receptors

*the choice of the cell surface receptor determines
which **cell types** and which **species** the virus can infect*



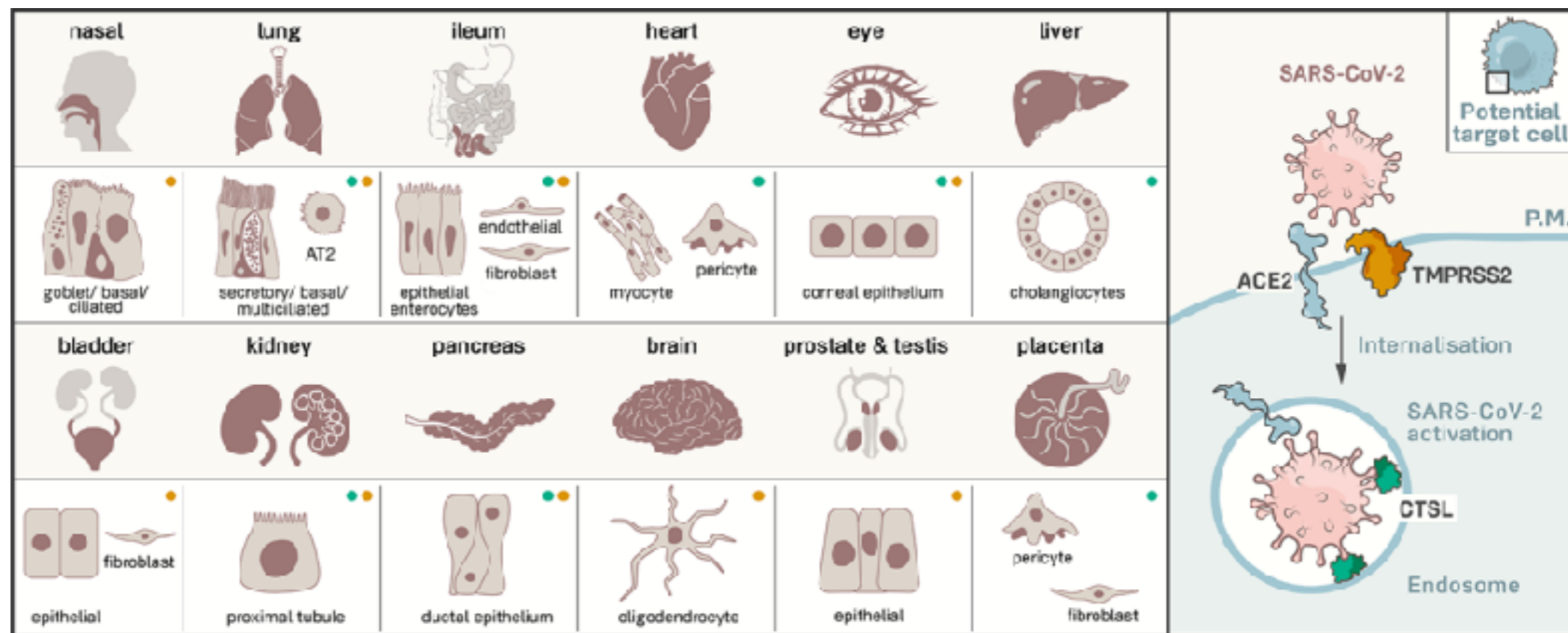
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Binding: Viral Proteins & Cellular Receptors

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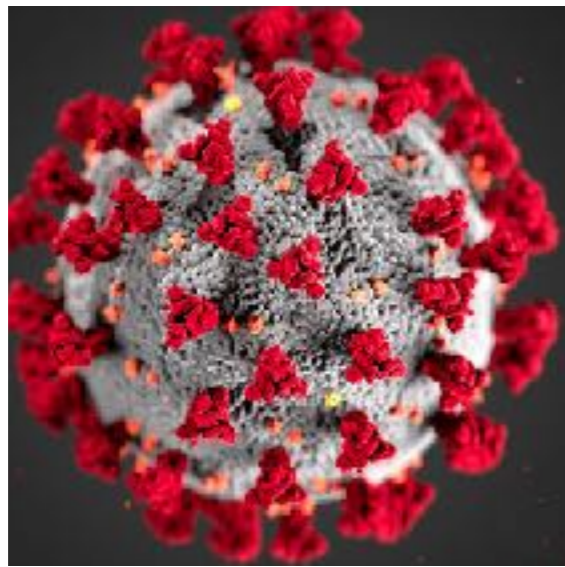


Muus, C. et al. *BioRxiv* April 21, 2020. DOI: 10.1101/2020.04.19.049254.

Figure from *The Scientist*. Receptors for SARS-CoV-2 Present in Wide Variety of Human Cells.

What Binds to Receptors?

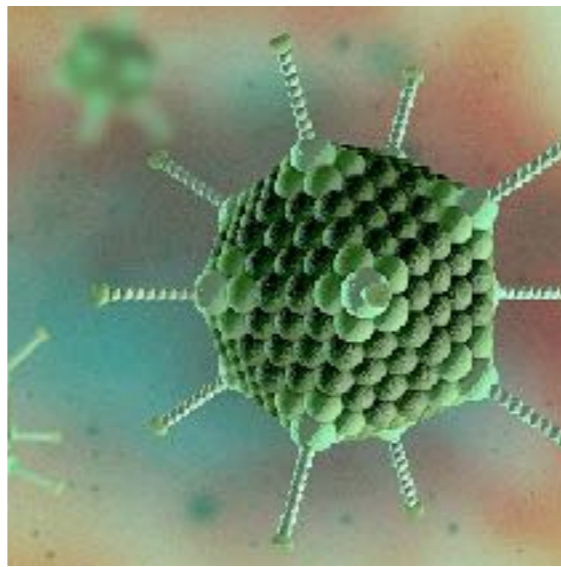
Enveloped



glycoproteins

coronavirus

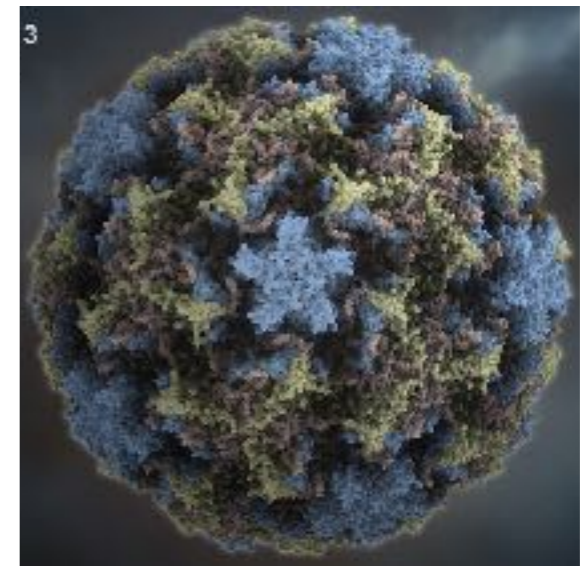
Non-Enveloped



fibers

adenovirus

Non-Enveloped

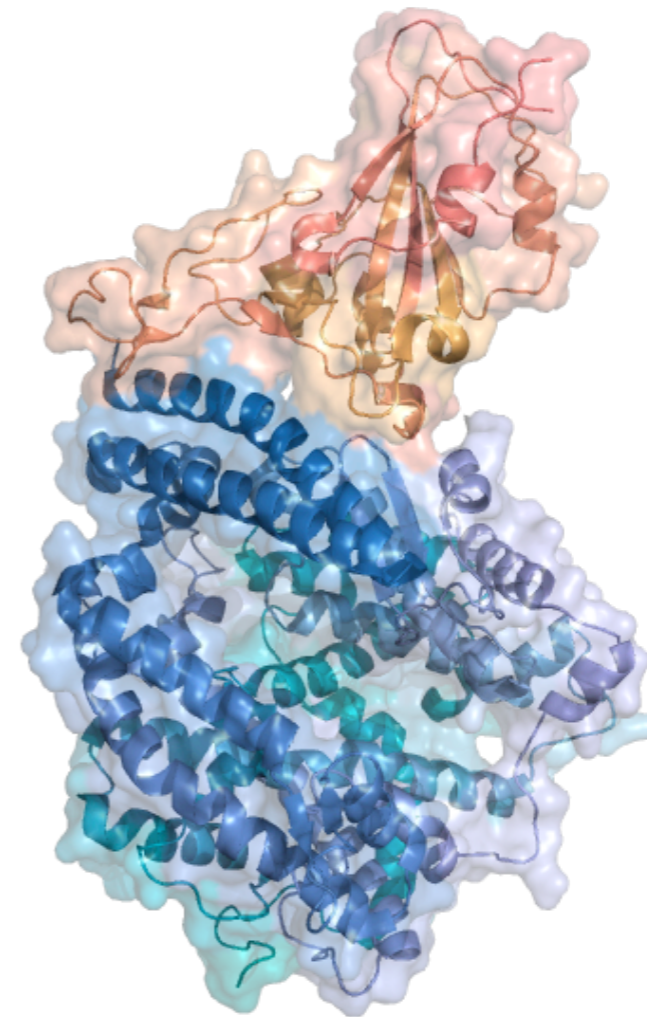


*surface protrusions
or indentations*

rhinovirus

Outline

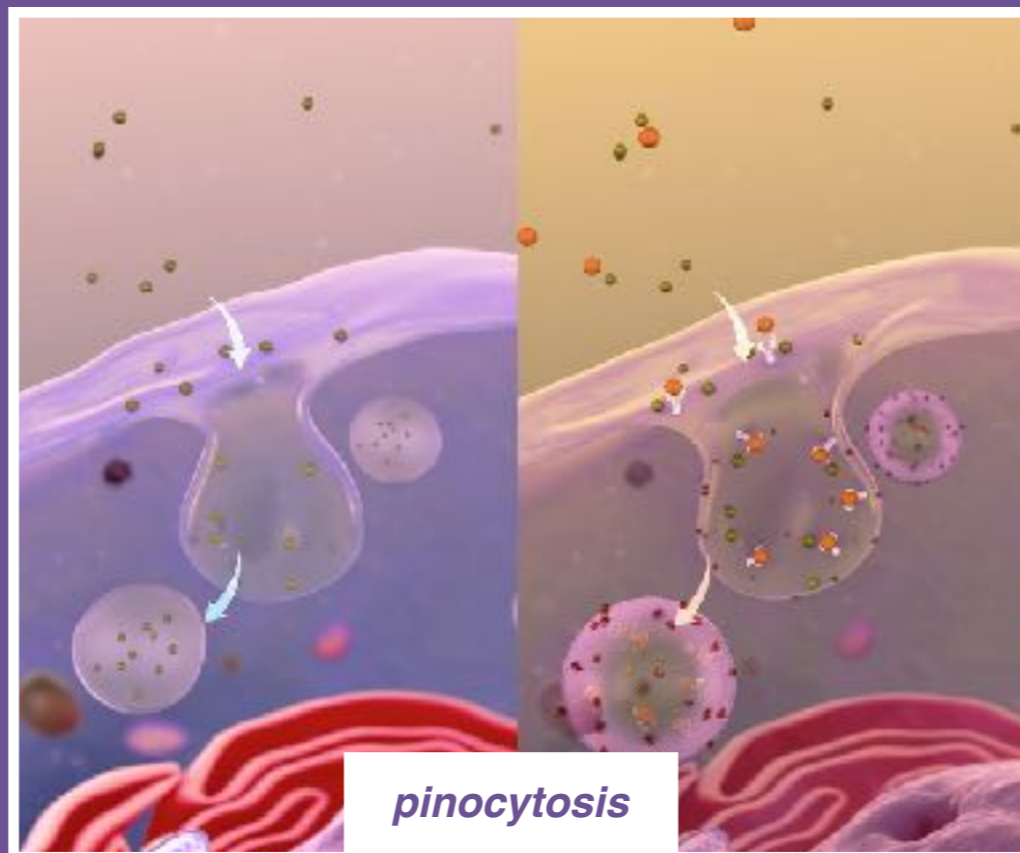
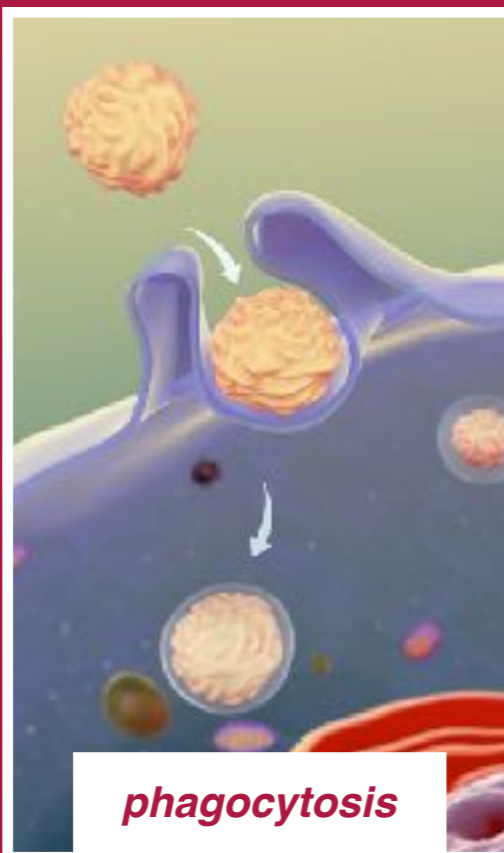
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Endocytosis

endocytosis is the uptake of fluids, solutes, membrane, and particles by invagination of the plasma membrane and formation of cytoplasmic vesicles

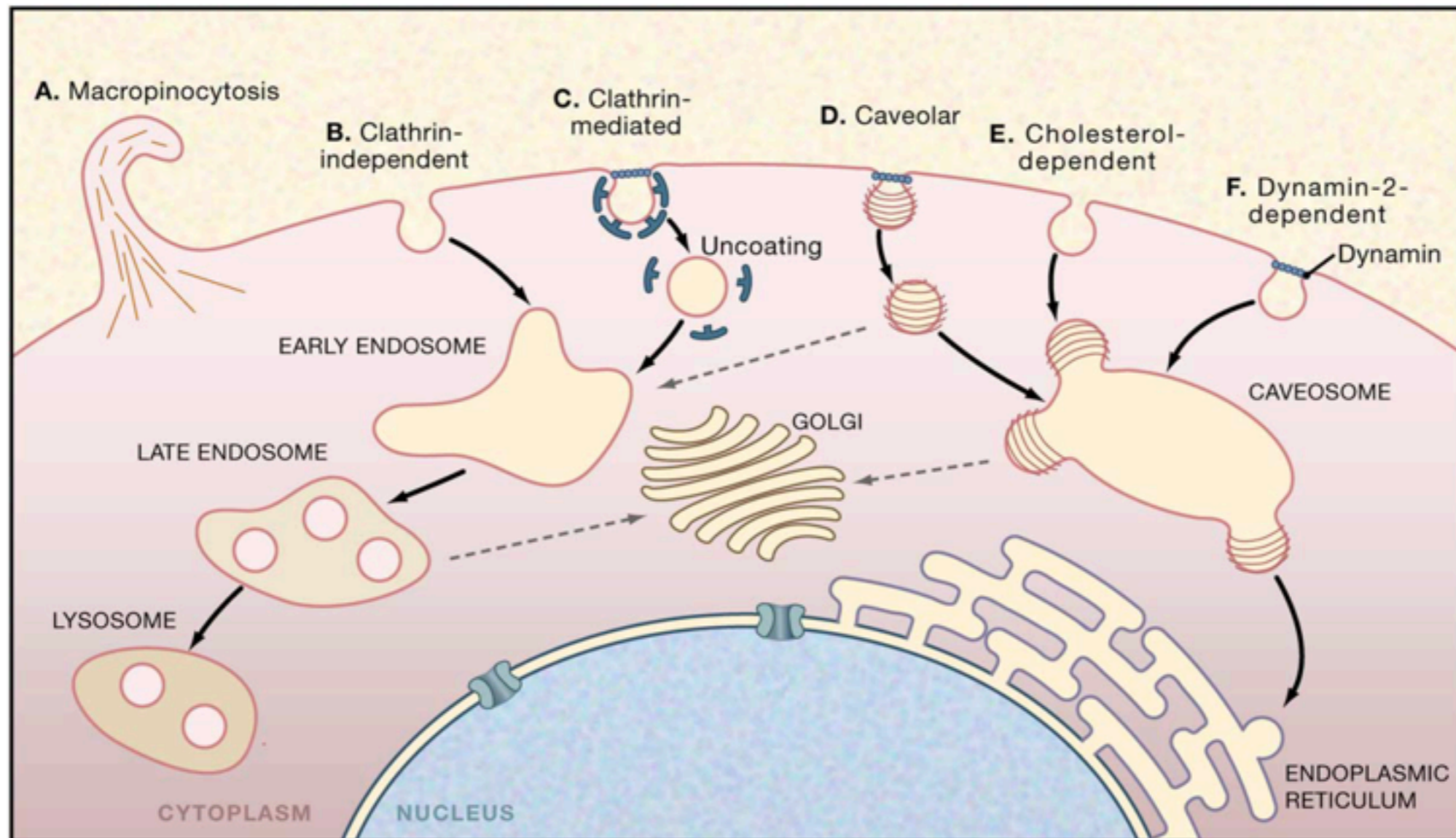
receptor-mediated endocytosis: uptake depends on its binding to a receptor



Endocytosis

dynamin vs dynamin-independent processes

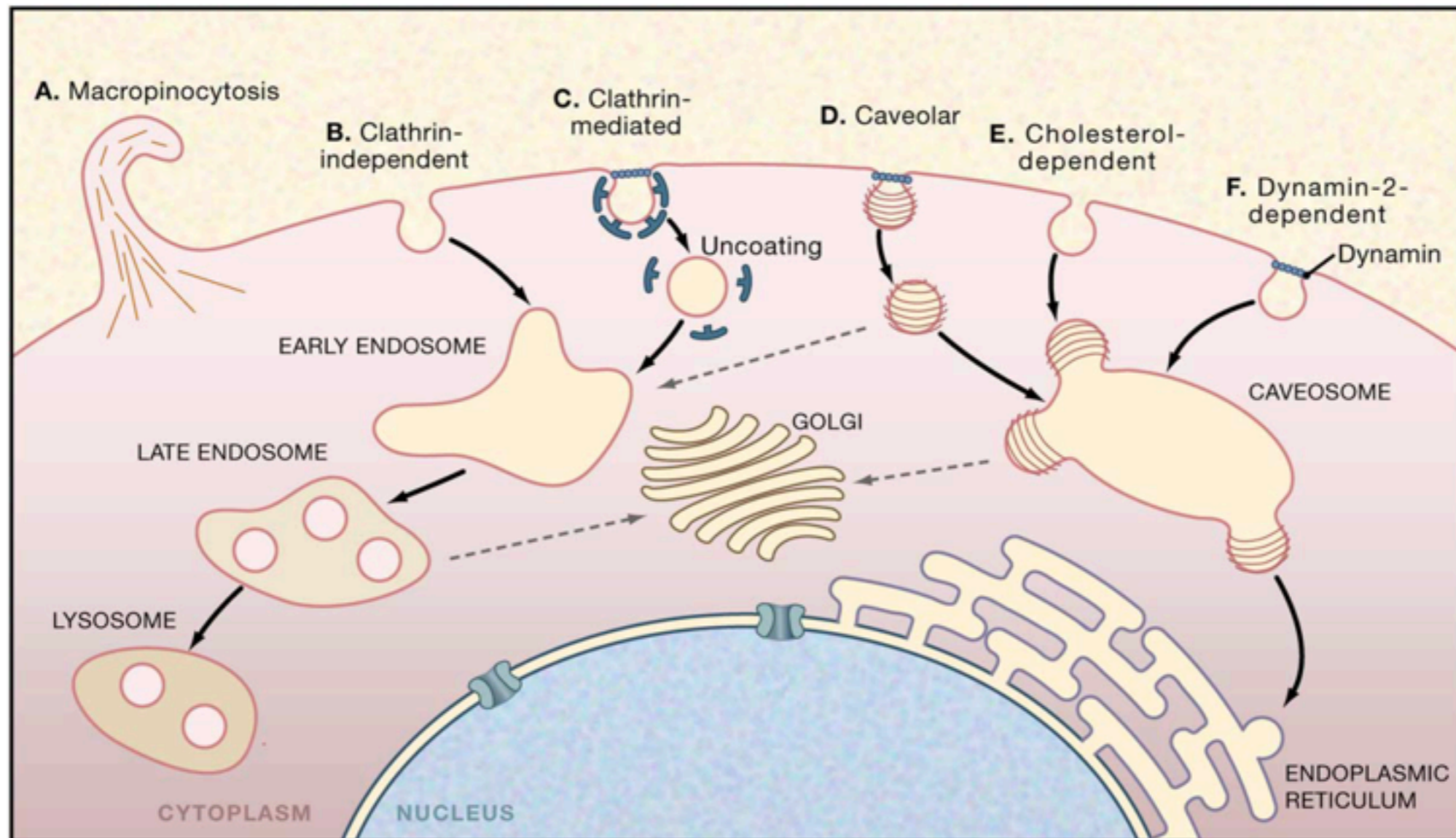
dynamin is a GTPase protein responsible in the scission of newly formed vesicles



Endocytosis

pH changes

◆ surface (7.0) ◆ early endosome (6.0–6.5) ◆ late endosome (5.0–5.5) ◆ lysosome (4.5–5)



Endocytosis

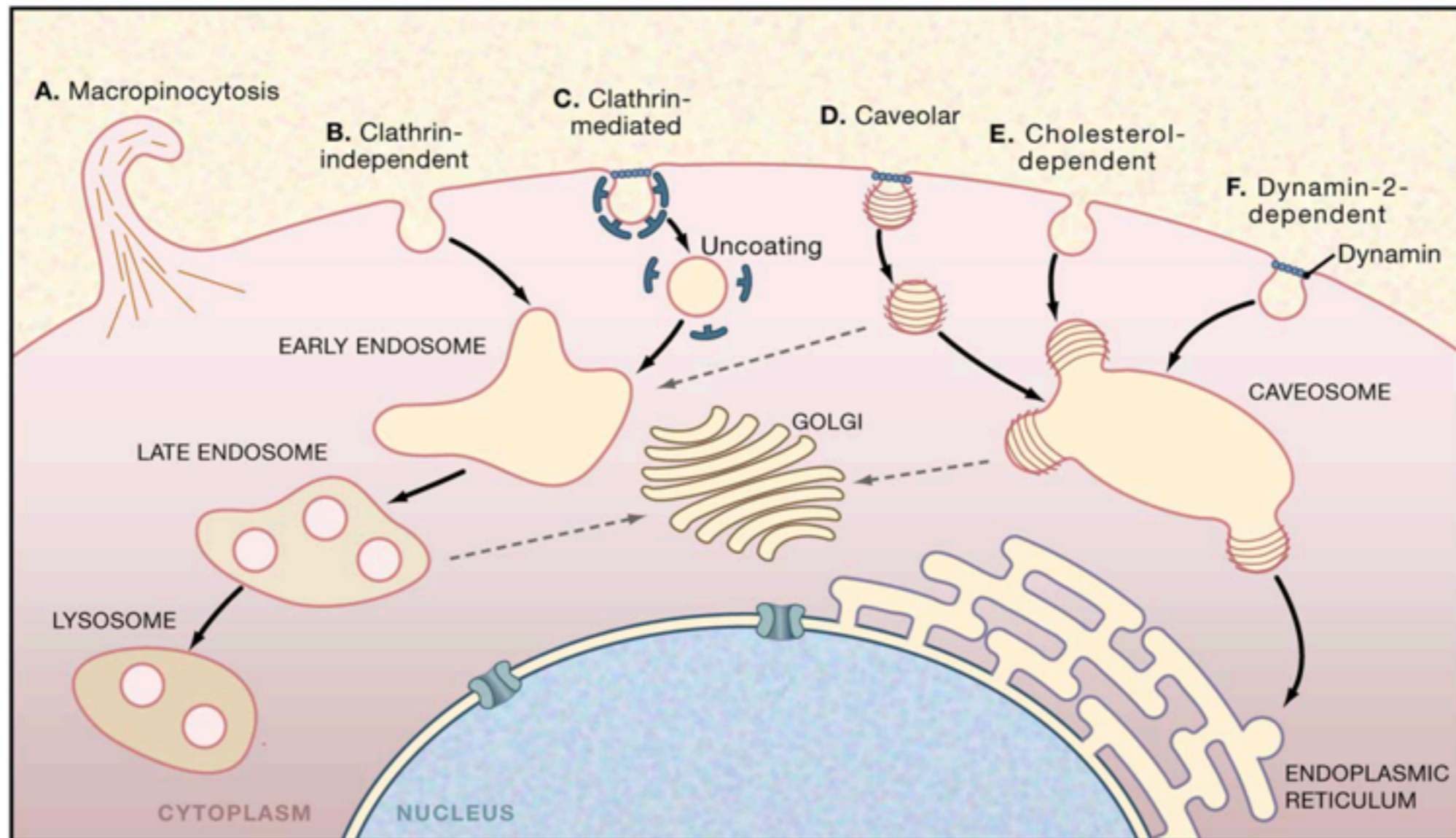
vaccinia
adeno B
HSV1

influenza
HPV16
LCMV

SFV
VSV
influenza
flavi
adeno 2/5

SV40
BK

*viruses can have multiple
endocytosis mechanisms*

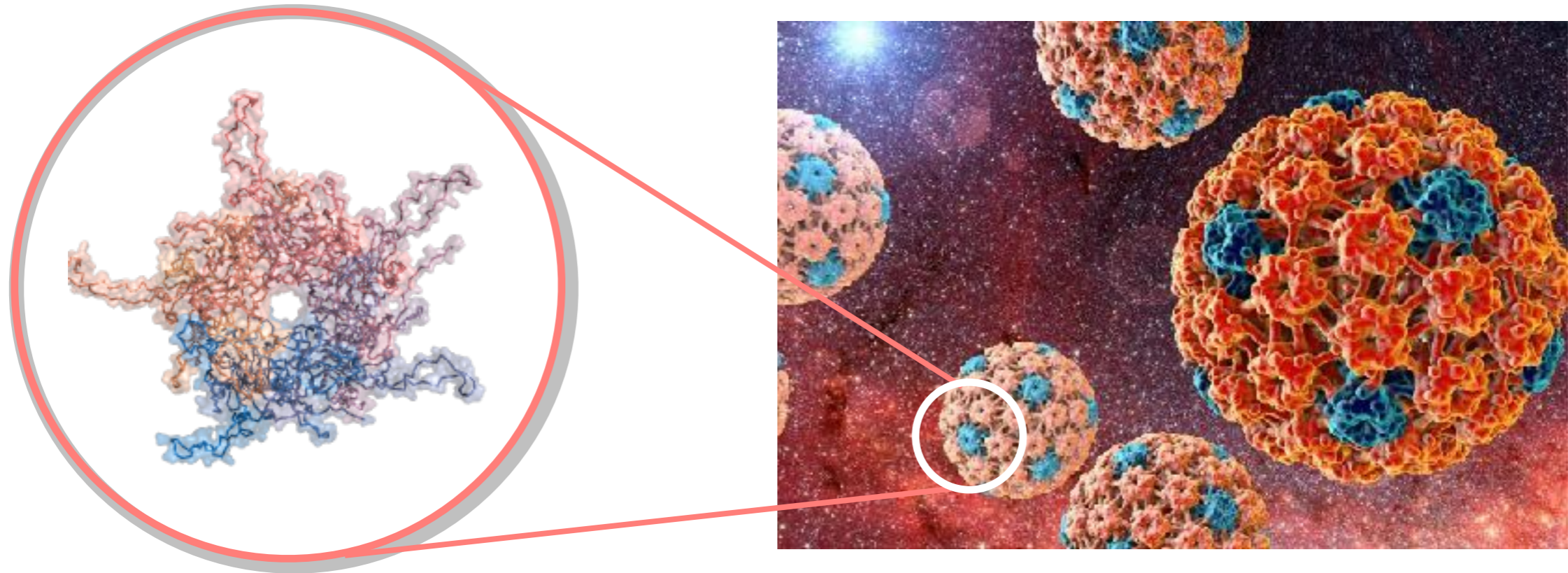


Endocytosis

endocytosis mechanism defines what cellular machinery is being used

- ◆ *clathrin, adapters, dynamin, caveolin*
- ◆ *cytoskeleton (actin, microtubule)*
- ◆ *signaling molecules (kinases)*
- ◆ *regulatory factors (Rho GTPases, Rabs, Arfs)*
- ◆ *ion channels (Na⁺/H⁺ exchangers, Ca⁺² channels)*
- ◆ *acidification machinery (vATPases, CLICs)*
- ◆ *lipids (cholesterol, phosphatidylinositides)*

Developing a Picture of Viral Entry: Human Papillomavirus 16



endocytosis to late endosome
noncoated pit (visually seen by microscopy)

$t_{1/2}$ endocytosis = 3 hours
 $t_{1/2}$ acid-activation = 10 hours

Developing a Picture of Viral Entry: Human Papillomavirus 16

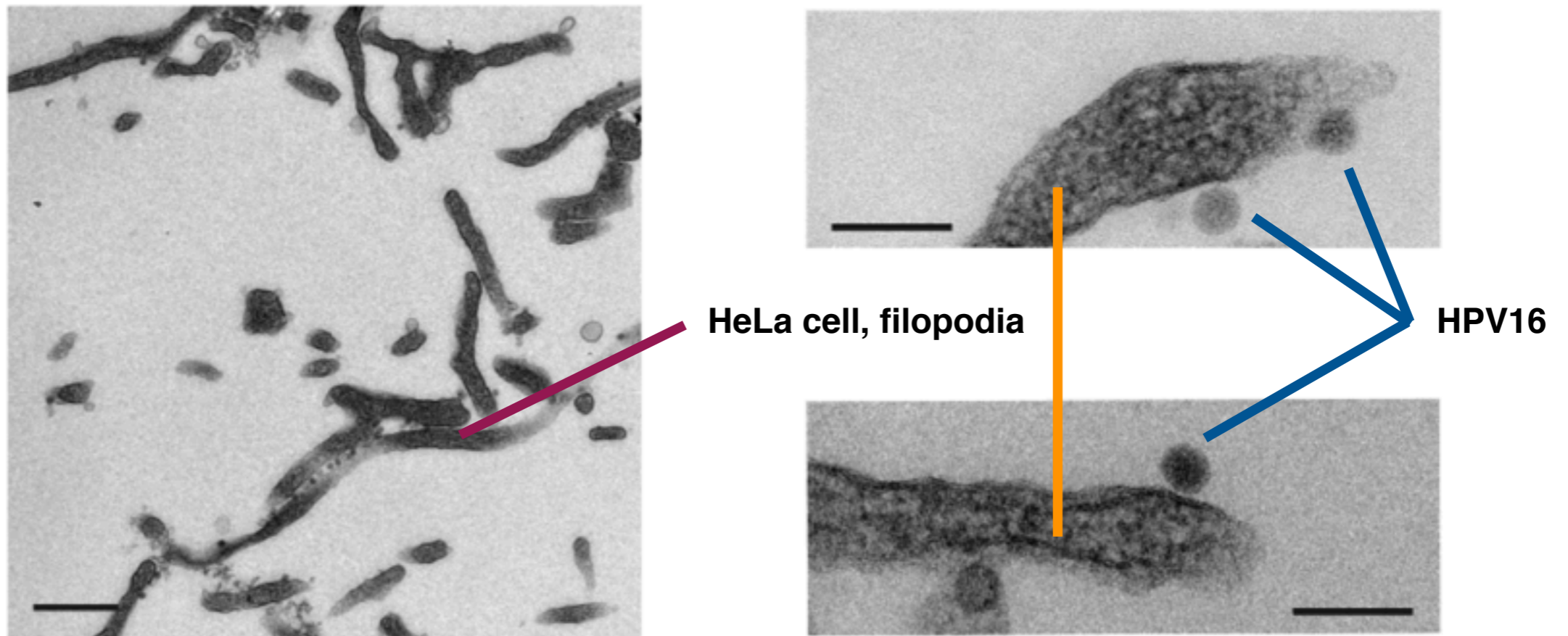
not required

- ◆ *clathrin*
- ◆ *AP2*
- ◆ *dynamin-2*
- ◆ *Arf 6*
- ◆ *cholesterols or lipid rafts*
- ◆ *caveolin, flotillin*
- ◆ *cdc42 or rac1*

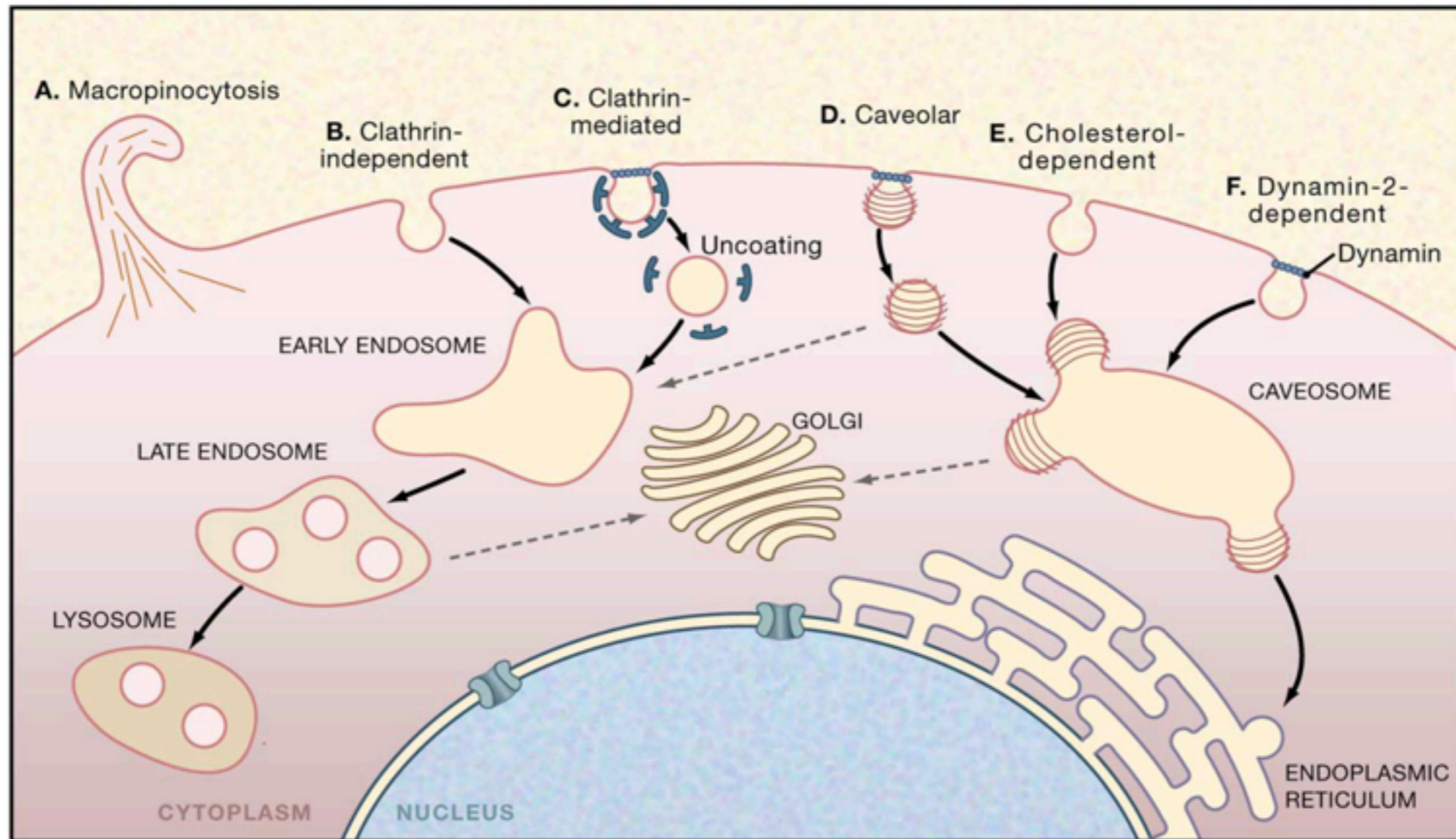
required

- ◆ *acidification*
- ◆ *protein kinase C zeta*
- ◆ *Tyr-kinases & phosphatases*
- ◆ *PI(3)kinase* ◆ *N-WASP*
- ◆ *Na⁺/H⁺ exchangers*
- ◆ *actin & microtubules*
- ◆ *Arf 1* ◆ *Rab5, Rab34*

Developing a Picture of Viral Entry: Human Papillomavirus 16

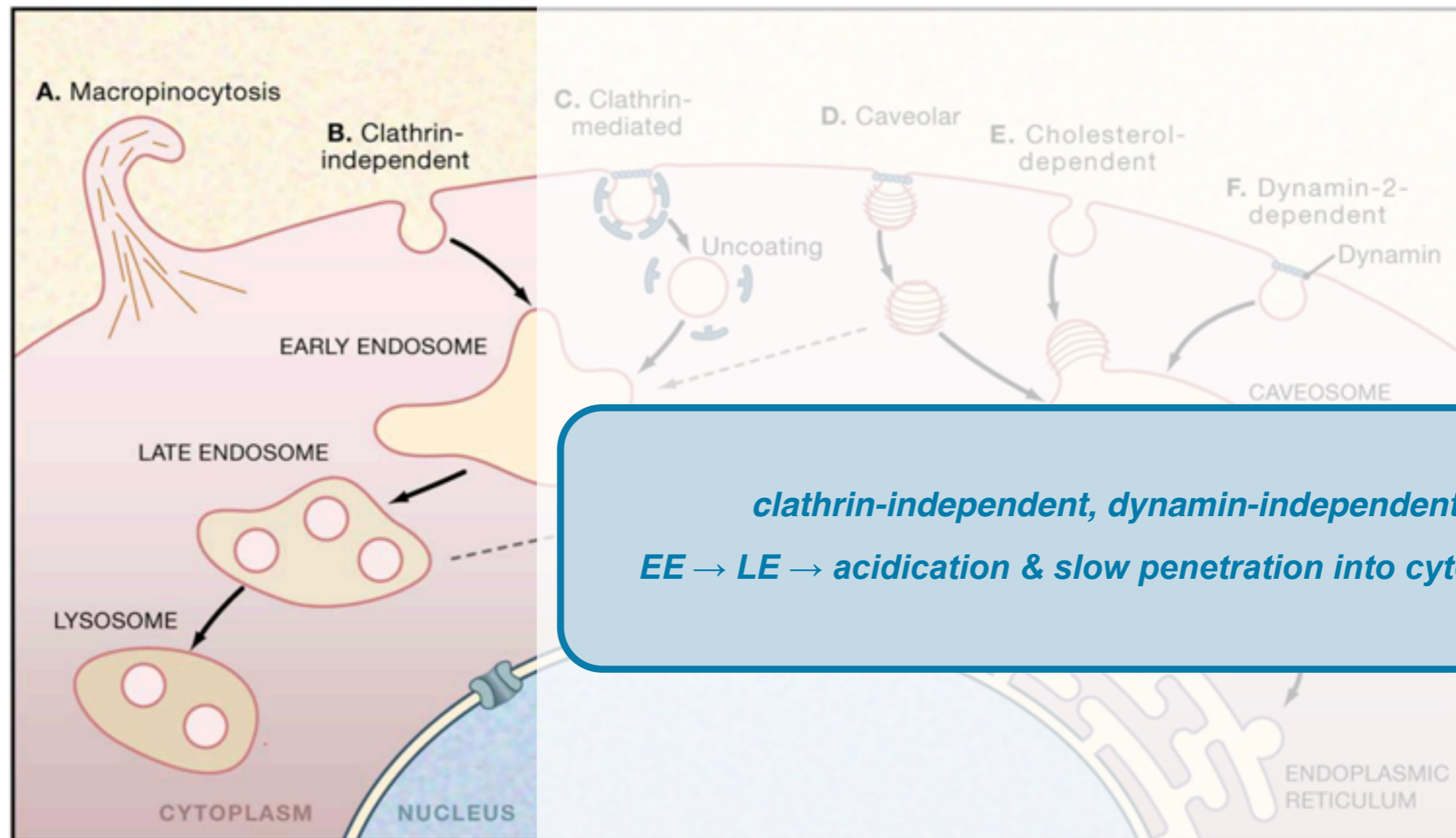


Developing a Picture of Viral Entry: Human Papillomavirus 16



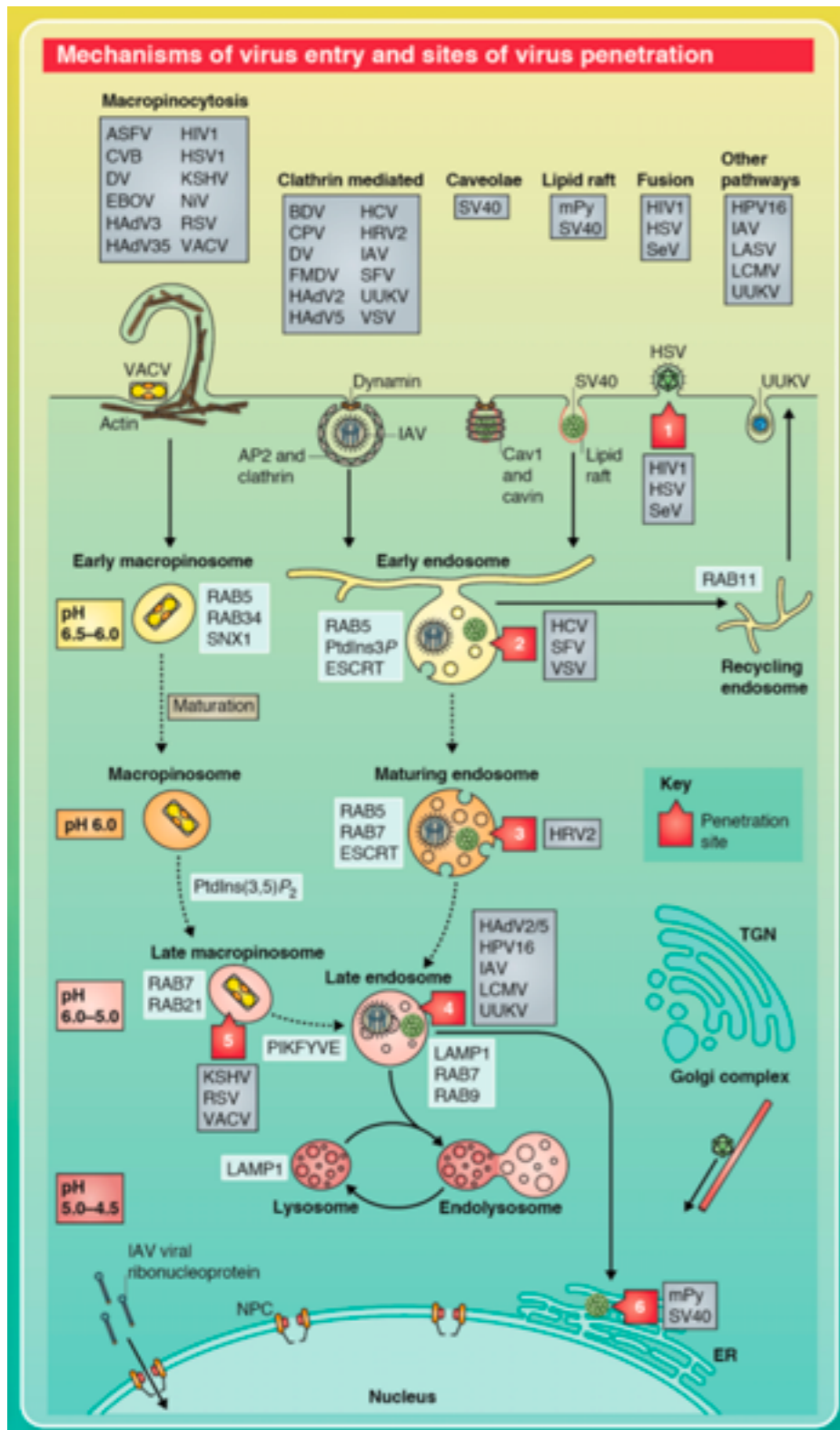
Schelhaas, M.; Ewers, H.; Rajamäki, M.-L.; Day, P. M.; Schiller, J. T.; Helenius, A. (2008) Human Papillomavirus Type 16 Entry: Retrograde Cell Surface Transport along Actin-Rich Protrusions. PLoS Pathog 4(9): e1000148. DOI: 10.1371/journal.ppat.1000148

Developing a Picture of Viral Entry: Human Papillomavirus 16



clathrin-independent, dynamin-independent
EE → LE → acidification & slow penetration into cytoplasm

Cell Penetration



penetration: the event that allows the viral genome to move into the cytosol

during viral entry, virus is mostly passive mostly responding to cellular cues & machinery

penetration requires the viral participation

penetration can occur anywhere along the endocytic route cell membrane, EE, LE, Lysosome, ER

membrane fusion (enveloped virus)

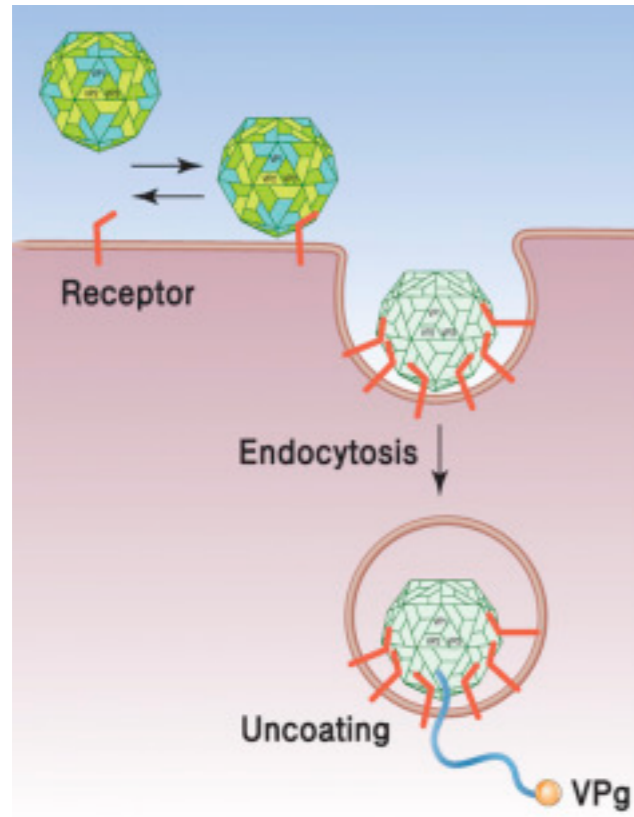
vacuole lysis

pore formation

ER associated degradation pathways

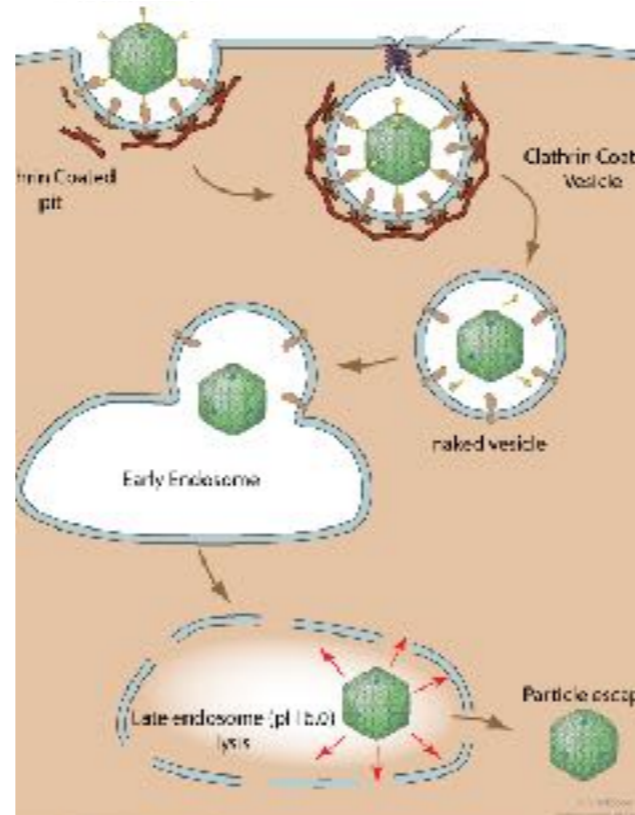
Examples of Viral Penetration

poliovirus



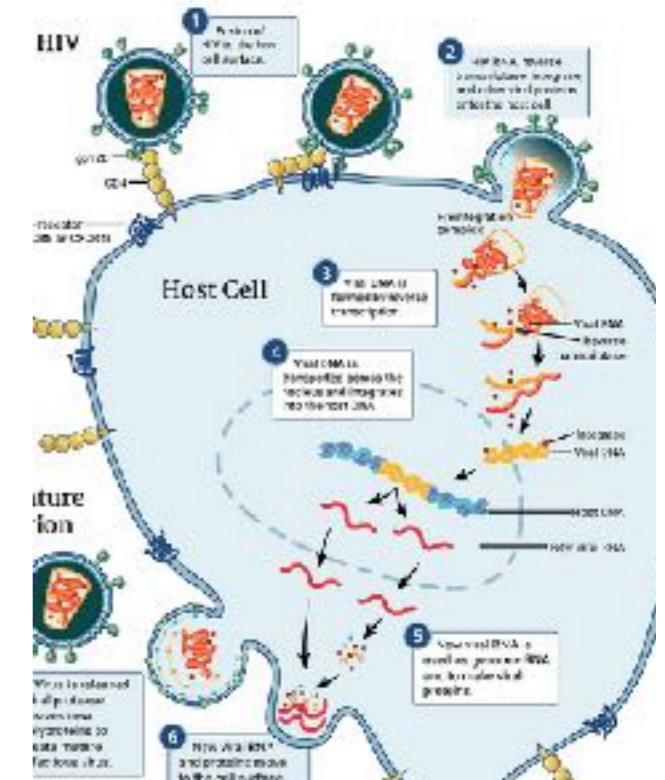
pores

adenovirus



vesicle lysis

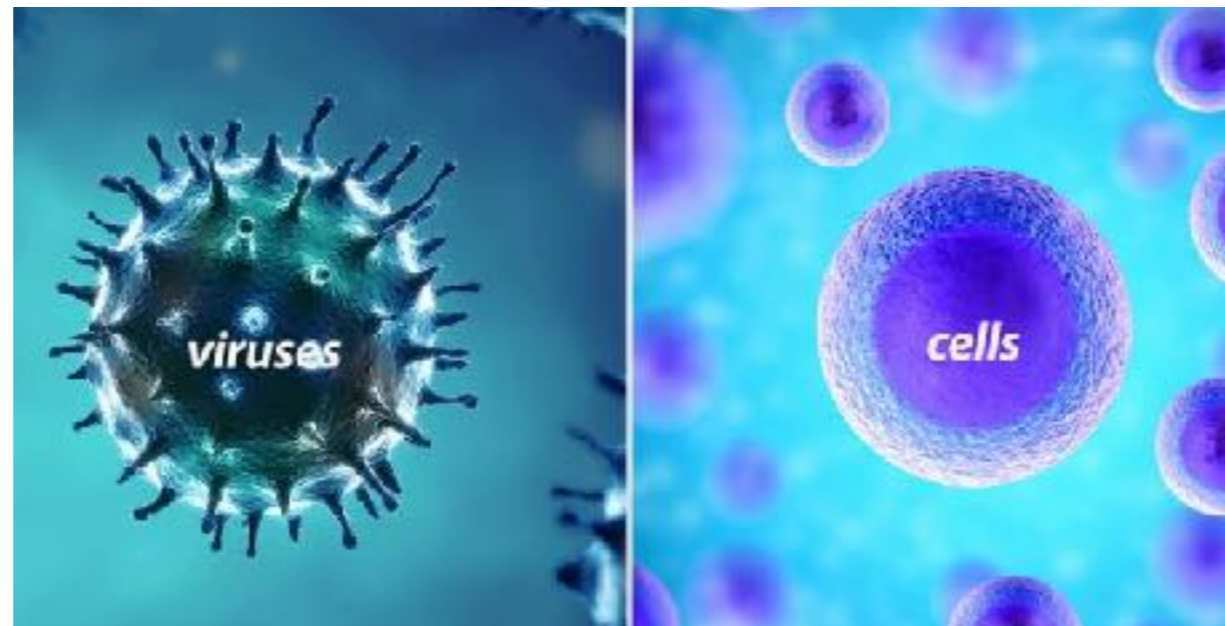
HIV-1



membrane fusion

A Thought

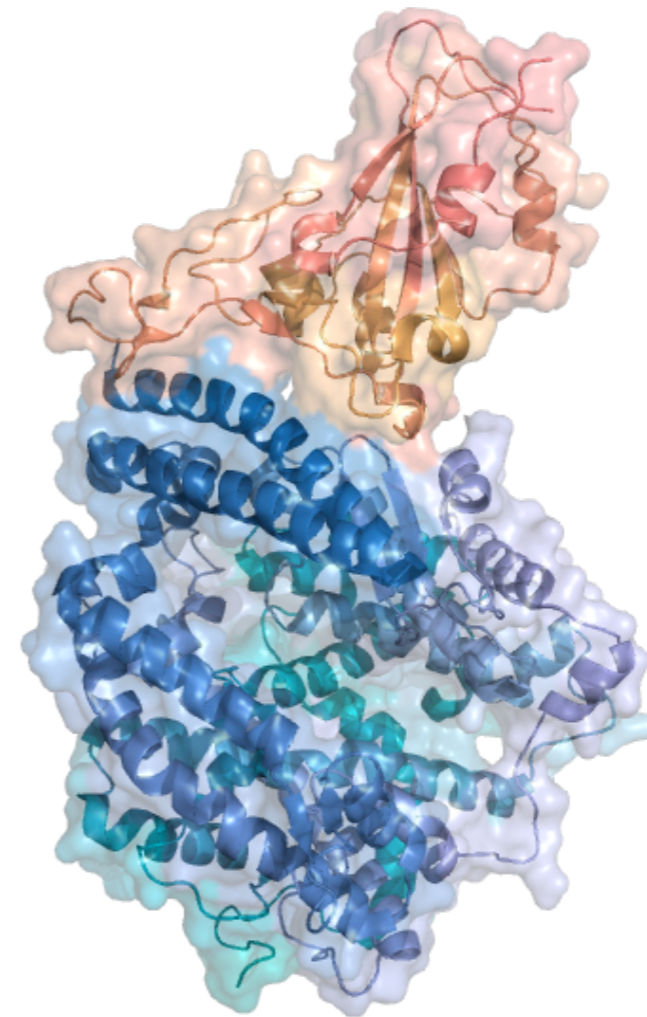
to understand a virus one must understand the cell that it infects



one can understand and learn about cells by studying viruses

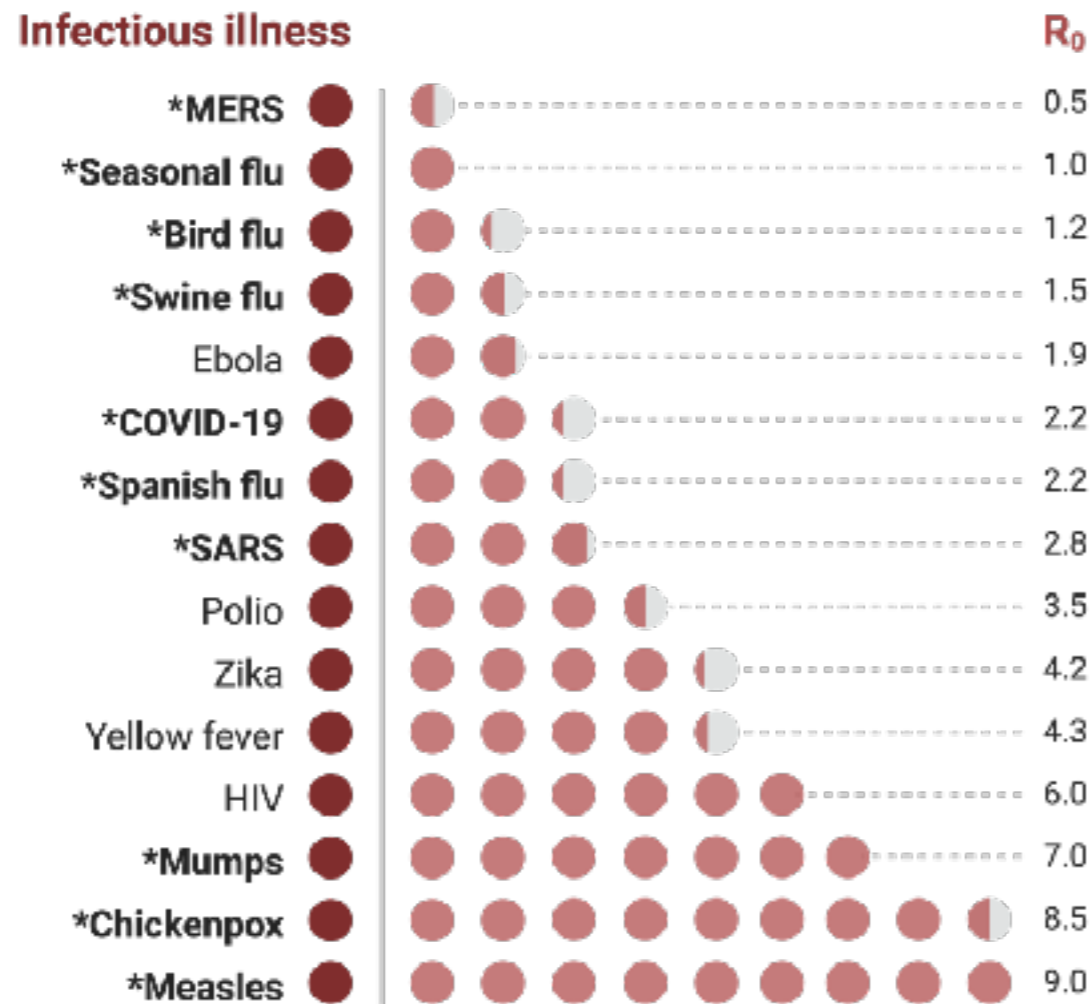
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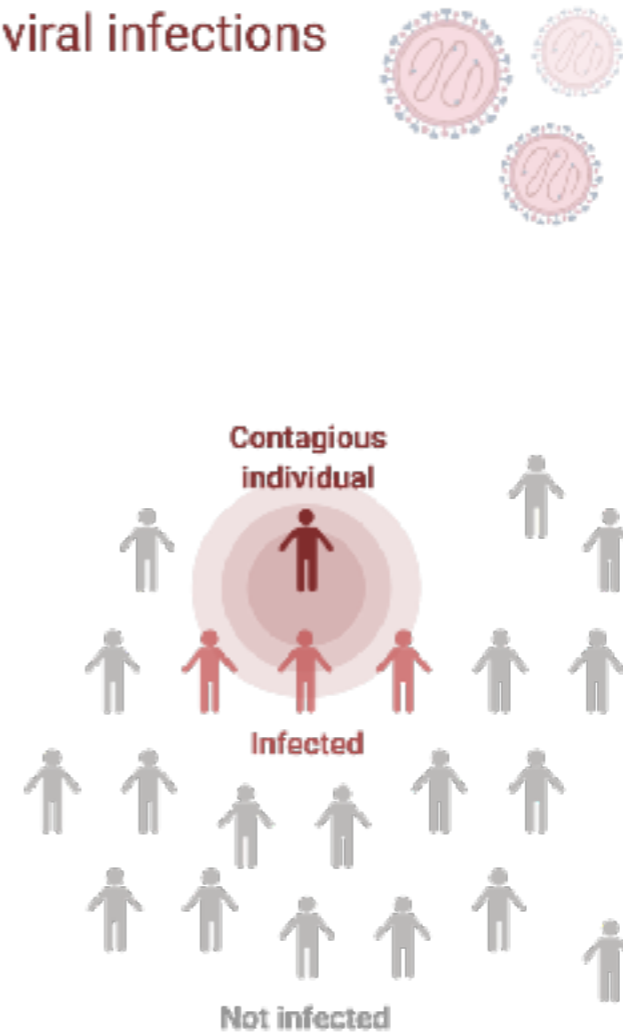


SARS-CoV-2: A Quick Overview

Average Basic Reproduction Number (R_0) of common viral infections



*Mode of transmission:
air droplets 



The **average basic reproduction number (R_0)** is an epidemiologic metric that describes the transmissibility of infectious agents. R_0 measures the expected number of secondary infections produced by a single infectious individual in a susceptible population during the mean infectious period.

SARS-CoV-2: A Quick Overview

AIDS *(40 million HIV-1 infected, 25 million deaths)*

Hepatitis B Virus *(240 million chronically infected worldwide (2.2 million US)
25% will succumb to liver disease or cancer)*


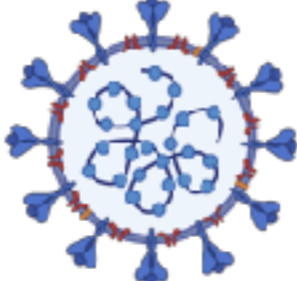
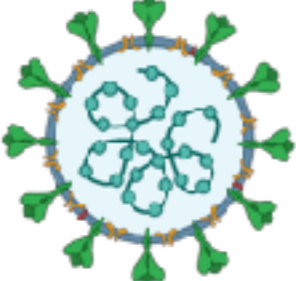
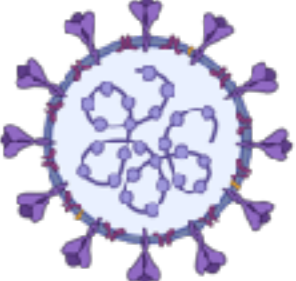
Rotavirus *(> 200,000 children deaths yearly)*

Influenza *(> 39 million deaths, 1918–19)*

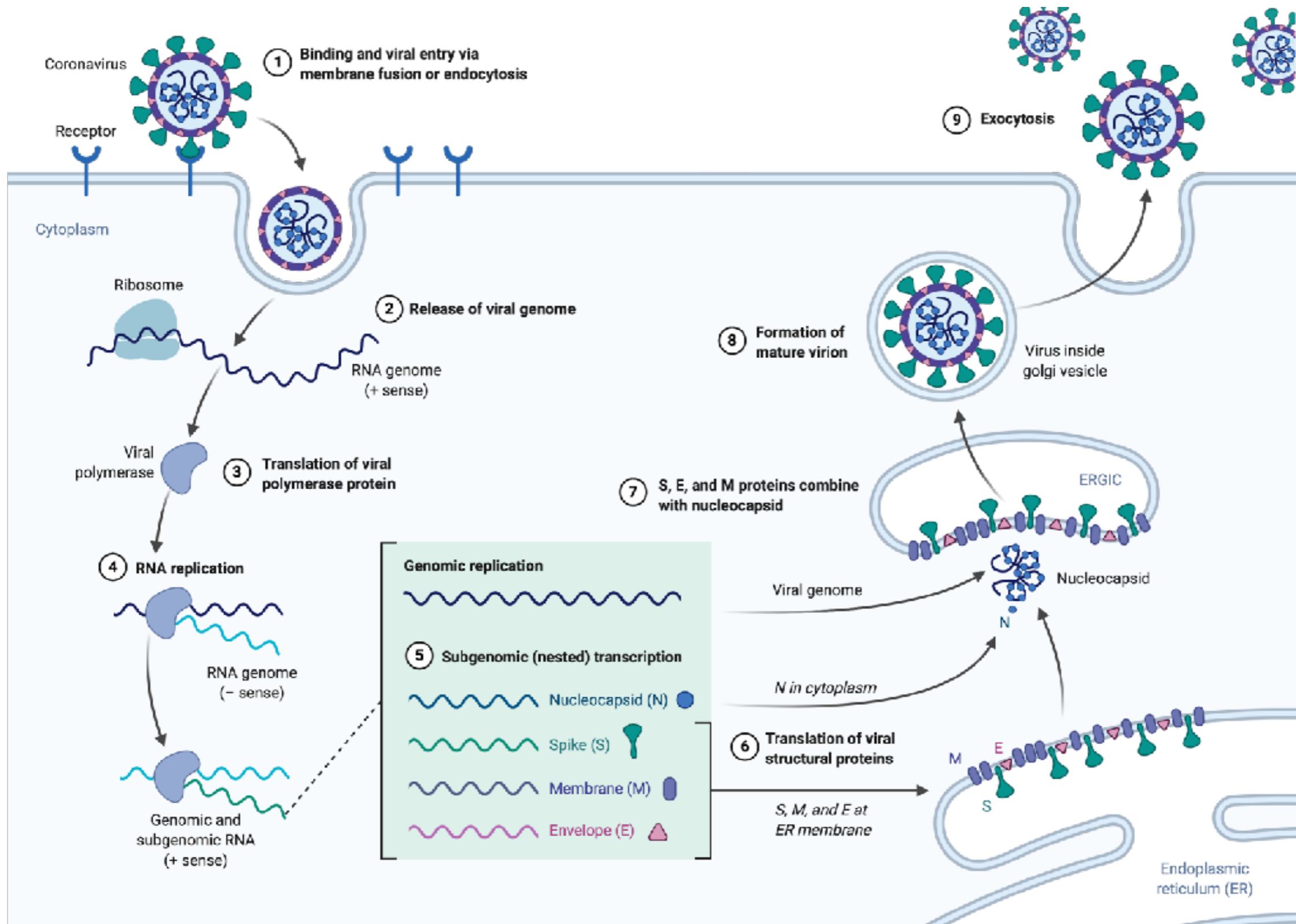
Avian Influenza (H5N1) *(pandemic threat in 2005)*

SARS-CoV-2 *(3.5 million infected, 250,000 deaths, declared pandemic in 2020)*

SARS-CoV-2: A Quick Overview

Disease	Flu	COVID-19	SARS	MERS
Disease Causing Pathogen	 Influenza virus	 SARS-CoV-2	 SARS-CoV	 MERS-CoV
R_0 Basic Reproductive Number	1.3	2.0 - 2.5 *	3	0.3 - 0.8
CFR Case Fatality Rate	0.05 - 0.1%	~3.4% *	9.6 - 11%	34.4%
Incubation Time	1 - 4 days	4 - 14 days *	2 - 7 days	6 days
Hospitalization Rate	2%	~19% *	Most cases	Most cases
Community Attack Rate	10 - 20%	30 - 40% *	10 - 50%	4 - 13%
Annual Infected (global)	~ 1 billion	3.5 million (5/20)	8098 (in 2003)	420
Annual Infected (US)	10 - 45 million	1.18 million (5/20)	8 (in 2003)	2 (in 2014)
Annual Deaths (US)	10,000 - 61,000	68,000 (5/20)	None	None

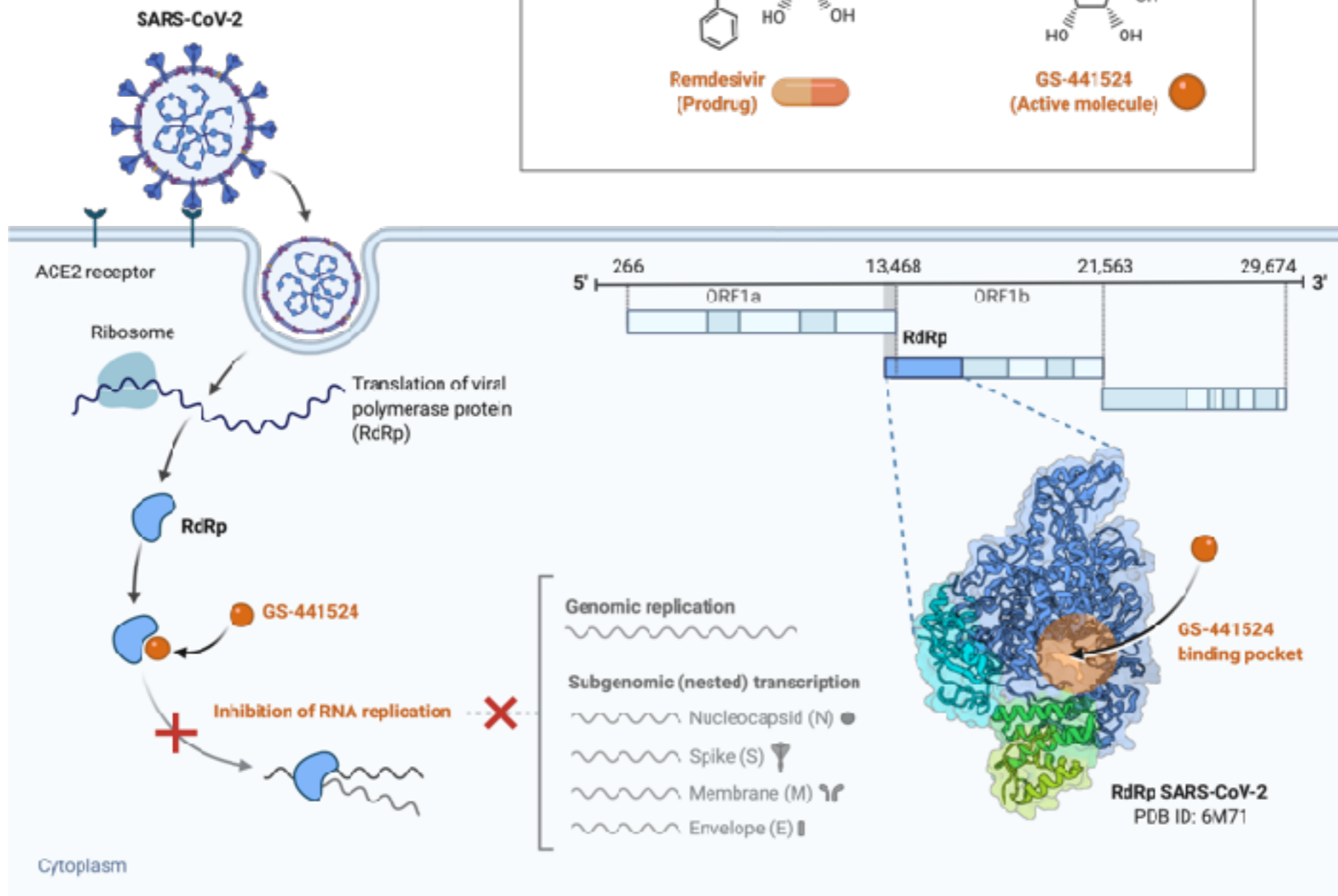
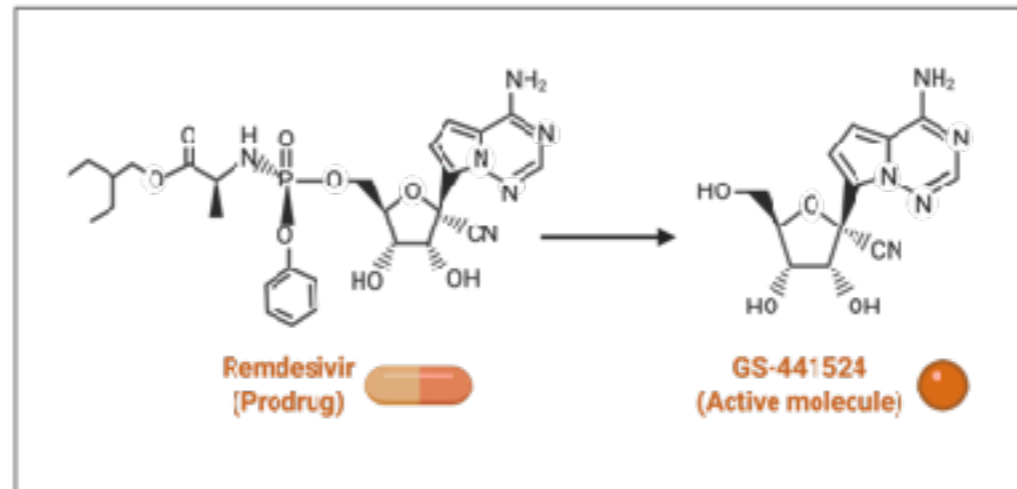
SARS-CoV-2: A Quick Overview



SARS-CoV-2: A Quick Overview

Remdesivir

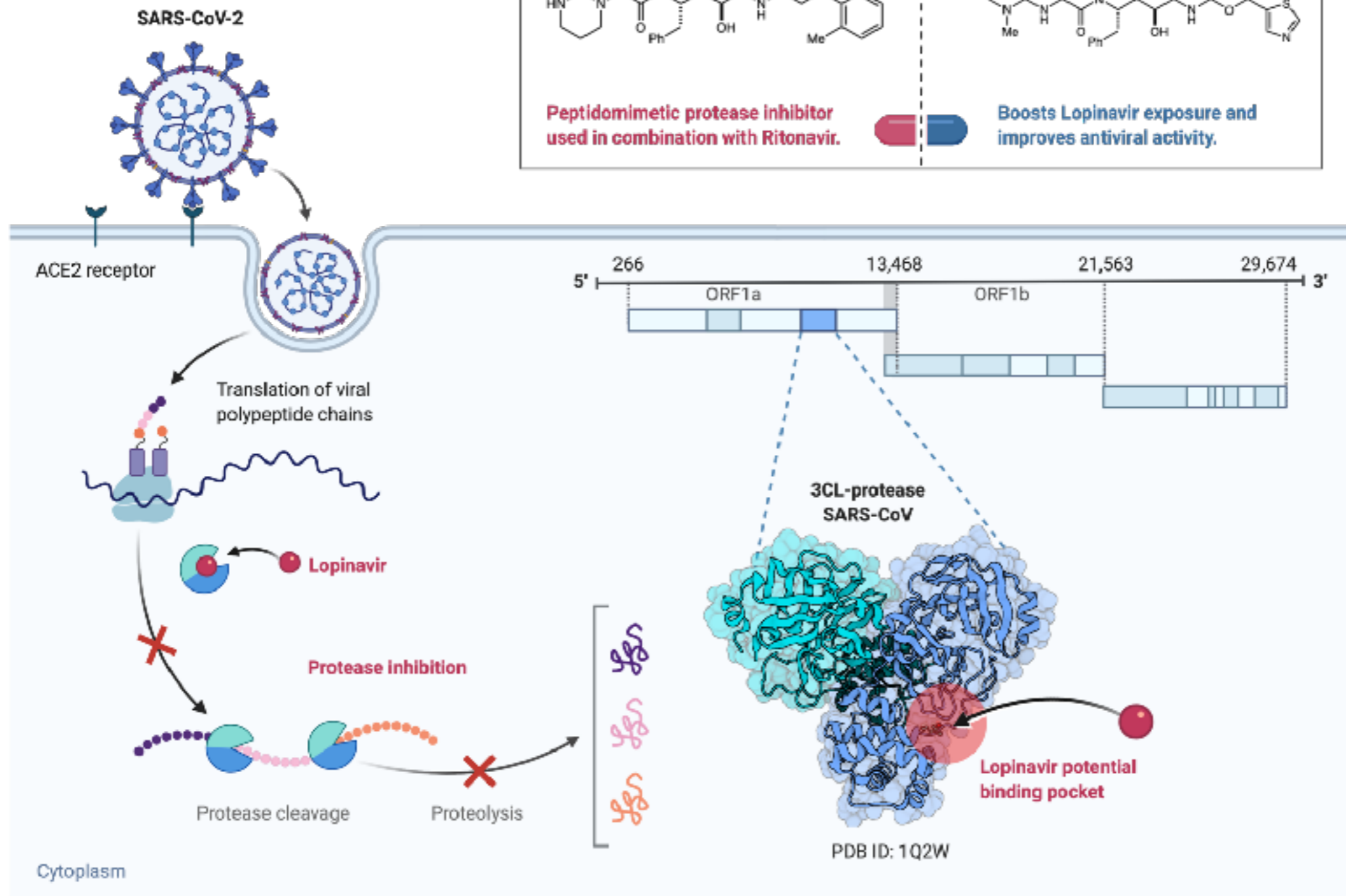
Potential repurposed drug candidate for COVID-19



SARS-CoV-2: A Quick Overview

Lopinavir

Potential repurposed drug candidate for COVID-19



SARS-CoV-2: A Quick Overview

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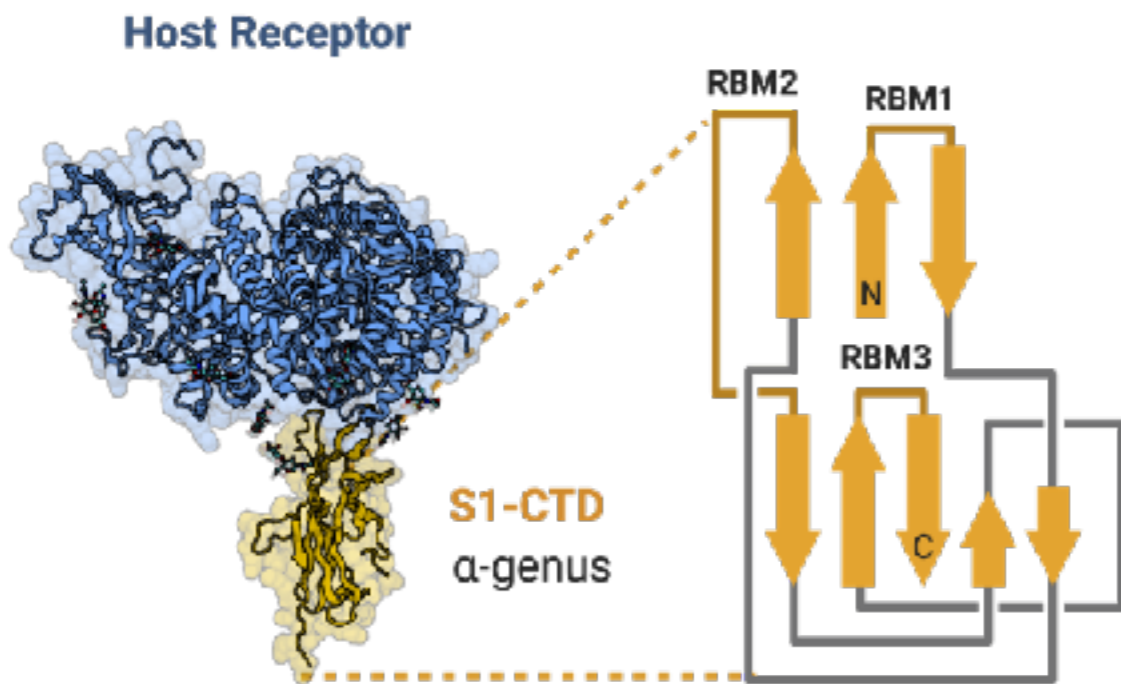
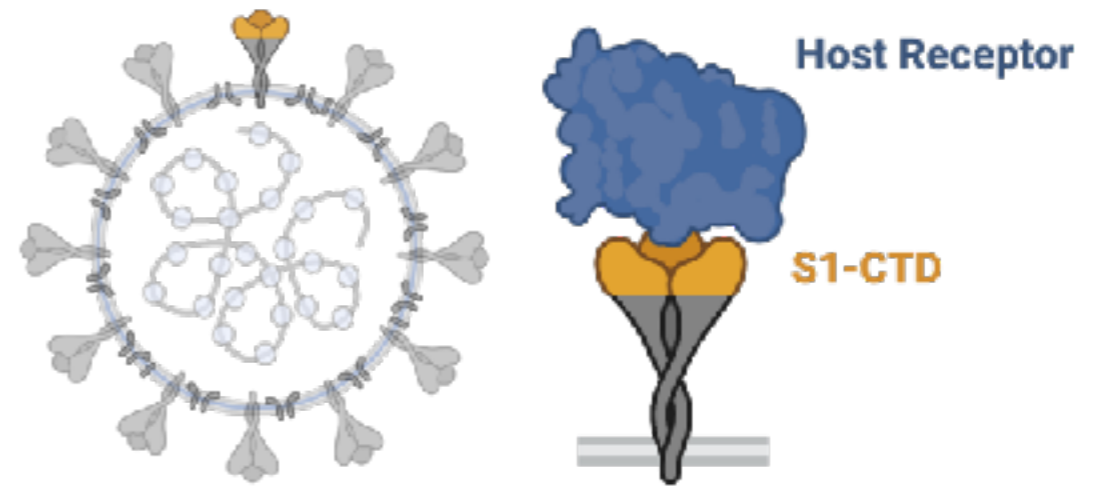
A SARS-CoV-2-Human Protein-Protein Interaction Map Reveals Drug Targets and Potential Drug-Repurposing

David E. Gordon^{1,2,3,4}, Gwendolyn M. Jang^{1,2,3,4}, Mehdi Bouhaddou^{1,2,3,4}, Jiewei Xu^{1,2,3,4}, Kirsten Obernier^{1,2,3,4}, Matthew J. O'Meara⁵, Jeffrey Z. Guo^{1,2,3,4}, Danielle L. Swaney^{1,2,3,4}, Tia A. Tummino^{1,2,6}, Ruth Huettenhain^{1,2,3,4}, Robyn M. Kaake^{1,2,3,4}, Alicia L. Richards^{1,2,3,4}, Beril Tutuncuoglu^{1,2,3,4}, Helene Foussard^{1,2,3,4}, Jyoti Batra^{1,2,3,4}, Kelsey Haas^{1,2,3,4}, Maya Modak^{1,2,3,4}, Minkyu Kim^{1,2,3,4}, Paige Haas^{1,2,3,4}, Benjamin J. Polacco^{1,2,3,4}, Hannes Braberg^{1,2,3,4}, Jacqueline M. Fabius^{1,2,3,4}, Manon Eckhardt^{1,2,3,4}, Margaret Soucheray^{1,2,3,4}, Melanie J. Bennett^{1,2,3,4}, Merve Cakir^{1,2,3,4}, Michael J. McGregor^{1,2,3,4}, Qiongyu Li^{1,2,3,4}, Zun Zar Chi Naing^{1,2,3,4}, Yuan Zhou^{1,2,3,4}, Shiming Peng^{1,2,6}, Ilsa T. Kirby^{1,4,7}, James E. Melnyk^{1,4,7}, John S. Chorba^{1,4,7}, Kevin Lou^{1,4,7}, Shizhong A. Dai^{1,4,7}, Wenqi Shen^{1,4,7}, Ying Shi^{1,4,7}, Ziyang Zhang^{1,4,7}, Inigo Barrio-Hernandez⁸, Danish Memon⁸, Claudia Hernandez-Armenta⁸, Christopher J.P. Mathy^{1,9,10,2}, Tina Perica^{1,2,9}, Kala B. Pilla^{1,2,9}, Sai J. Ganesan^{1,2,9}, Daniel J. Saltzberg^{1,2,9}, Rakesh Ramachandran^{1,2,9}, Xi Liu^{1,2,6}, Sara B. Rosenthal¹¹, Lorenzo Calviello¹², Srivats Venkataramanan¹², Jose Liboy-Lugo¹², Yizhu Lin¹², Stephanie A. Wankowicz^{1,13,9}, Markus Bohn⁶, Phillip P. Sharp⁴, Raphael Trenker¹⁴, Janet M. Young¹⁵, Devin A. Caverio³, Joseph Hiatt^{16,3}, Theodore L. Roth^{16,3}, Ujjwal Rathore³, Advait Subramanian^{1,17}, Julia Noack^{1,17}, Mathieu Hubert¹⁸, Ferdinand Roesch¹⁹, Thomas Vallet¹⁹, Björn Meyer¹⁹, Kris M. White²⁰, Lisa Miorin²⁰, Oren S. Rosenberg^{21,22,23}, Kliment A Verba^{1,2,6}, David Agard^{1,24}, Melanie Ott^{3,21}, Michael Emerman²⁵, Davide Ruggero^{26,27,4}, Adolfo García-Sastre²⁰, Natalia Jura^{1,14,4}, Mark von Zastrow^{1,1,4,28}, Jack Taunton^{1,2,4}, Alan Ashworth^{1,27}, Olivier Schwartz¹⁸, Marco Vignuzzi¹⁹, Christophe d'Enfert²⁹, Shaeri Mukherjee^{1,17}, Matt Jacobson⁶, Harmit S. Malik¹⁵, Danica G. Fujimori^{1,4,6}, Trey Ideker³⁰, Charles S. Craik^{6,27}, Stephen Floor^{12,27}, James S. Fraser^{1,2,9}, John Gross^{1,2,6}, Andrej Sali^{1,2,6,9}, Tanja Kortemme^{1,9,10,2}, Pedro Beltrao⁸, Kevan Shokat^{1,4,7}, Brian K. Shoichet^{1,2,6}, Nevan J. Krogan^{1,2,3,4}

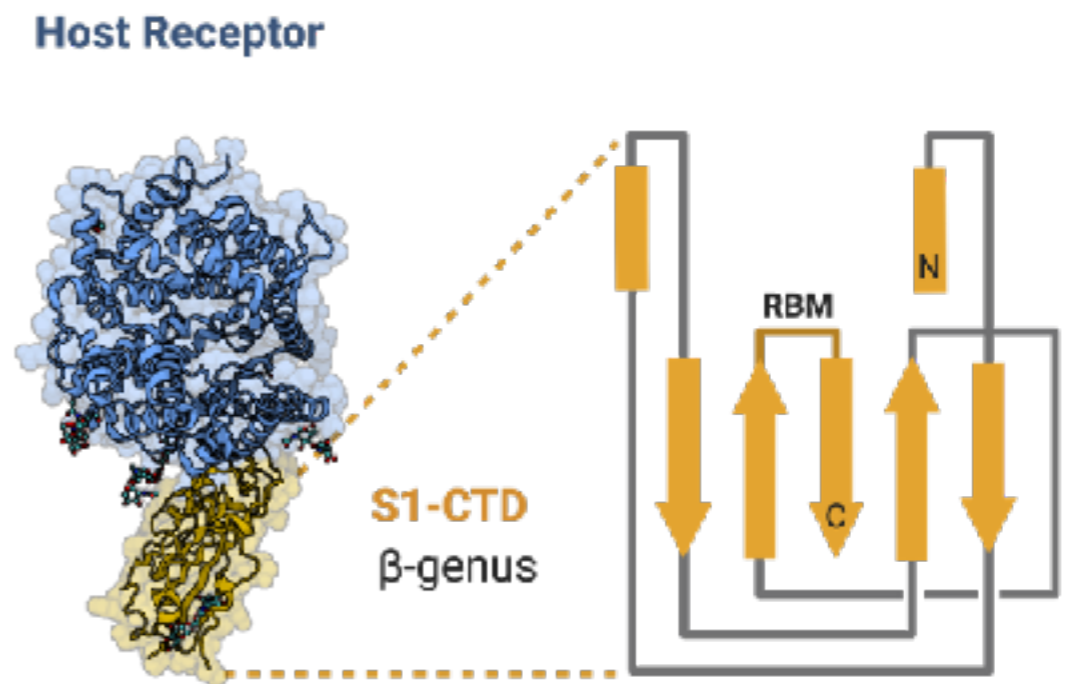
SARS-CoV-2: A Quick Overview

Receptor Binding Motifs (RBM)

SARS-CoV-2 spike protein (S1-CTD)



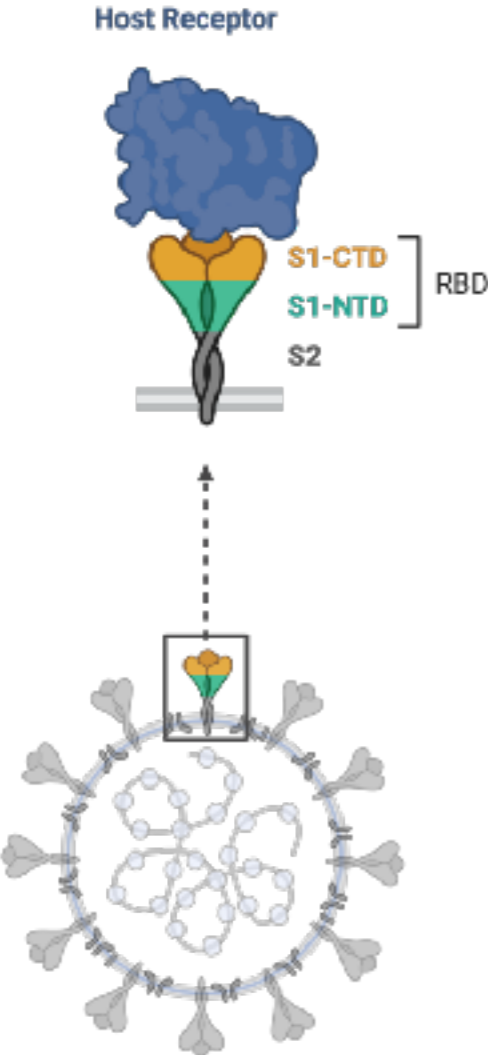
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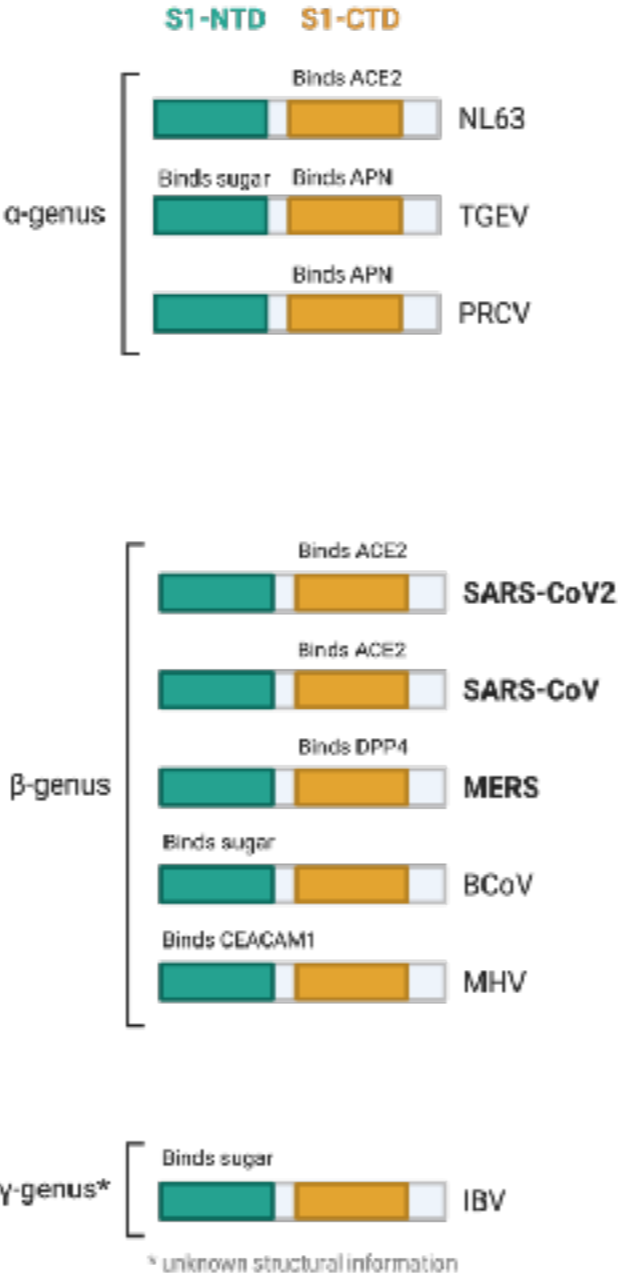
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SARS-CoV-2: A Quick Overview

Schematic of spike-receptor binding mechanism

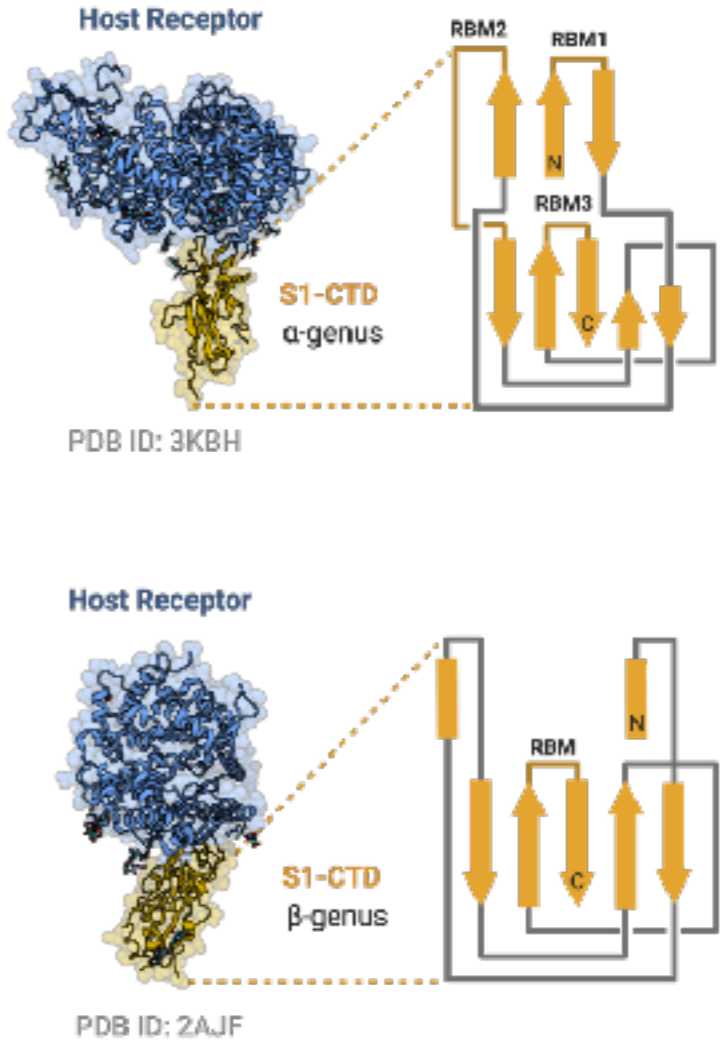


Receptor Binding Domain (RBD)



Receptor Binding Motifs (RBM)

Receptor binding motifs of S1-CTD bind to host receptor



SARS-CoV-2: A Quick Overview

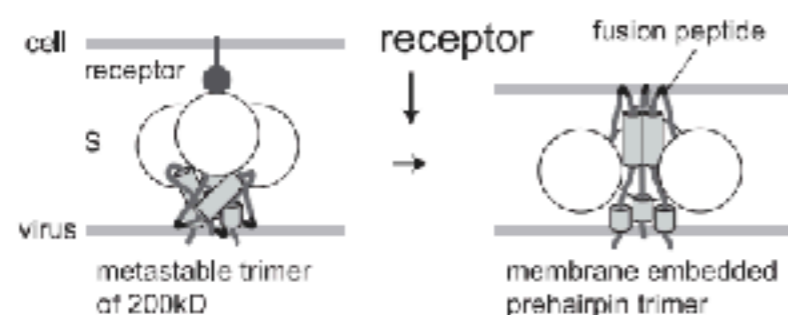
JOURNAL OF VIROLOGY, Nov. 2009, p. 11133-11141
0022-538X/09/\$12.00 doi:10.1128/JVI.00959-09
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Vol. 83, No. 21

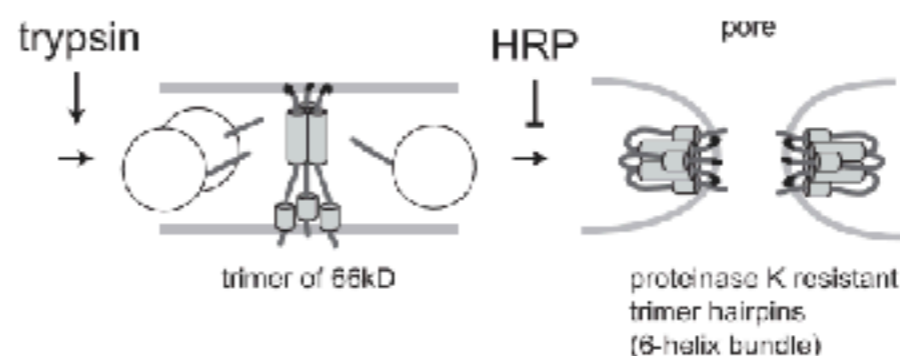
Two-Step Conformational Changes in a Coronavirus Envelope Glycoprotein Mediated by Receptor Binding and Proteolysis[∇]

Shutoku Matsuyama* and Fumihiko Taguchi†

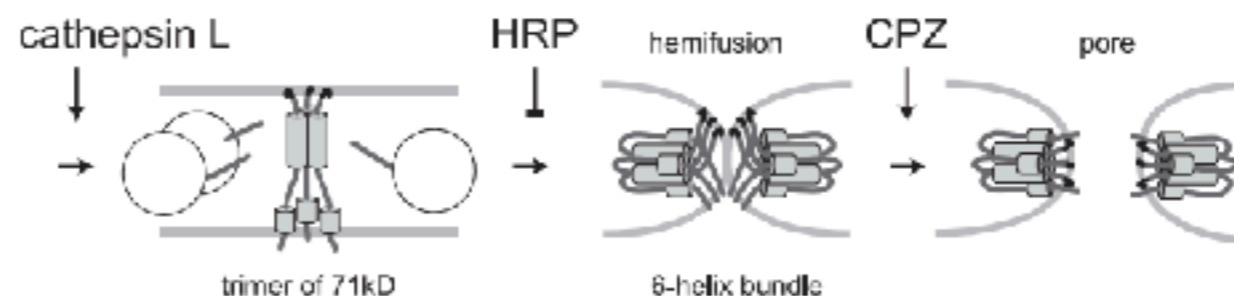
Step 1



Step 2 triggered by trypsin



Step 2 triggered by cathepsin L



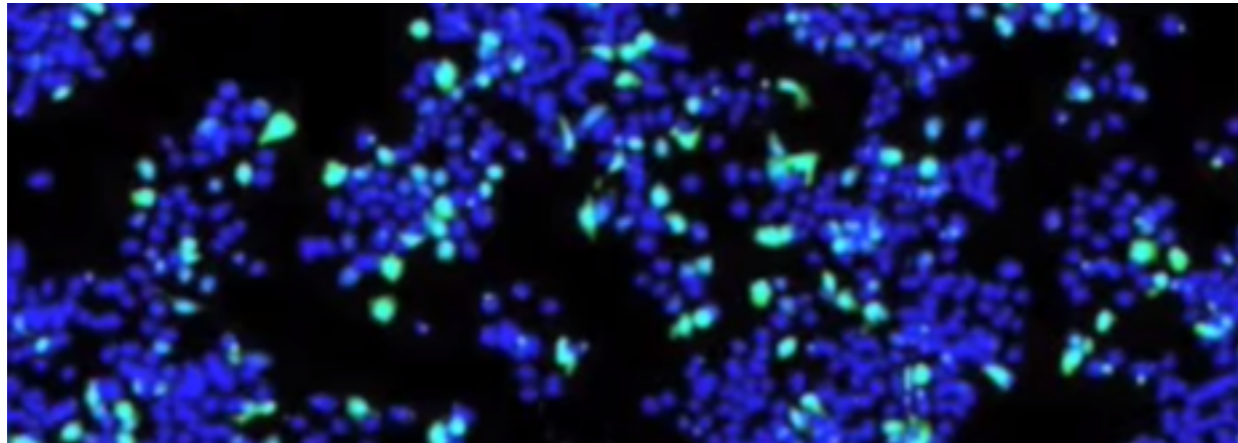
*serine protease TMPRSS2
or cathepsin CTSL*

Current State-of-the-Art for Understanding Systems Biology in Virology

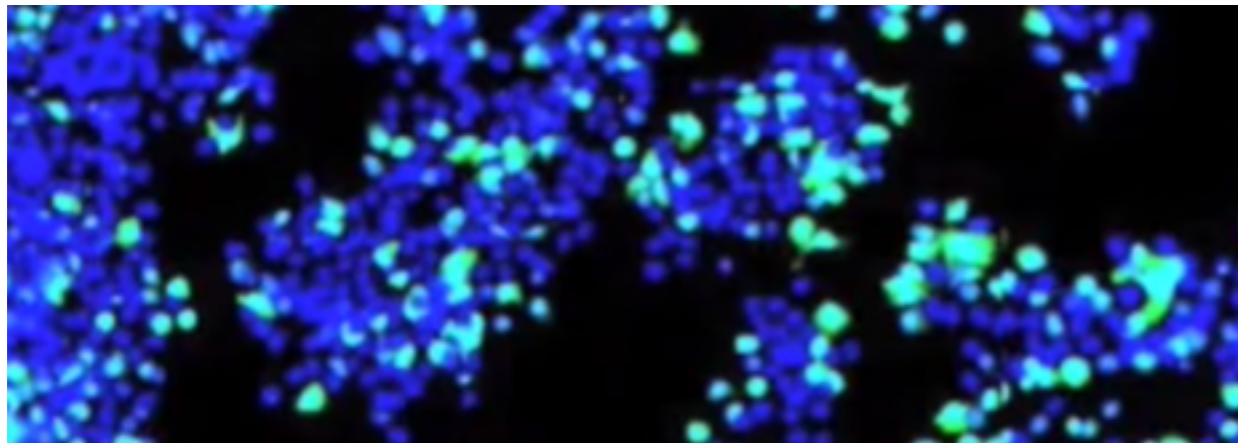
centered on siRNA silencing & high throughput experimentation

- ◆ 7000 druggable genes
- ◆ HTE using 384 well plates
- ◆ 3 siRNA per gene (testing in triplicate)
- ◆ mature viruses (MV) expressing GFP (green fluorescent protein)
- ◆ automated microscopy (hits contain no green)
- ◆ Hit = 2/3 or 3/3 siRNA causing 50% or less infection compared to controls

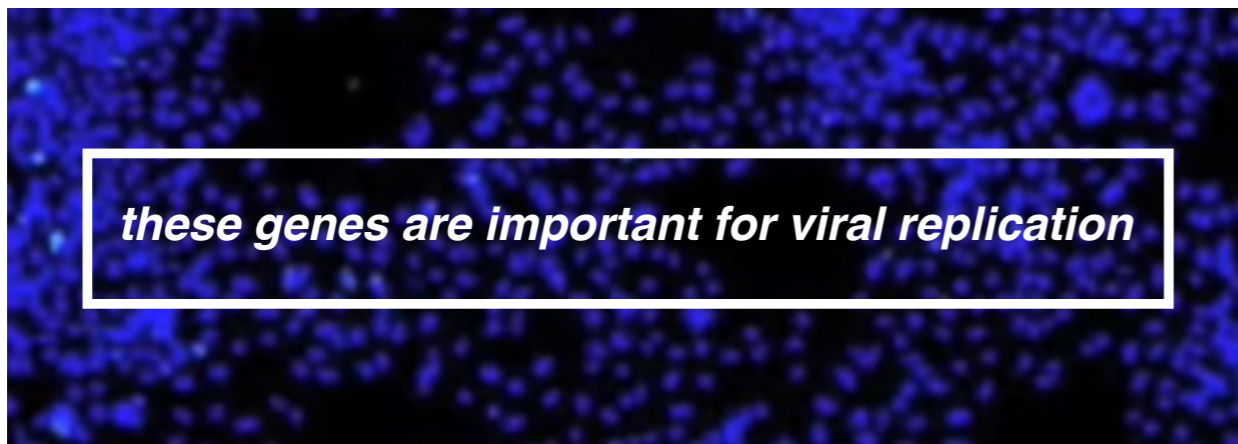
Current State-of-the-Art for Understanding Systems Biology in Virology



control
all normal human cell genes expressed



siRNA knockout of one gene
increased viral replication (rare)

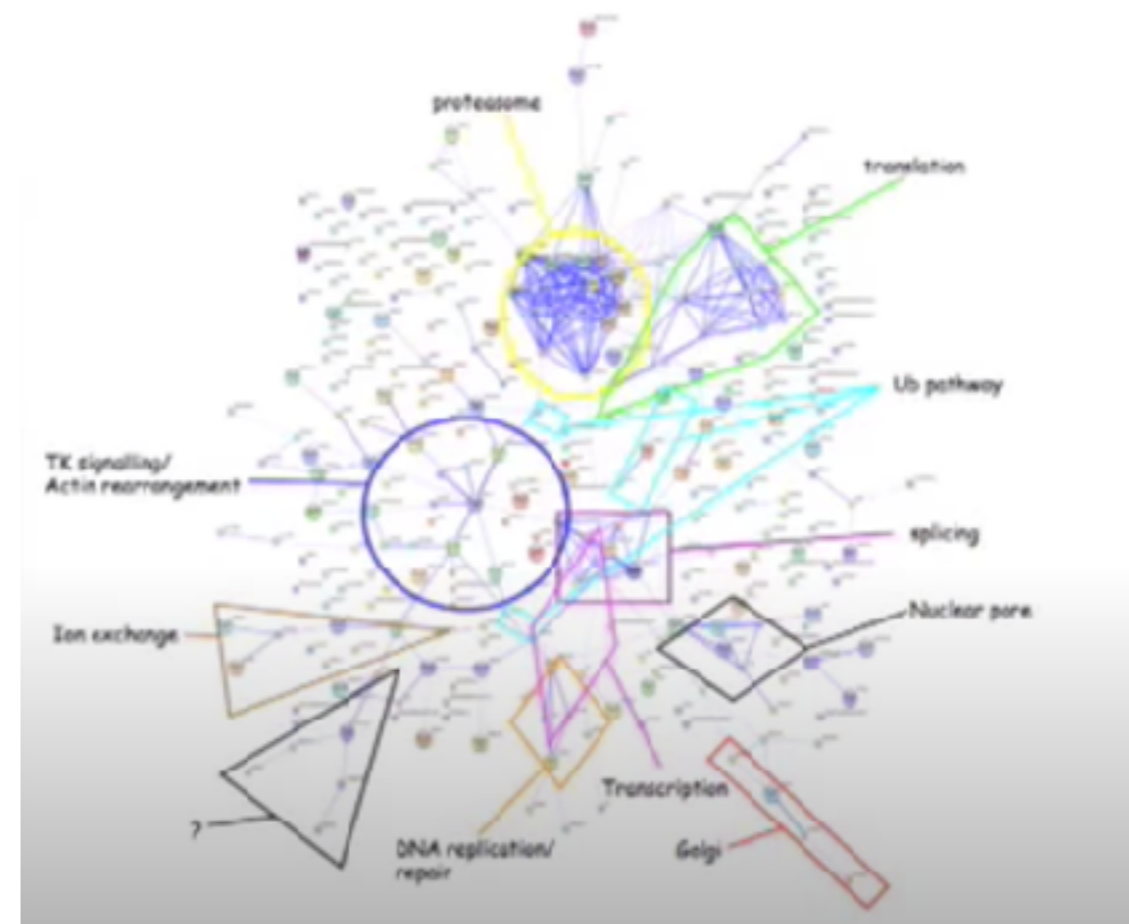
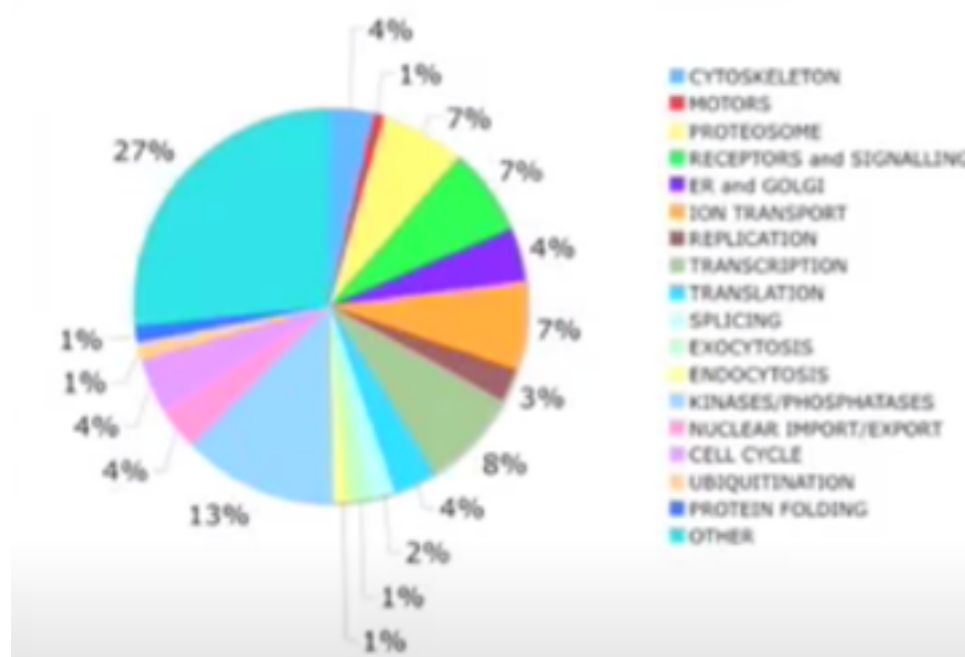
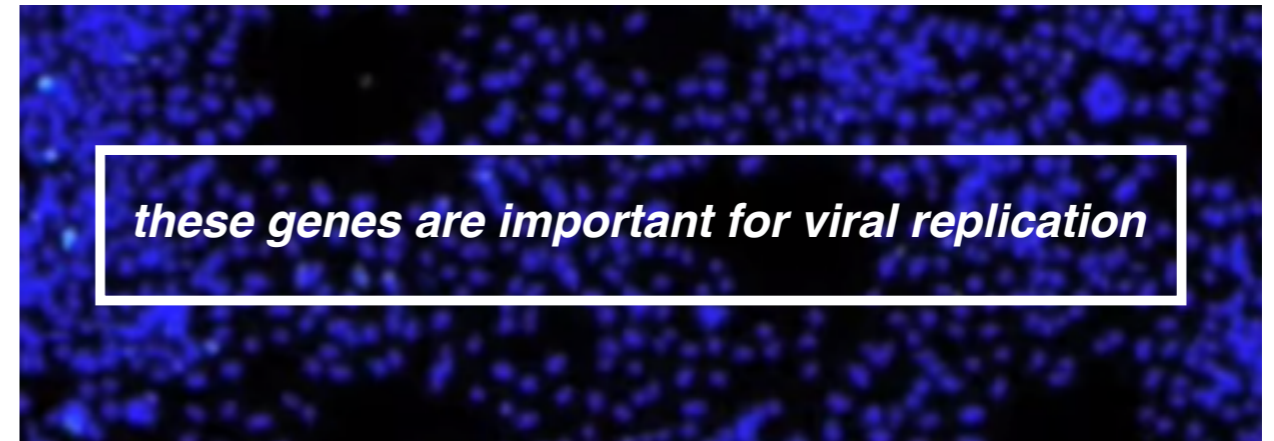


siRNA knockout of one gene
decrease in viral replication

Current State-of-the-Art for Understanding Systems Biology in Virology

results

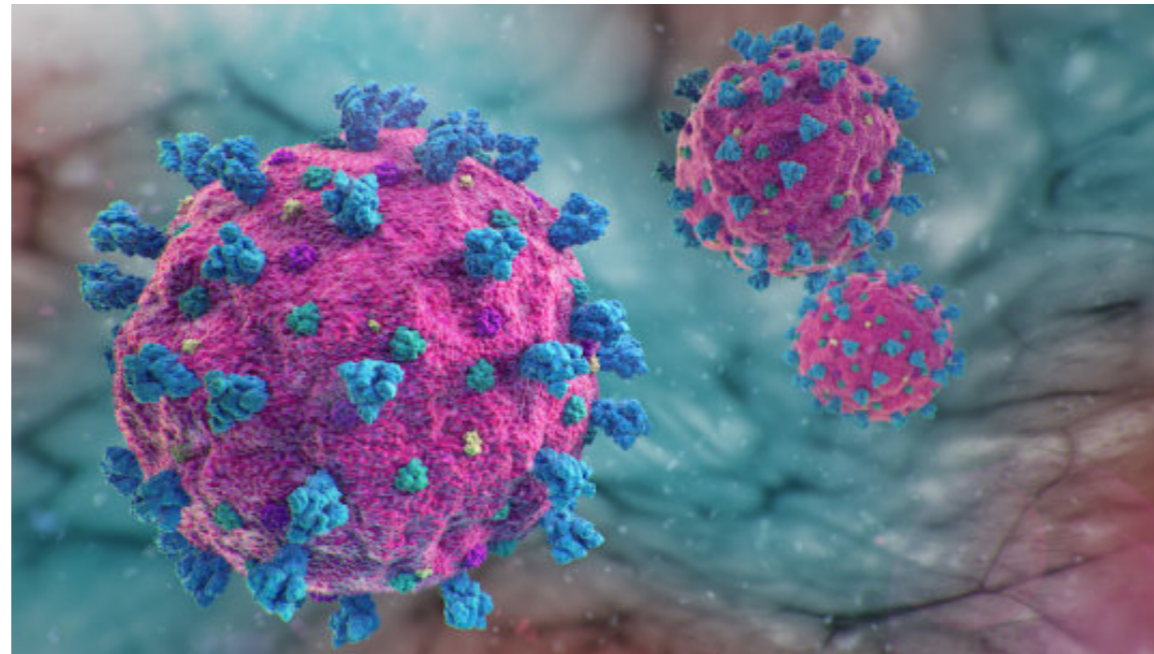
- ◆ 146 primary hits (3/3)
- ◆ 158 secondary hits (2/3)
- total of 304 inhibit infection**
- ◆ 4 increase infection
- ◆ string analysis of those genes



Additional Resources

- ◆ **iBiology (video lecture series on many biology topics)**
 - ◆ **Virus Entry (Ari Helenius, ETH Zurich)**
 - ◆ **Virus Ecology & Evolution (Paul Turner, Yale)**
 - ◆ **Danger from the Wild: HIV (David Baltimore, Caltech)**
 - ◆ **Discovering Reverse Transcriptase (David Baltimore, Caltech)**
 - ◆ **Studying Coronavirus (Tracey Goldstein & Koen Van Rompay, UCD)**
- ◆ **BioRender (“ChemDraw” for biomolecules, Princeton License)**

Viral Entry



*MacMillan Group Meeting
May 04, 2020*

Daniel Kim